

From Eye to Insight



# The 1<sup>st</sup> Cryo Electron Microscopy and 3D Image Processing of Macromolecular Assemblies and Cellular Tomography (CEM3DIP) 2016

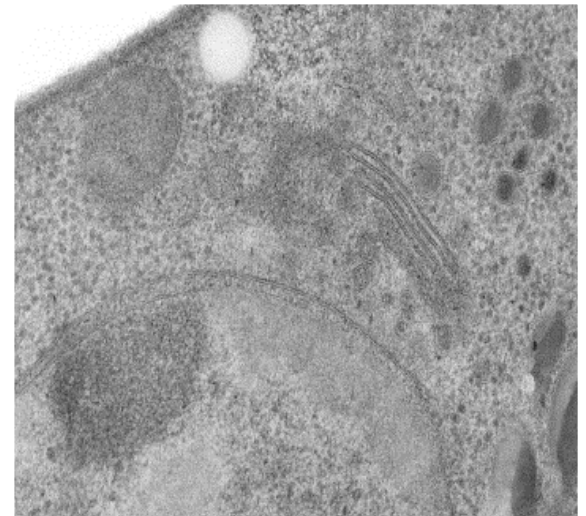
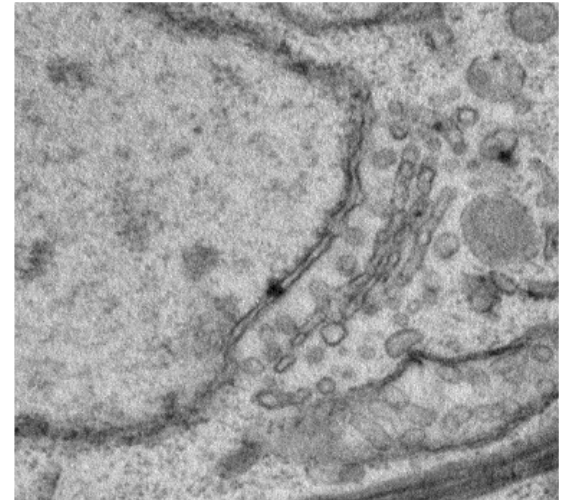
**EM CRYO PREPARATION WORKFLOWS**  
**Update on the Latest Instrumentation**

Science  Lab



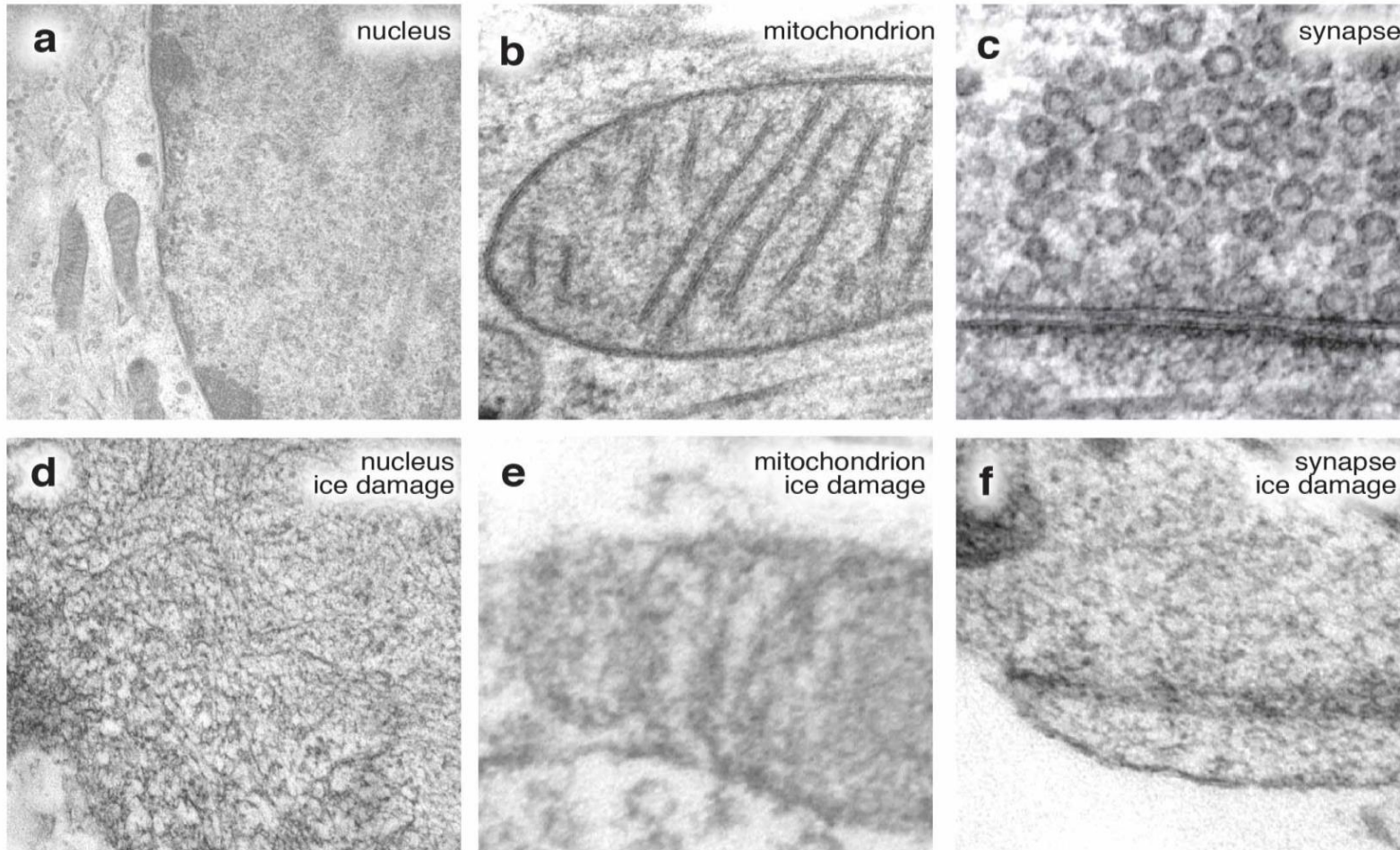
# Why Cryo?

- Low Contrast in EM
  - Critical Temperature  $-140^{\circ}\text{C}$
  - Sensitive to Electron Beam
  - Contamination (ice)
- 
- + Low Artefact Formation During Fixation
  - + Most Native Fixation and Analysis Method
  - + Fastest Fixation
  - + Frozen Hydrated Samples



*Toxoplasma gondii*

# The “No-Ice Challenge”



Watanabe S<sup>1</sup>, et al. Nature, 2013 Dec 12, 504 (7479):242-7 doi: 10.1038 nature 12809



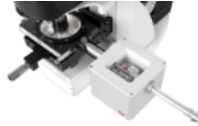
# Connectivity and Cryo-Preparation Workflows

2.



Cryo Fixation by  
High Pressure Freezing

1.



Cryo-CLEM

4.



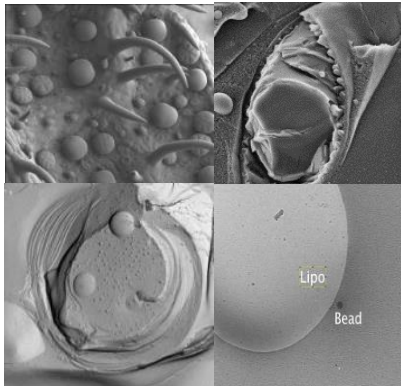
Cryo-Transfer

3.

Freeze Fracture, Freeze  
Etching and Coating



cryoEM



Cryo-Transfer



5.

Cryo Ion Milling



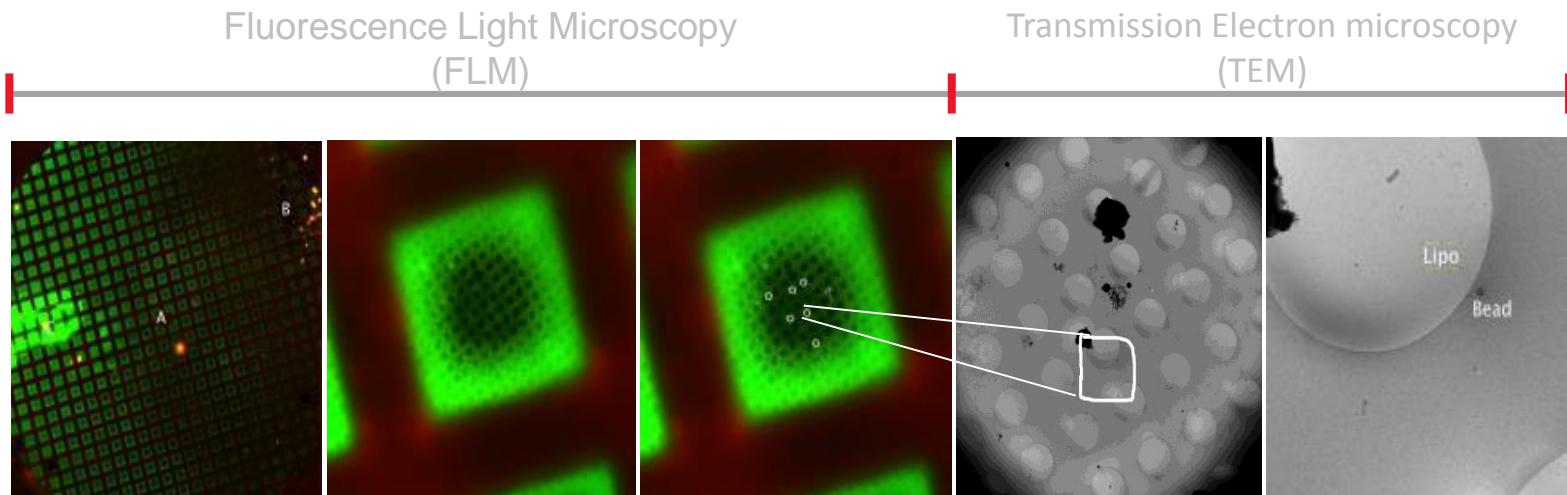
Cryo Ultramicrotomy



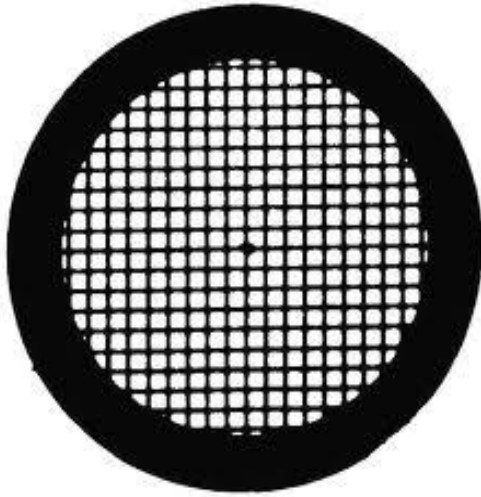


# 1. Correlative Light and Electron Microscopy

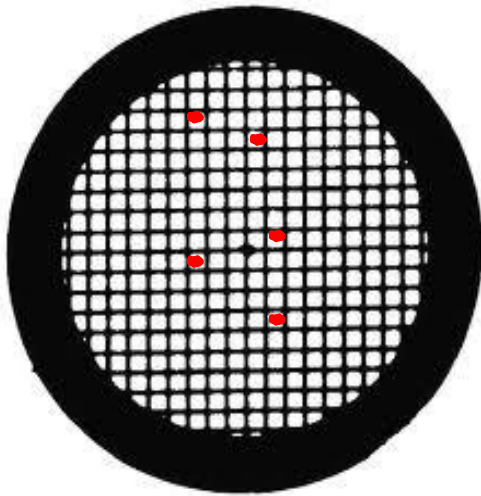
- Correlative Light and Electron Microscopy (CLEM) combines fluorescence light microscopy (FLM) and electron microscopy (EM) **imaging of the same sample**.
- A method which allows FLM **rapid screening** of large areas and fast determination of regions of interest in EM. This **reduces the user interaction time** on the EM significantly.
- Provides **deeper understanding** of analyzed sample by **overlaying complementary information** such as high resolution LM localization and EM ultrastructural context data.



# CLEM Application – Rapid Screening



Without FLM prior the EM the user might have to check all grid squares to find adequate target structures



With FLM prior the EM the user knows which grid squares contains promising target structures

# Cryo CLEM System



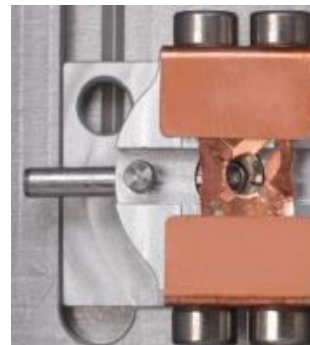


# Cryo CLEM System

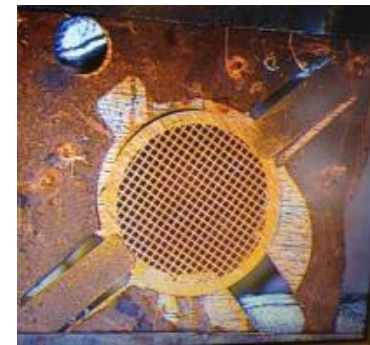
Plunge frozen  
or  
HPF/UM sample



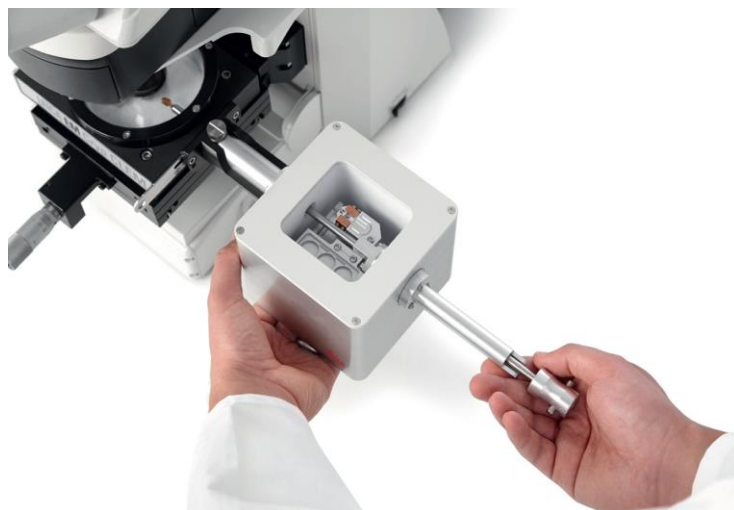
Cryo transfer system  
with cartridge loading station



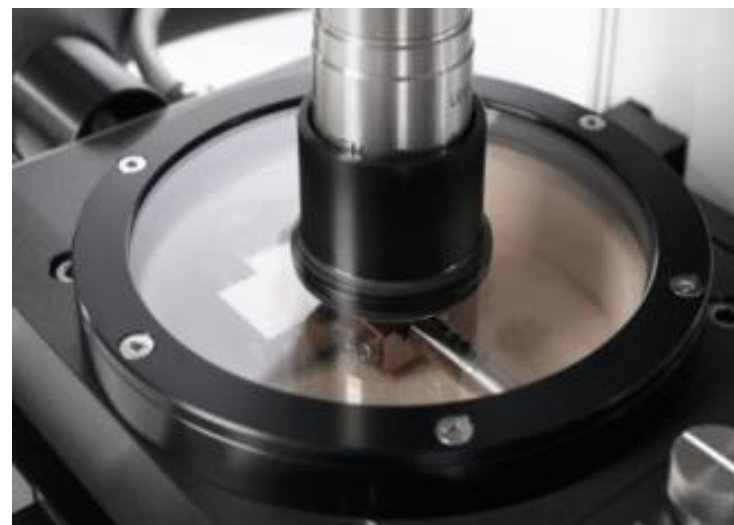
Cartridge loading station  
with empty cartridge



Cartridge with 3mm  
grid



Cryo transfer system docked to cryo stage



Leica cryo stage with cover and cryo  
CLEM objective.  
Temp. range  $-195^{\circ}\text{C}$  to  $+60^{\circ}\text{C}$

# Cryo CLEM System - Objective



Leica HCX PL APO 50x / 0,90 CLEM objective

- Achromatically corrected
- Thermally decoupled lens
- NA 0.9
- Low working distance of 0.28 mm
- Localization of ~50nm

# Workflow Solution

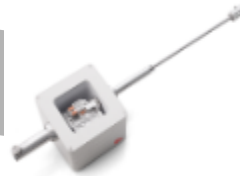


High Pressure Freezing,  
Cryo-Ultramicrotomy

or



Grid Plunging



Cryo-Transfer  
System

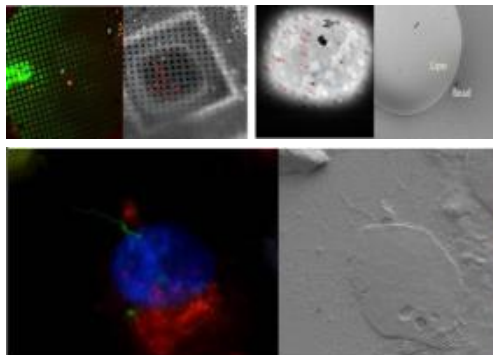


Leica Cryo Light Microscopy

**Cryo CLEM**

Leica LAS X Widefield Images used for  
correlation of LM marked structures in EM

## Image Analysis



**Cryo-TEM**



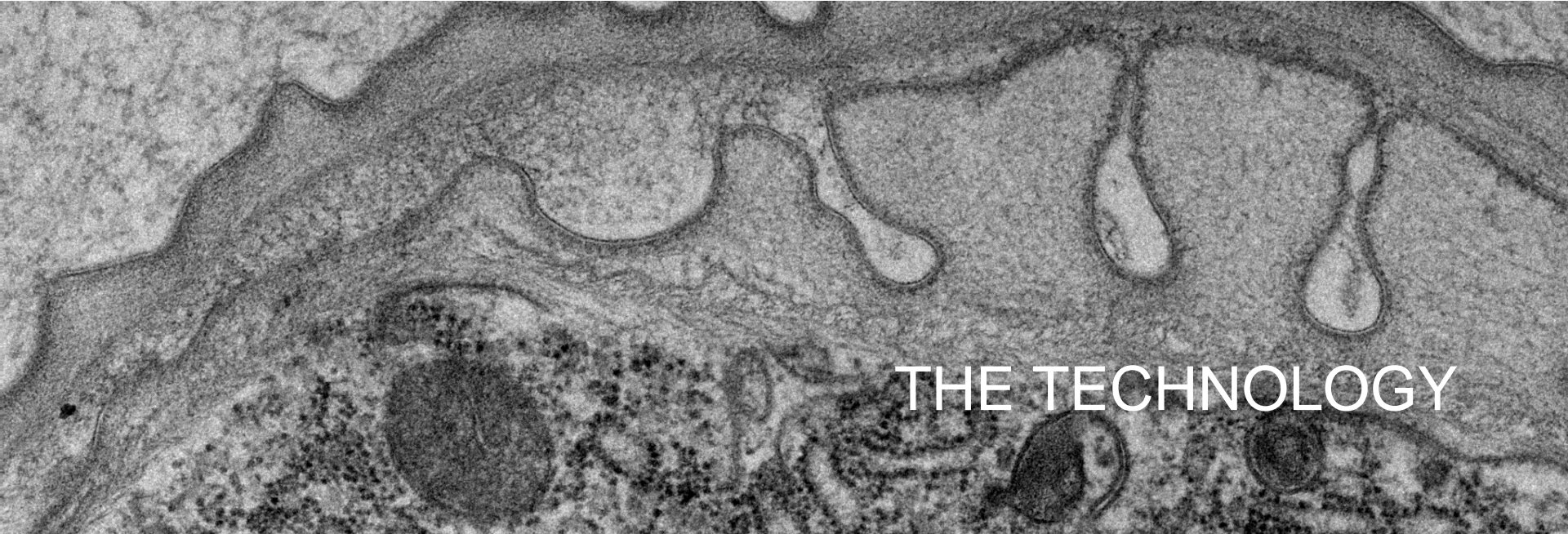
Transfer to cryoTEM





## A NEW ERA IN HIGH PRESSURE FREEZING





## THE TECHNOLOGY

C. elegans, courtesy of Elly van Donselaar, Martin Harterink and Karin Vocking, Utrecht University,

A High Pressure Freezer arrests aqueous samples in their native state to deliver the best possible sample preservation.

# LEICA EM ICE



Leica EM ICE is a platform for game-changing discoveries.



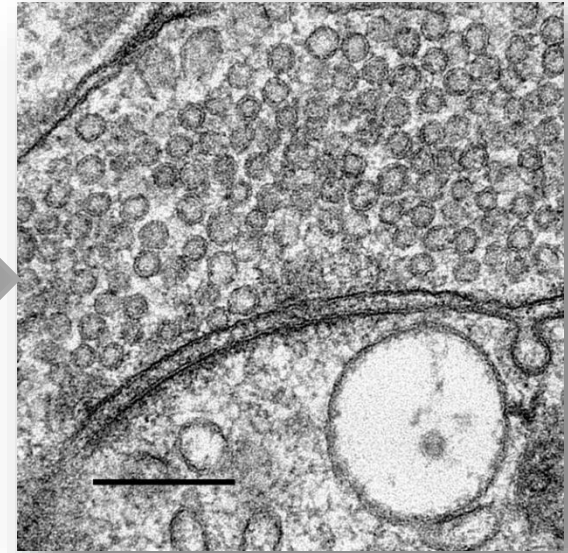
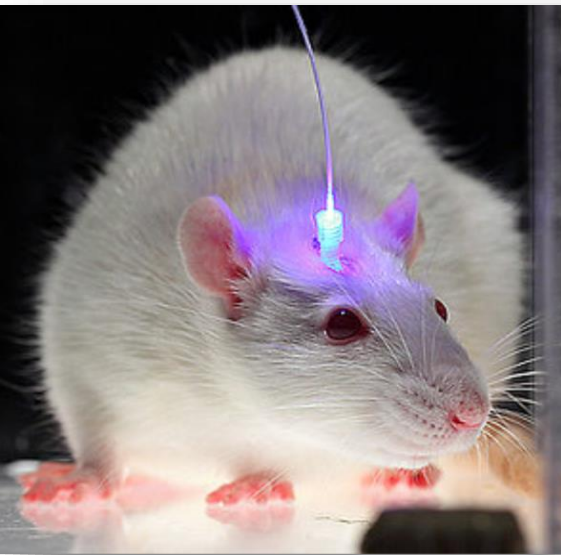
# LEICA EM ICE LIGHT STIMULATION

New possibilities for researchers in life science and industry

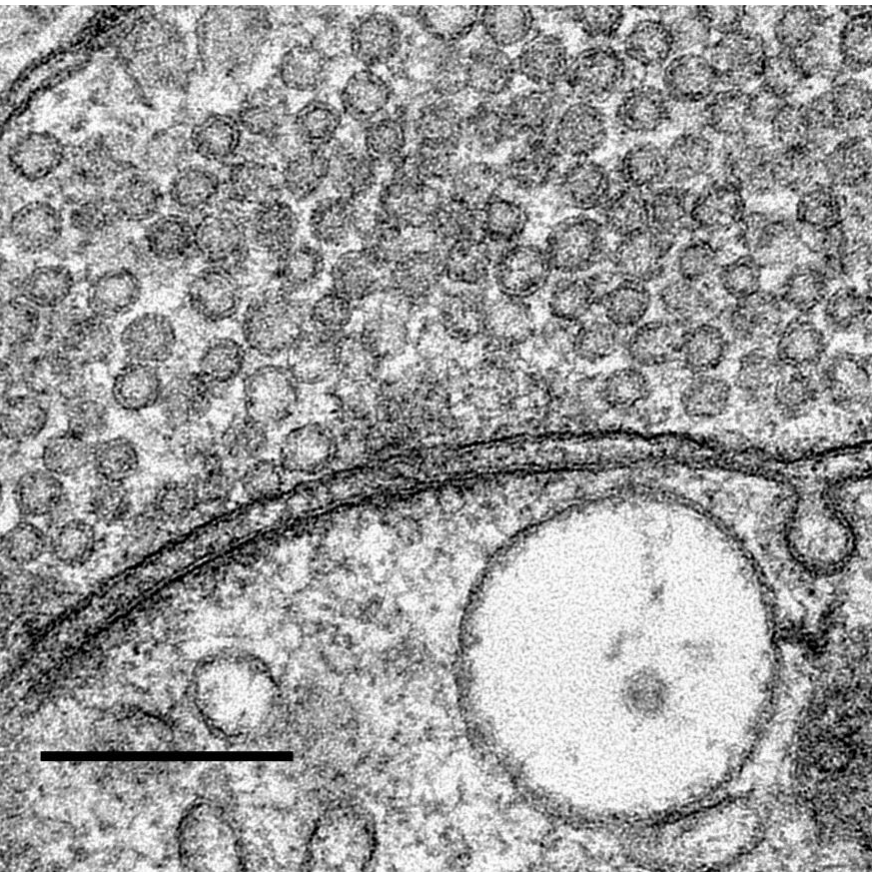


# WHY LIGHT STIMULATION?

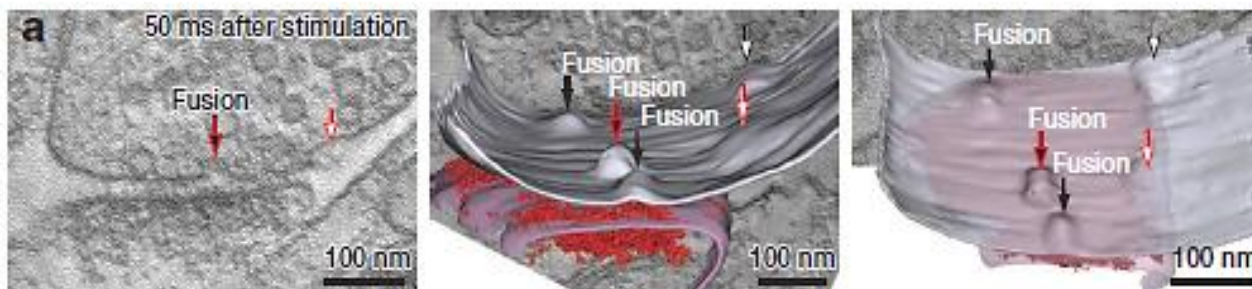
Utilizing light to understand the complex process of neurotransmission





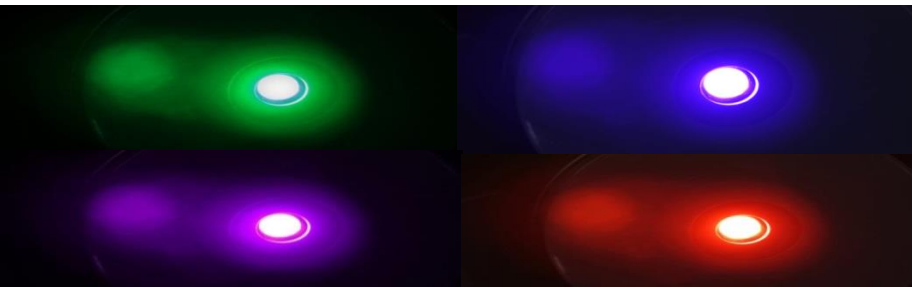


The synchronization of light stimulation and high pressure freezing allows the visualization of highly dynamic process at a nanometer resolution and millisecond precision.



**Nature. 2013 Dec 12;  
504(7479):242-7**





**Leica EM ICE**

**Temperature**  
Chamber: 26 °C  
Table: 25 °C

**Pressure**  
System: 9.6 bar

**LN<sub>2</sub>**

**Status**  
**System ready!**

close cover to start freezing

**Specimen Storage**

**Light Stimulation**

<program name> Edit

1 2 3 4 5

LED	Duration	Period	Pulse	Dark phase
amber 590 nm	1000ms	1000ms	100ms	50ms

Activate light stimulation

Container 1 0 / 1

Settings Bake out View Light 09:49 2015-04-10

## Variety of Light Spectra

- Five modules, five wavelengths automatically recognized by the instrument software

## Program Your Experiment

- Precise correlation between the light impulse and the time of freezing

- One minute recovery time between freezing cycles
- One second from fresh-to-frozen
- Only 20 minutes cool-down time
- Only 30 L daily consumption of LN2

**LEICA EM ICE**





The image shows a close-up of a Leica automated sample loading system. A red L-shaped metal frame is positioned over a black base. A white rectangular block is on the left. A silver cylindrical component is being moved into a slot in the red frame. To the right, several small, colorful (green, orange, white) plastic components are scattered on a black surface. The text 'FOCUS ON YOUR SAMPLE' is overlaid in white, with 'One move, fully automated loading' below it. At the bottom, red text describes the process: 'Closing the cover triggers the perfect alignment of cartridge assembly followed by the freezing process'.

# FOCUS ON YOUR SAMPLE

One move, fully automated loading

Closing the cover triggers the perfect alignment of cartridge assembly followed by the freezing process



## SAMPLE STORAGE DEWAR

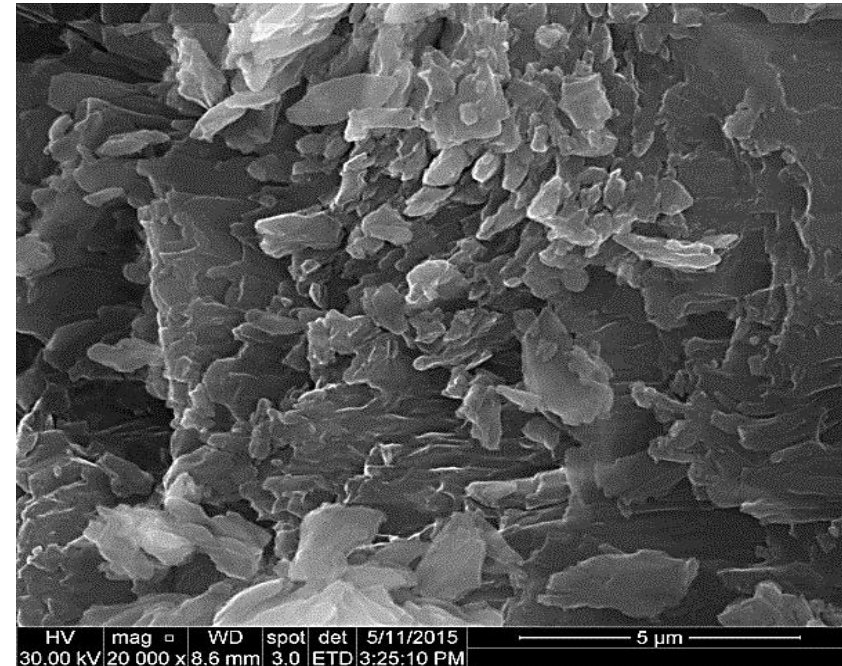
- Three separate positions
- Nine consecutive freezing cycles
- Programmed rotation



# DISCOVER NEW POSSIBILITIES



Apply light stimulation to  
any light sensitive compounds



Sunscreen lotion frozen after millisecond UV  
light stimulation.

### 3. Cryo-preparation – Freeze Fracture

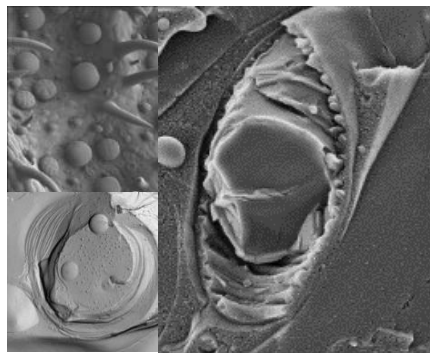


Cryo Fixation  
High Pressure Freezing



Cryo-Transfer

Cryo SEM



Cryo-Transfer



Freeze Fracture, Freeze  
Etching and Coating

# A REVOLUTION IN FREEZE FRACTURE



# READY TO OPERATE

- Load locks for sample, microtome, e-beams
- New cooling, shielding, microtome, e-beams
- High vacuum 10<sup>-8</sup>mbar
- Connection to VCT500





# PRECISE AND CLEAN CUTTING

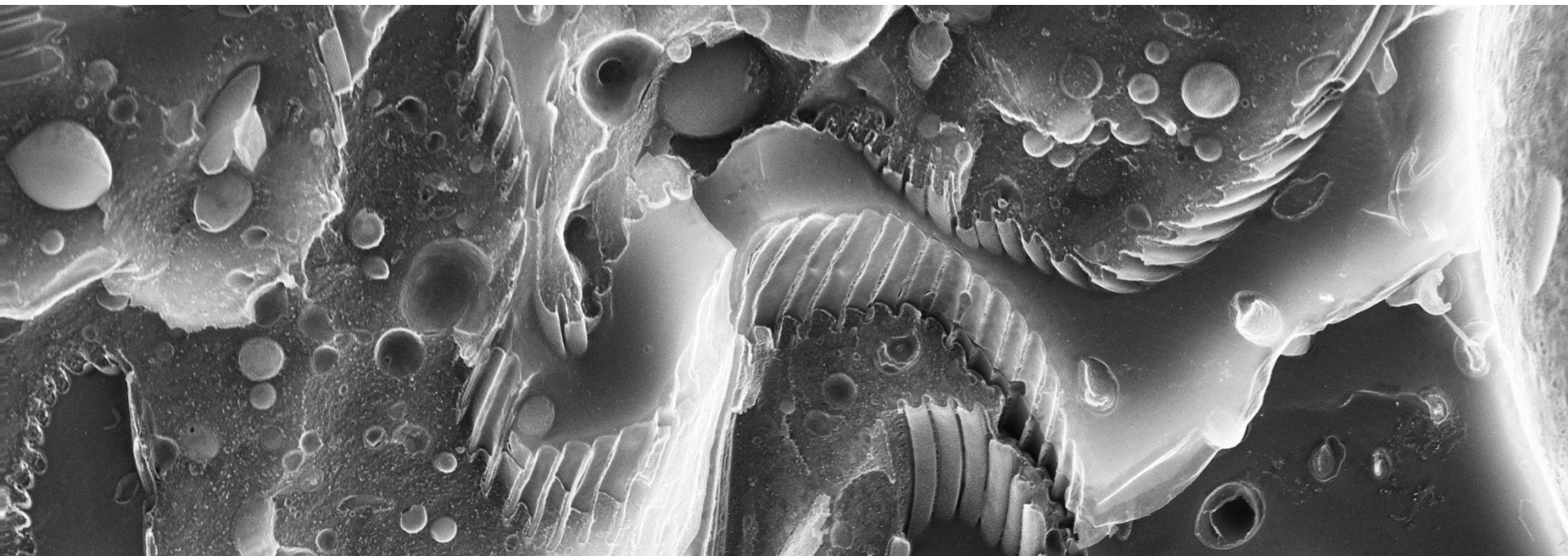
Exchangeable and 3 axis movable knife



# FREEZE FRACTURE

Freeze Fracture breaks frozen specimens to reveal internal structures

*Euglina gracilis*, courtesy of Andres Käch, Center for Microscopy and Image Analysis, University of Zurich, Switzerland

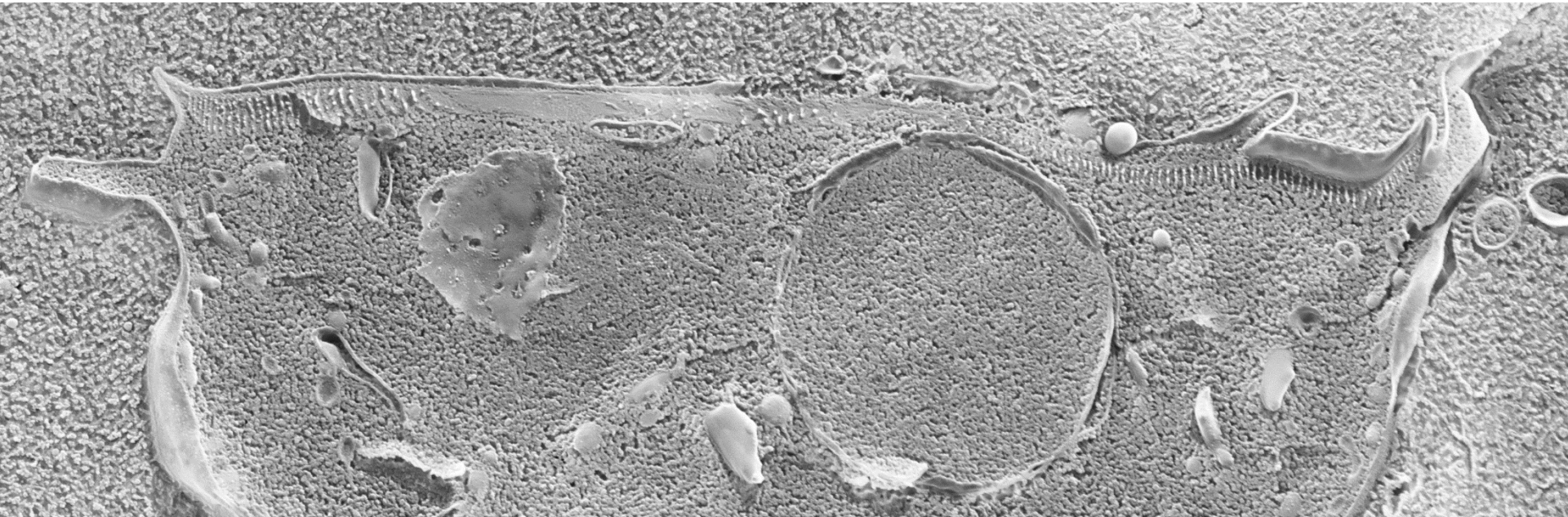




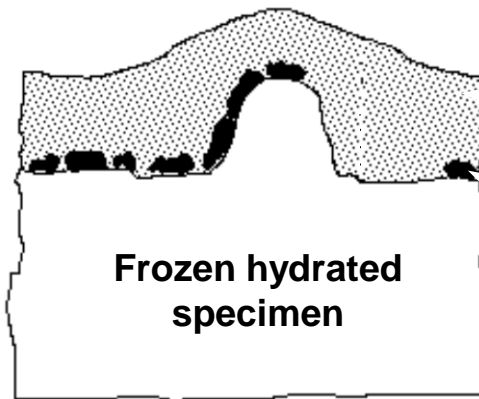
# FREEZE ETCHING

Freeze Etching is the sublimation of surface ice under vacuum to reveal details of the fractured face that were originally hidden.

*Giardia lamblia*, courtesy of Andres Käch, Center for Microscopy and Image Analysis, University of Zurich, Switzerland



# FREEZE FRACTURE & REPLICAS



Carbon

Platinum



Replica



**cryo-SEM**

Backscattered signal



**TEM**

Amplitude contrast



## 4. THE FUTURE IS CONNECTIVITY

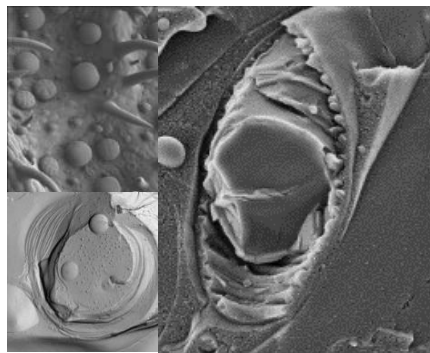


Cryo Fixation  
High Pressure Freezing



Cryo-Transfer

Cryo SEM



Cryo-Transfer

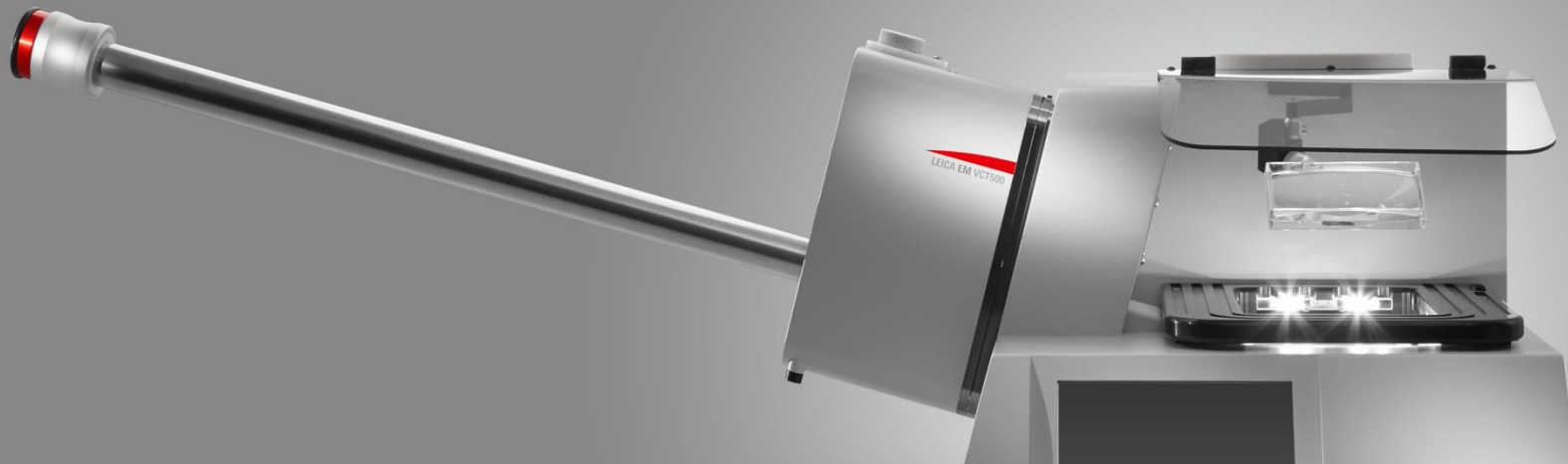
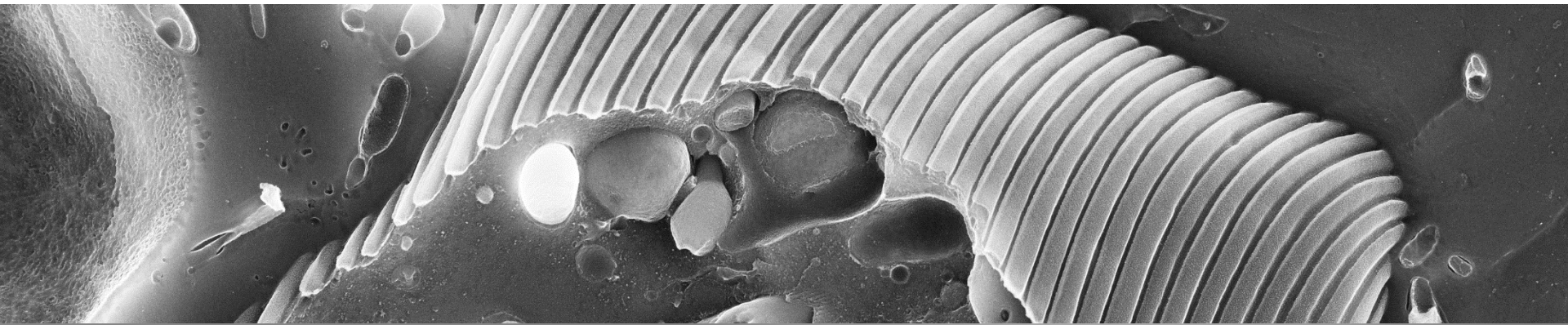


Freeze Fracture, Freeze  
Etching and Coating

# WHY VACUUM CRYO TRANSFER?

Contamination-free sample transfer between preparation and analysis systems.

*Euglena gracilis*, courtesy of Andres Käch, Center for Microscopy and Image Analysis, University of Zürich, Switzerland





## CONNECT WITH LEICA EM VCM

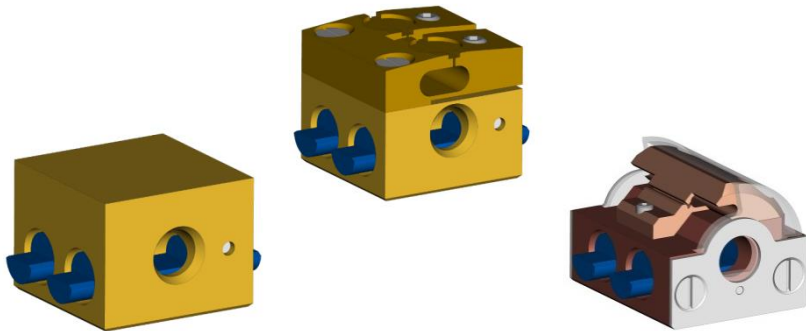
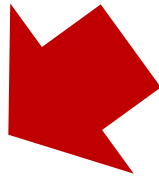


- All transfer under vacuum
- Contamination-free sample handling
- Improved connectivity



# FLEXIBILITY TO CONNECT

- Glovebox
- SEM
- FIB
- Analysis systems, e.g. XPS
- Atomic probe
- Synchrotron
- ....

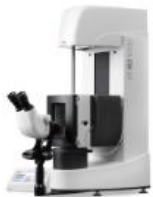




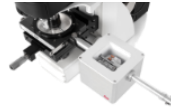
# Connectivity for cryo-preparation



Cryo Fixation, e.g.  
High Pressure Freezing



Cryo-Transfer



Cryo-CLEM



Freeze Fracture, Freeze  
Etching and Coating



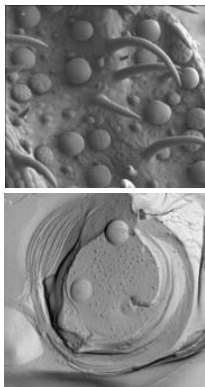
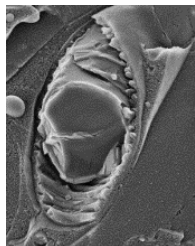
Cryo Ultramicrotomy



**5.** Cryo Ion Milling

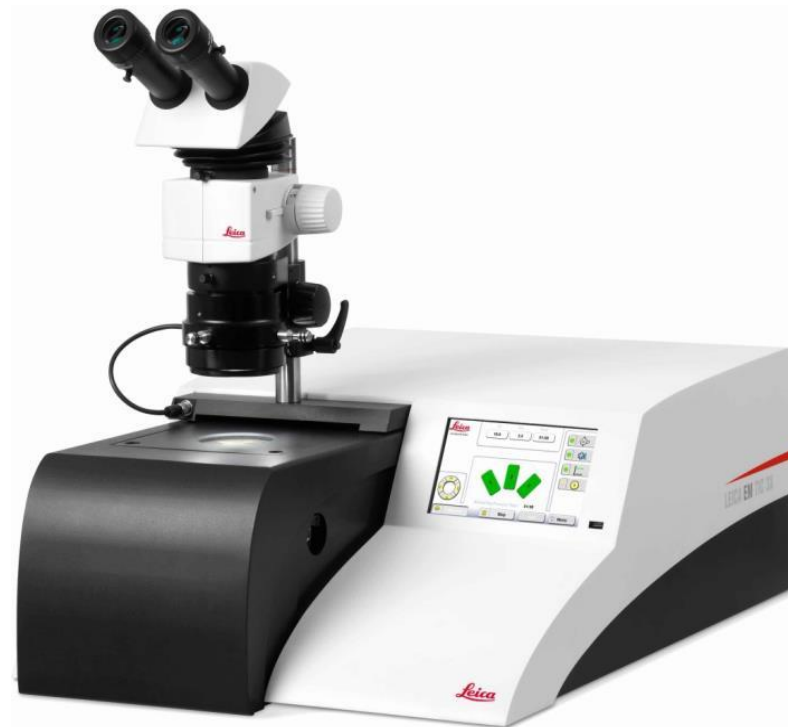


Cryo-Transfer



## 5. Broad Ion Beam Milling Workflow

Sample preparation under controlled environmental conditions

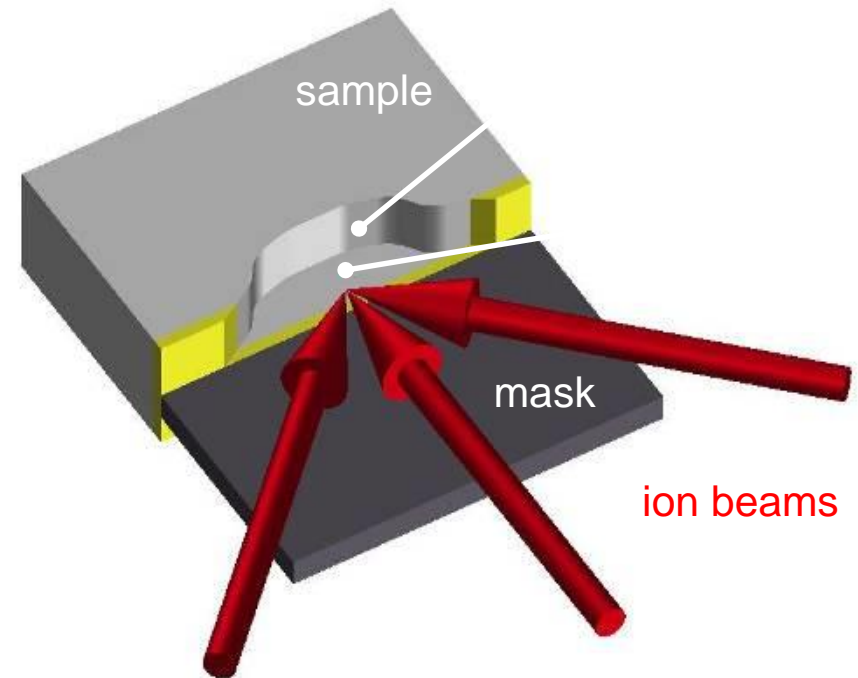


## Ion beam slope cutting with Leica EM TIC 3X

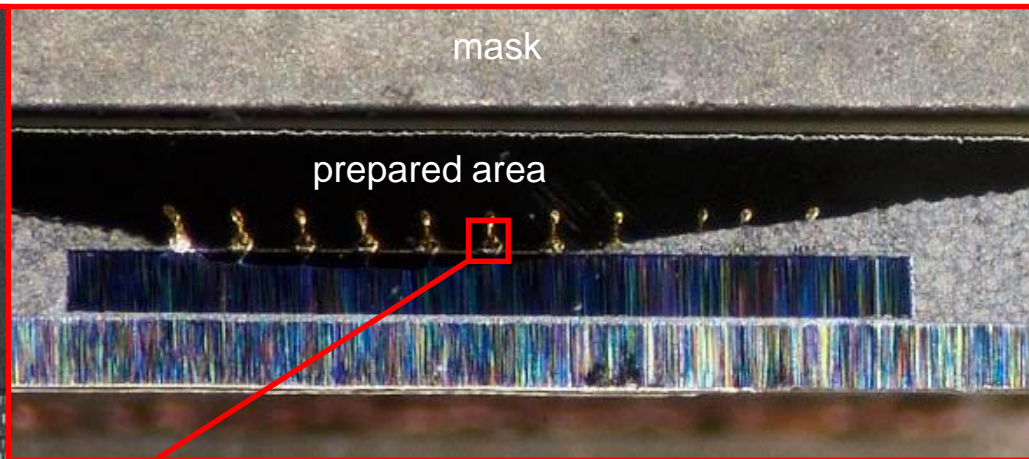
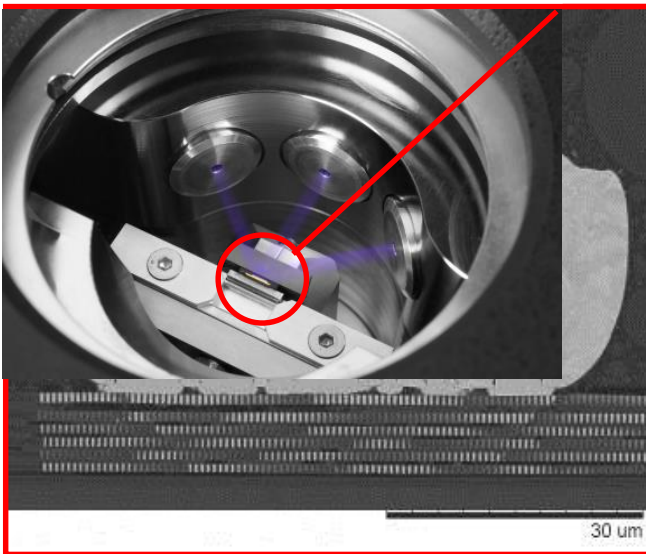
- Three ion beams hitting the sample from different directions (reduction of curtaining)
- Fixed sample (better heat transfer)

### Features

- Cutting depth  $>1000\mu\text{m}$
- Cutting width  $>4000\mu\text{m}$
- Cutting speed  $>150\mu\text{m/h}$



## Leica EM TIC 3X “Triple ion beam” slope cutting





# What About Temperature, or Environmentally Sensitive Samples?

## Heat-sensitive polymer fibres with water-soluble portion



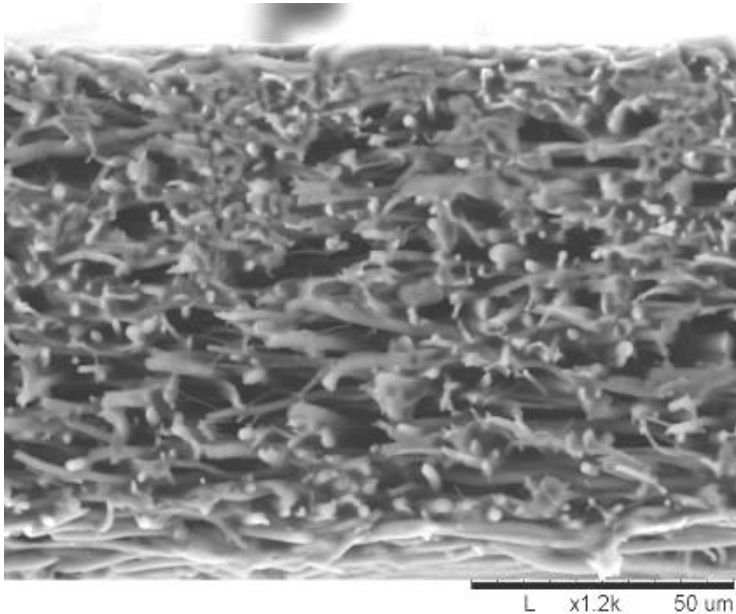
Without cooling ( $\sim 80^{\circ}\text{C}$ )



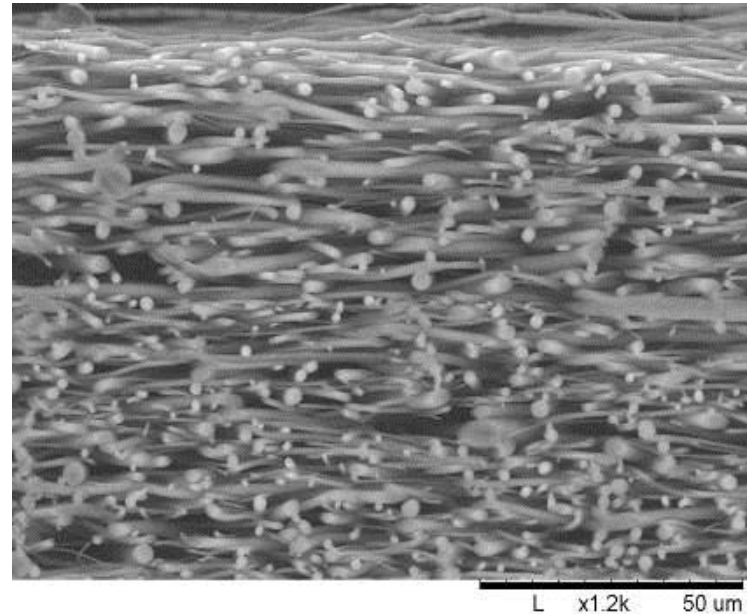
With cooling stage ( $-120^{\circ}\text{C}$ )

# Comparison

Heat-sensitive polymer fibres with water-soluble portion

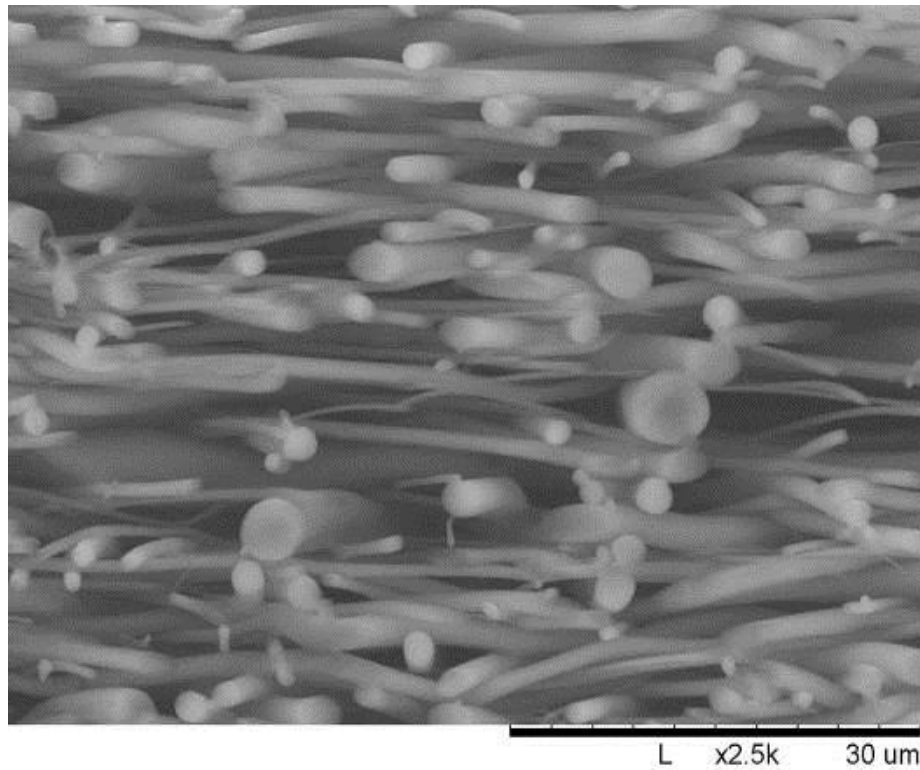


UC7 with FC7 -140° C



TIC 3X -120° C

# Cooling stage result





# Environmentally controlled workflow

Sample  
Freezing



**Cryo-loading station** with **cryo-saw**  
LN2 sample pre-preparation



**EM TIC 3X** with cryostage &  
VCT docking station  
(cryo-) BIB milling

(Cryo-)  
SEM  
FIB



**VCT 100 shuttle**  
(cryo-) vacuum transfer

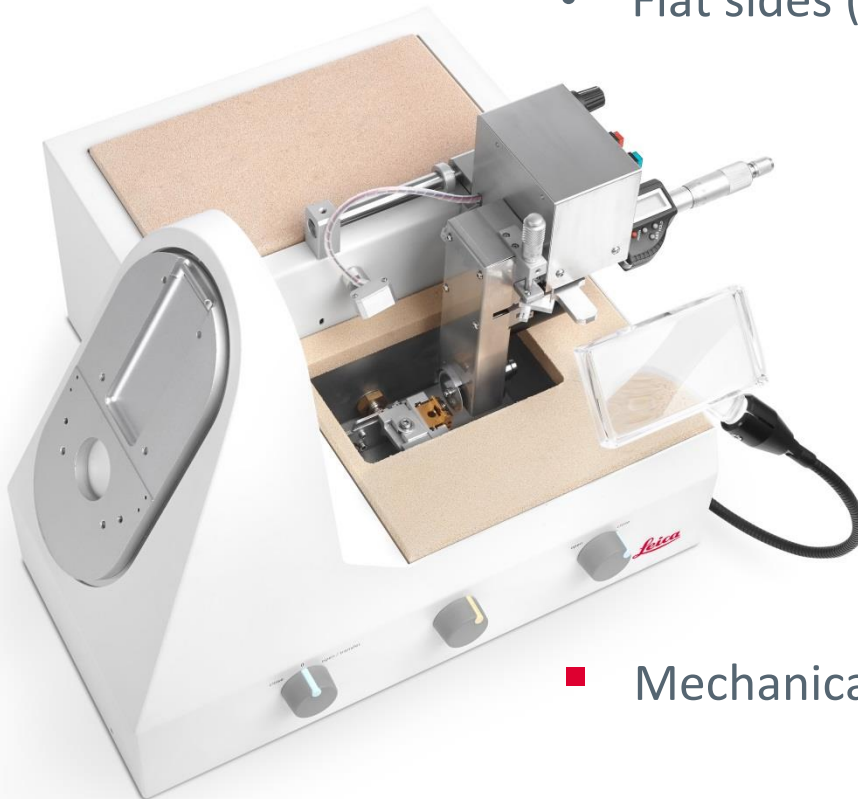


**EM ACE 600** with VCT docking station  
(cryo-) coating

# Cryo-saw and loading station

Sample size:

- Maximum 10x7x4(thick)mm
- Flat sides (saw-finish

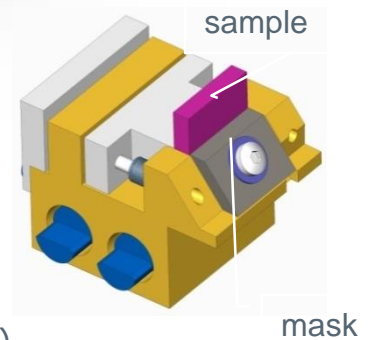


- Mechanical preparation under LN2 conditions

# Preparation Process

- Freeze
- transfer to cryo-saw preparation station filled with LN2
- inserted in the VCT holder and pre-prepared with cryo-saw

VCT holder  
(sample size max. 10x7x4mm)



# Leica EM VCT500 connectivity

Cryogenic sample transfer to and from the broad ion beam etching system, TIC 3X and to (cryo)SEM

- After shaping to size, sample is placed in loading position
- VCT shuttle is attached and the Dewar filled with LN2
- Sample taken on board VCT shuttle, transferred and attached to the EM TIC 3X docking port
- Sample moved into position for ion beam slope cutting under cryo-conditions.





# New environmentally controlled workflow



Sample  
Freezing



**Cryo-loading station** with **cryo-saw**  
LN2 sample pre-preparation



**EM TIC 3X** with cryostage &  
VCT docking station  
(cryo-) BIB milling

**CryoSEM/FIB**

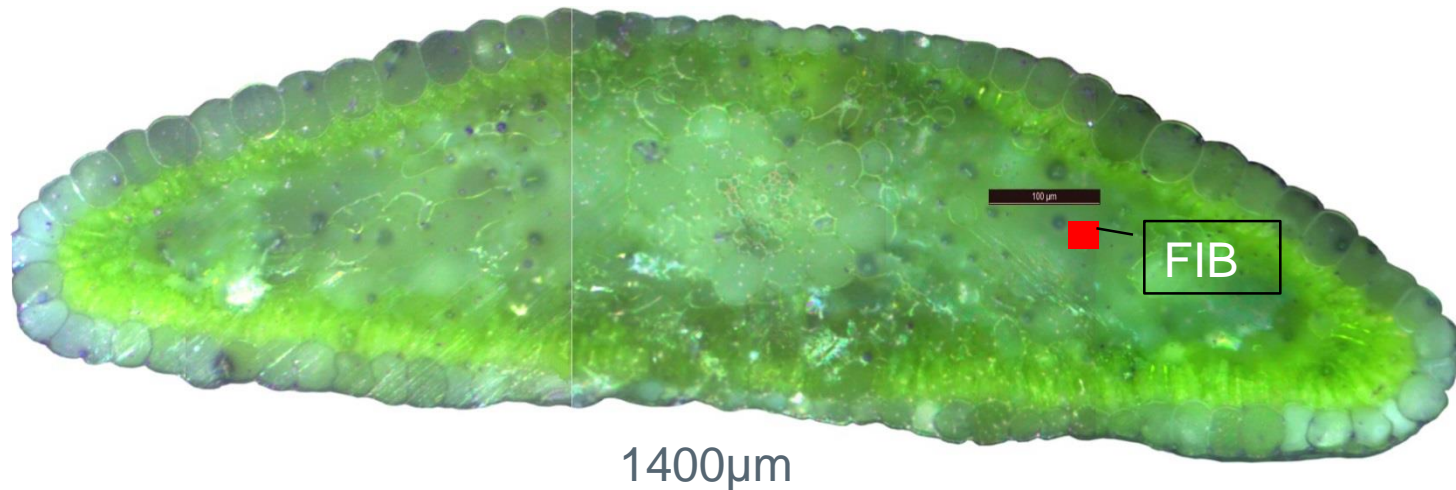


**VCT 100 shuttle**  
(cryo-) vacuum transfer



**EM ACE 600** with VCT docking station  
(cryo-) coating

# Applications

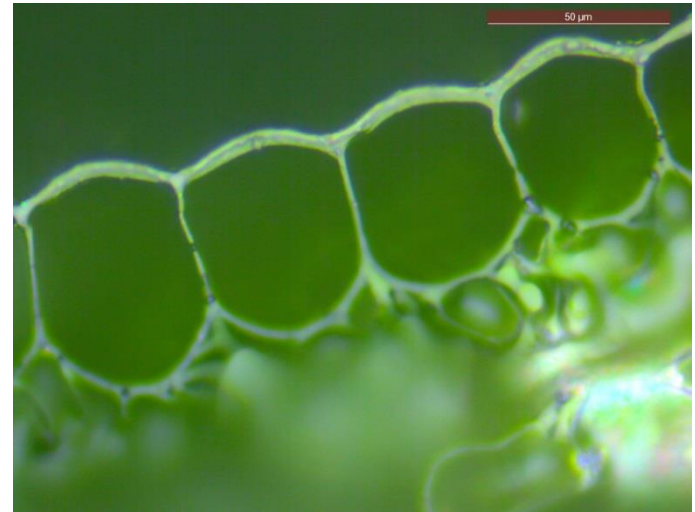


Asparagus; plunged in LN2, pre-prepared with the Cryo-Saw-Loading station, Ion beam cross sectioned with the TIC 3X – VCT at -120° C and 6kV; transferred with the VCT100 and investigated in a "cryo-LM" 20x objective

# Applications

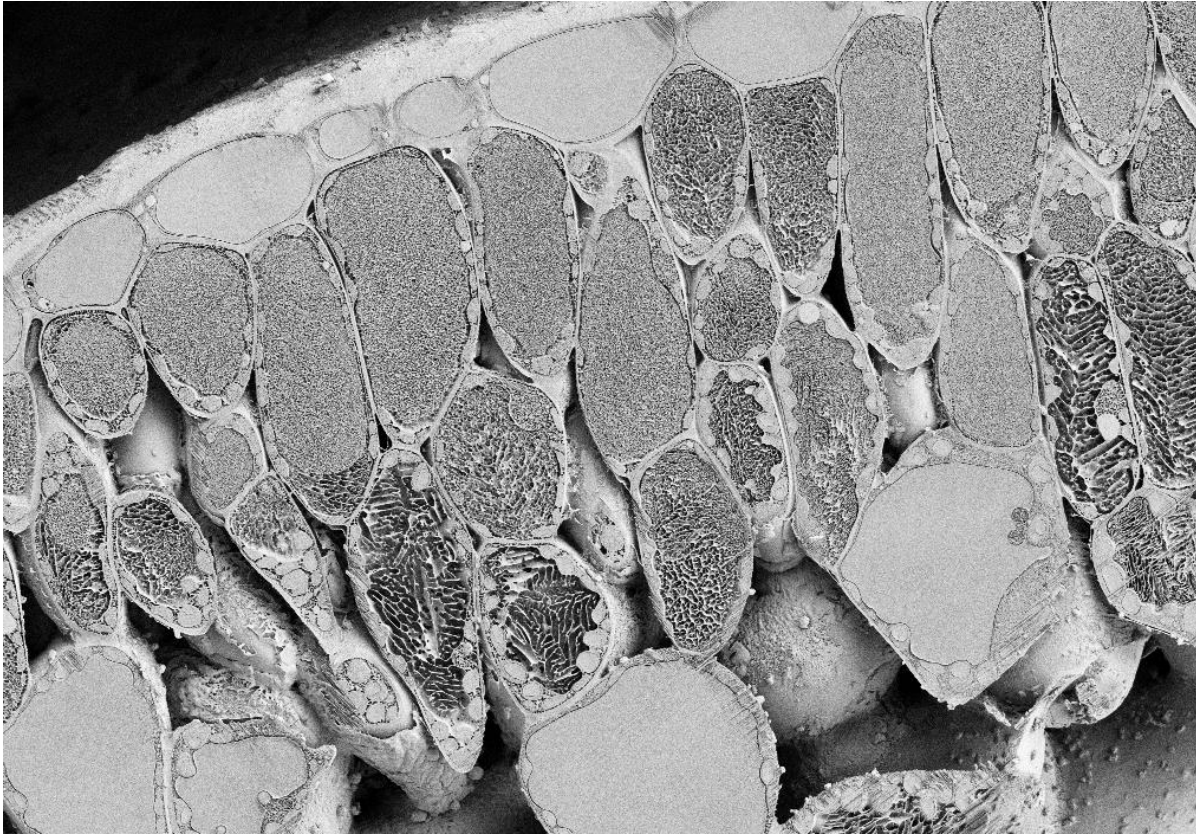


Asparagus; Cryo-TIC preparation; "Cryo-LM"



Asparagus; Cryo-TIC preparation; LM after the sample warmed up

# Applications



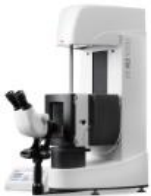
*Taxus* sp., plunged in LN2, pre-prepared with the Cryo-Saw-Loading station, ion beam cross sectioned with TIC 3X/VCT at -120C and 6kV; transferred with the VCT100 and investigated in a cryo-FIB SEM  
Image courtesy of Harvard University



# Connectivity for cryo-preparation



Cryo Fixation, e.g.  
High Pressure Freezing



Cryo-Transfer



Cryo-CLEM



Freeze Fracture, Freeze  
Etching and Coating



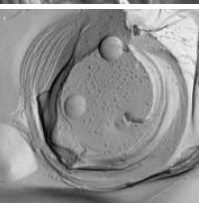
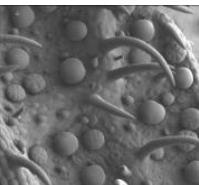
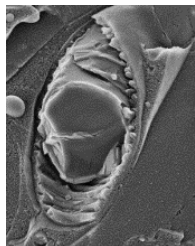
Cryo Ultramicrotomy



Cryo Ion Milling



Cryo-Transfer



# Future Developments

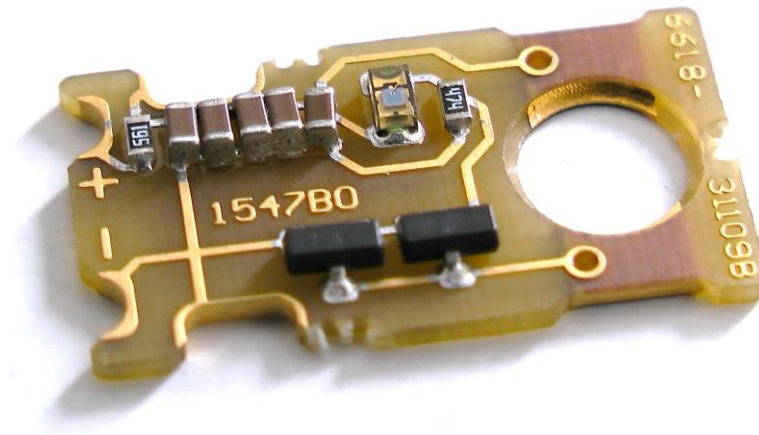
# HPF with Electrical Stimulation



# The Concept

- Middle plate includes a circuit board with capacitors to store the electrical energy

Middle plate with capacitors top-side view



Middle plate back-side view



Spacer ring



Cover ring



# The Concept

- Blue LED module used as the trigger for releasing the power and having e-current delivered to the sample
- Software automatically recognising the ES mode. All settings saved in the log-file

The screenshot displays the Leica EM ICE software interface, which is divided into several functional areas:

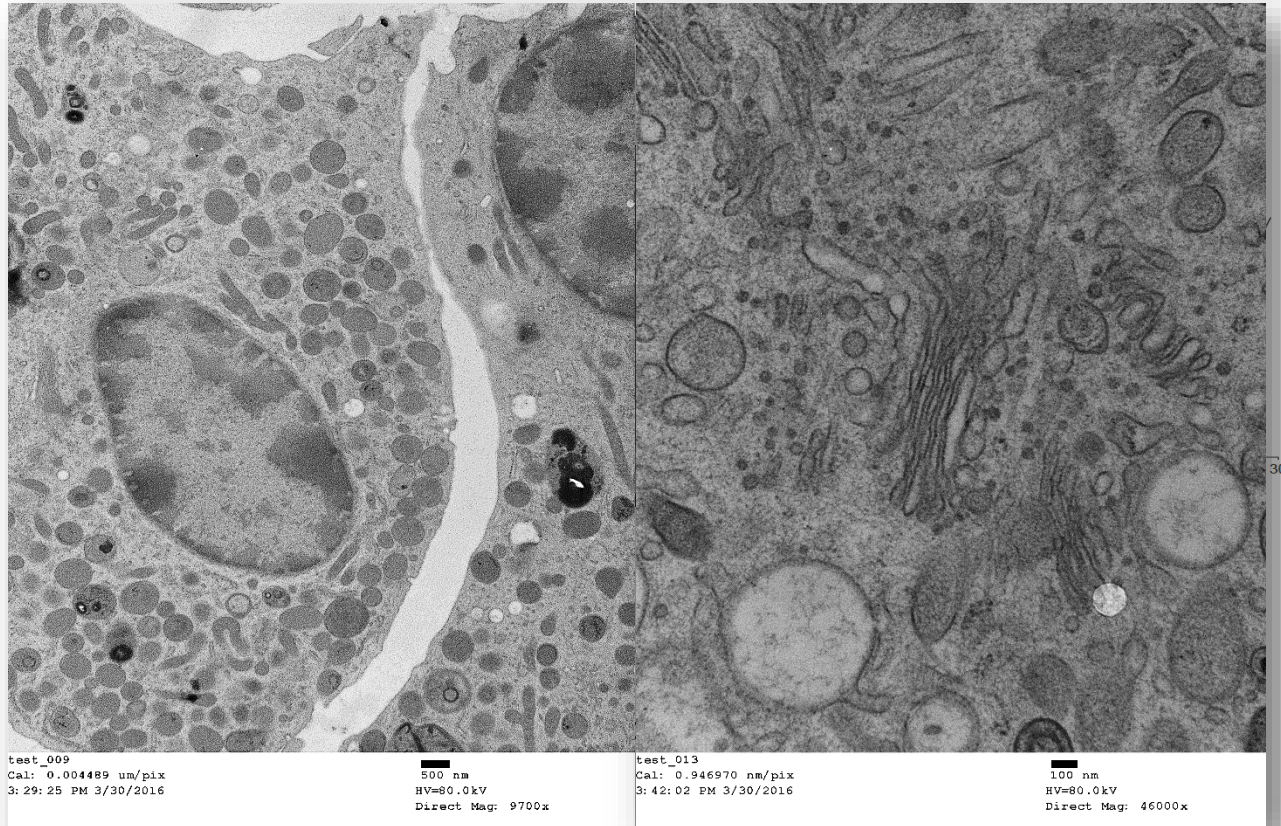
- Top Left:** Leica MICROSYSTEMS logo and the text "EM ICE".
- Temperature Section:** Labeled "Temperature", it shows "Chamber: 37 °C".
- Pressure Section:** Labeled "Pressure", it shows "System: 9.4 bar". Below this is a visual representation of an LN<sub>2</sub> cylinder with a "MIN" and "LOW" level indicator.
- Status Section:** A central area with a green "System ready!" message, an illustration of the specimen stage, and the instruction "Close cover to start freezing.".
- Electrical Stimulation Section:** Titled "Electrical Stimulation", it includes a "Specimen Storage" dropdown menu showing "<program name 1>" with an "Edit" button. Below are five numbered buttons (1-5), with button 1 selected. A table of parameters is shown:

	Dark phase	Period	Pulse	# Periods
LED	→	[Pulse]	[Pulse]	[Pulse]
	0ms	1000ms	900ms	1

Below the table, it specifies "blue 460 nm" and includes a "Check cartridge" button.
- Container Section:** Shows a container icon, "Container 1", and "0 / 2".
- Bottom Bar:** Contains buttons for "Settings" (gear icon), "Bake out" (wavy lines icon), "View" (line graph icon), "Light" (light bulb icon), and a timestamp "16:01 2016-05-23".

# Proof of Principle/Functional Model

## Initial Sample Test



# Future Developments

HPF for protein crystallography

# Leica Protein Crystallography Solution



HPF



- Standardization
- Reliability
- High throughput



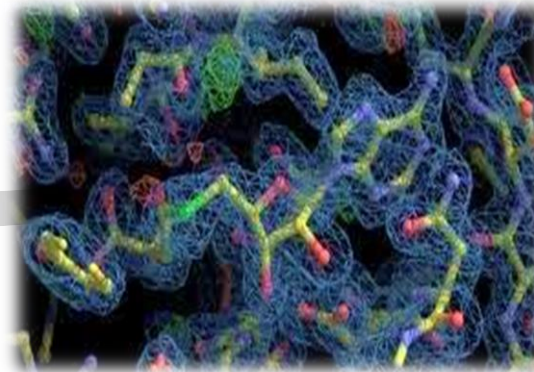
Manual freeze

- Screening for a suitable cryoprotectant
- Disintegration and loss of crystals
- Time consuming



elements  
for a

Drug discovery  
development





Thank You

