

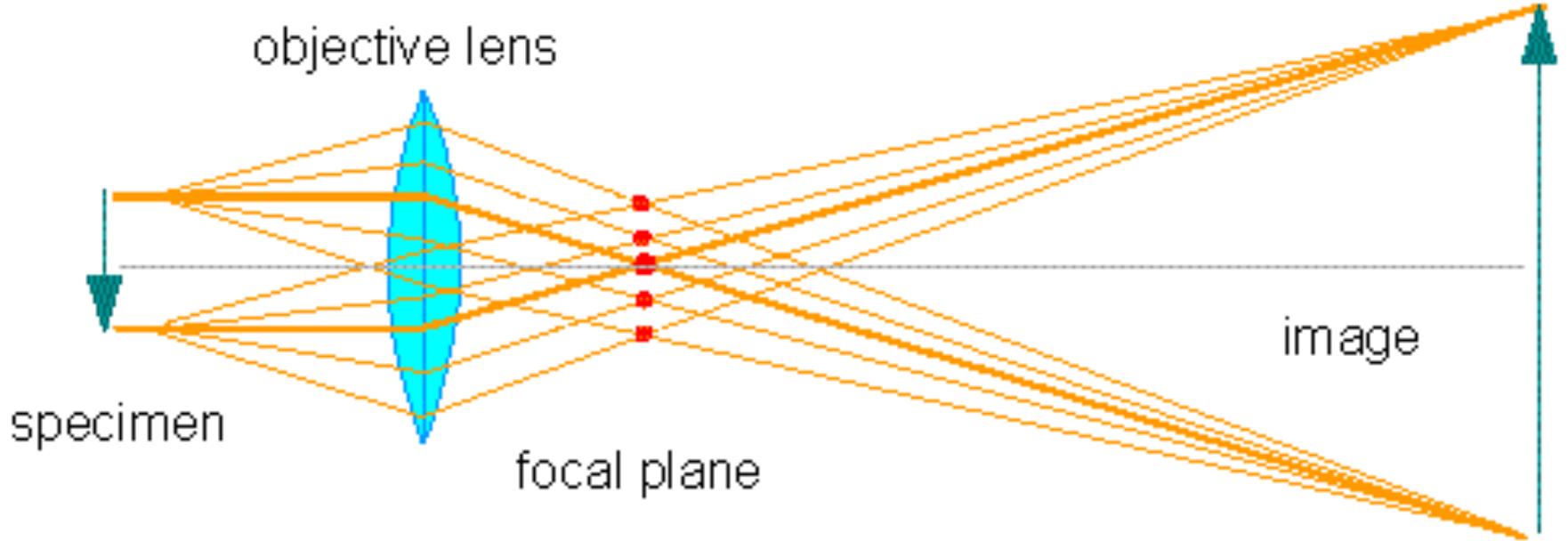
# **Principles of TEM Image formation, particle detection from TEM images and noise handling**

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Indian Institute of Technology, Delhi

# Principle of image formation



$$L_1/L_2 = A_1/A_2,$$

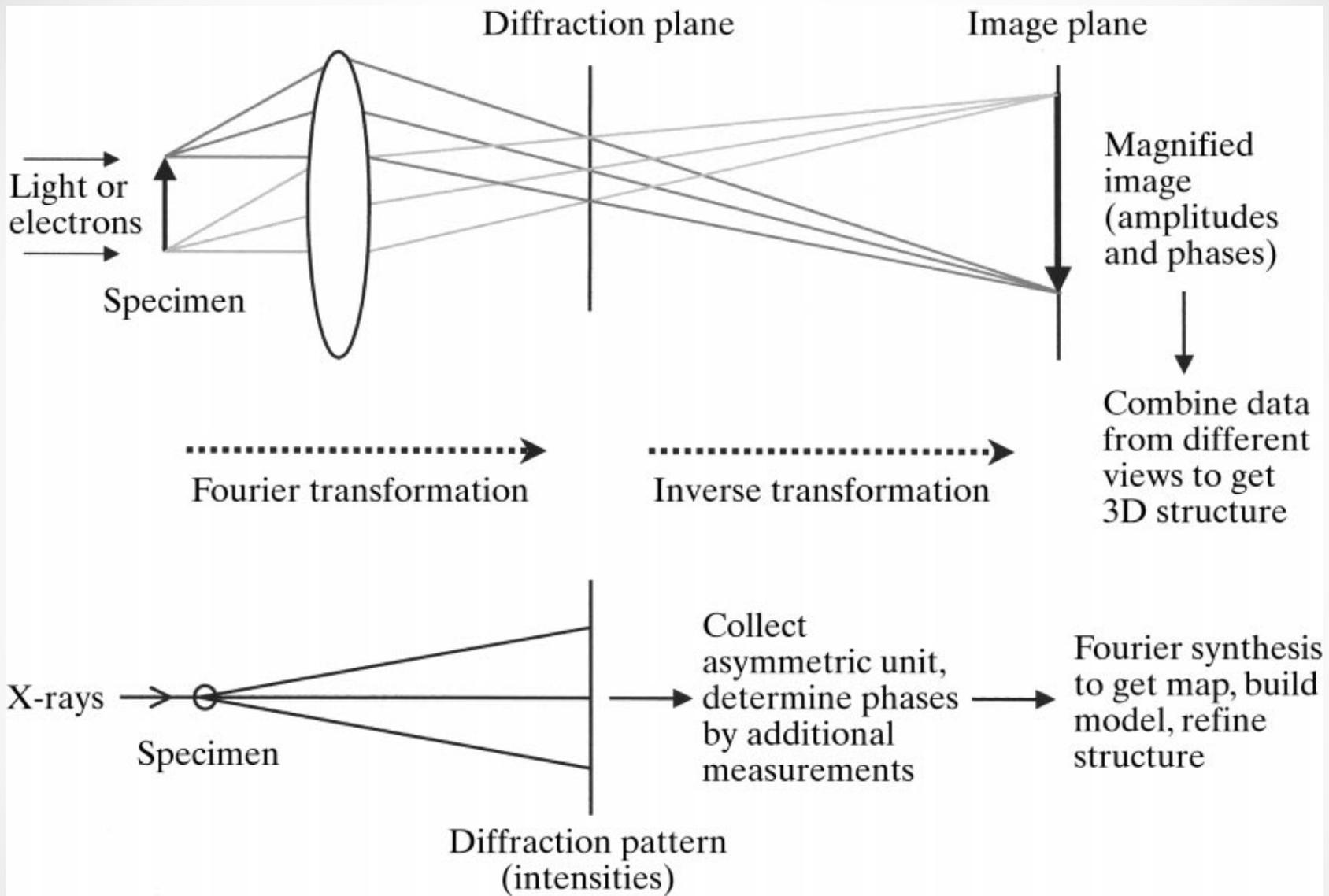
where,  $L_1$  = distance of object from lens

$L_2$  = distance of object from image

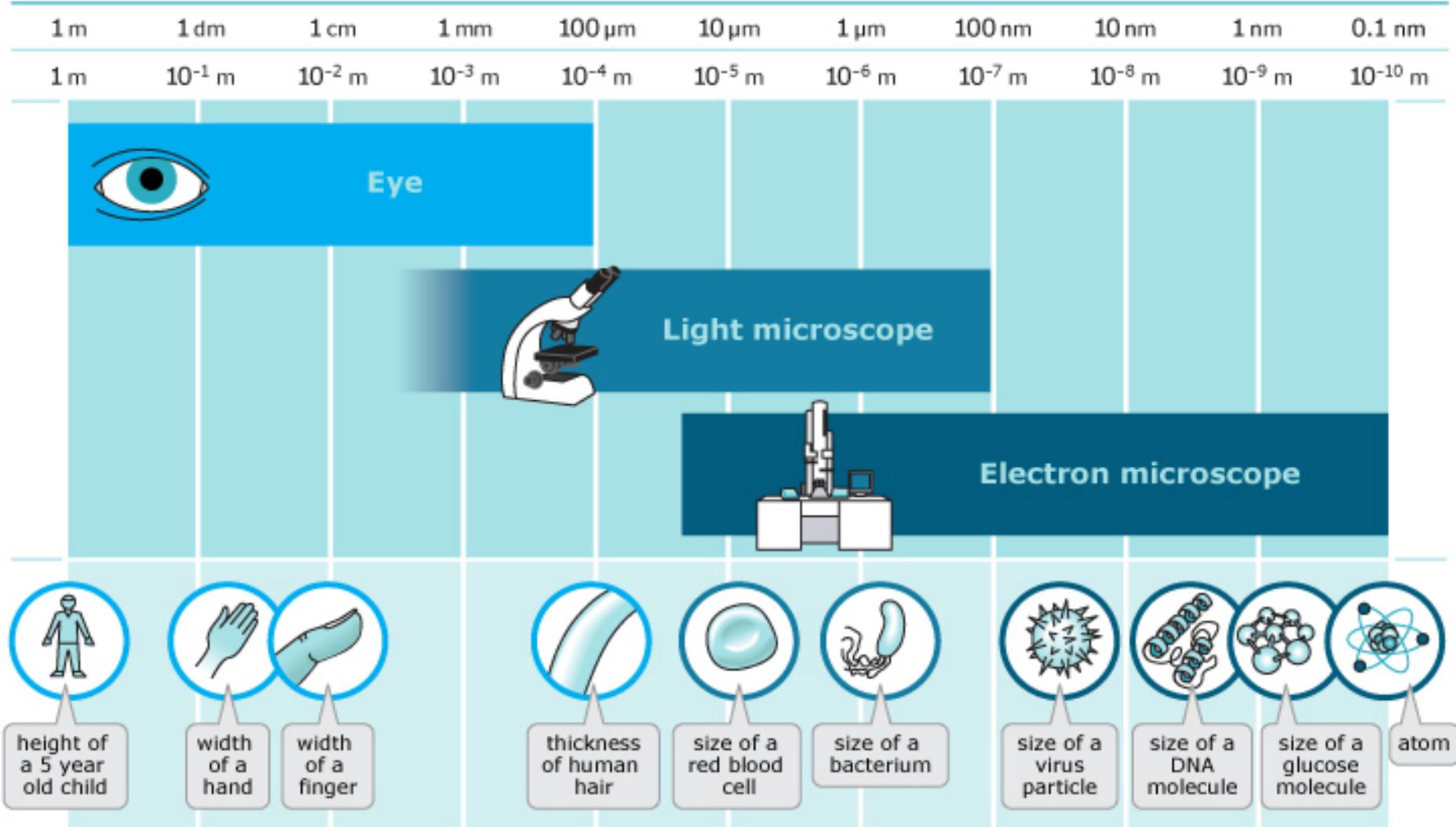
$A_1$  = size of object

$A_2$  = size of image

# Principle of image formation



# Resolving power of microscopes



# First electron microscope

Major discoveries:

- *Accelerated electrons behave like light in vacuum*
- *Travel in straight lines, wave like properties*
- *Wavelength 100,000 x shorter than visible light*
- *Electric and magnetic fields could be used to bend and focus electrons*

First electron microscope (TEM) designed and built by Ernst Ruska in 1931

Lens for electrons constructed in 1926 by H. Busch



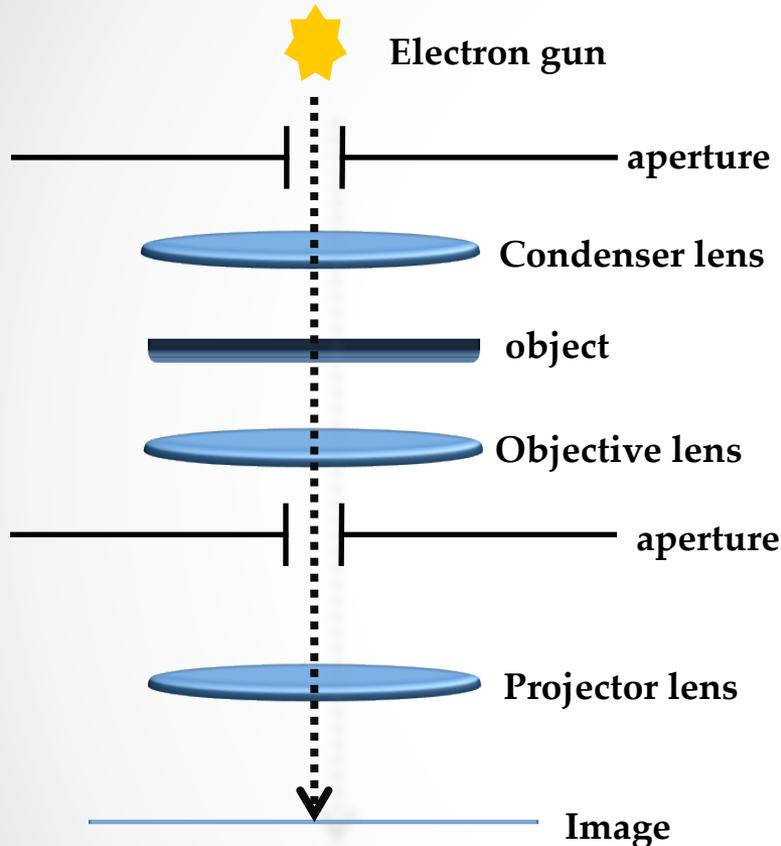
Resolution ~ 100 nm

(~ 200 nm for modern light microscopes)

# Current versions



# Schematic of an electron microscope



Condenser lenses convert diverging electron beam into parallel beam

Objective aperture prevents electrons scattered at high angles from reaching image plane, improves contrast

Provides initial magnification (20-50x)

Aberration affects image formation

Intermediate and projector lenses further magnify image

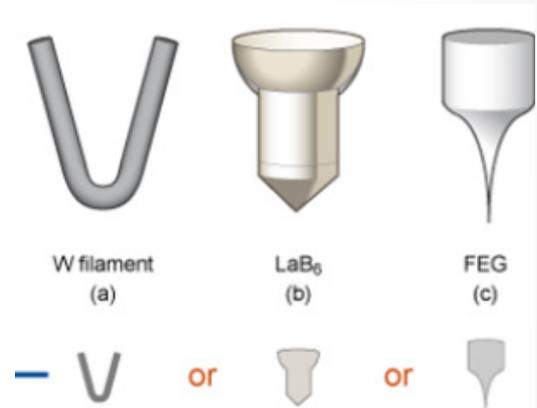
# Sources of electrons

## Tungsten filament

- Heated to 2000-3000 °C
- Thermionic emission
- Electrons accelerated by electric field between anode and filament
- Energy distribution 2.5 eV
- 40-50 Kx magnification

## LaB<sub>6</sub> crystal

- Thermionic emission
- Electrons produced from crystal vertex
- Lower temperature required, lower work function
- Better brightness and lifespan compared to tungsten
- Require higher vacuum levels
- Energy distribution 1.5 eV
- 50-100 Kx magnification



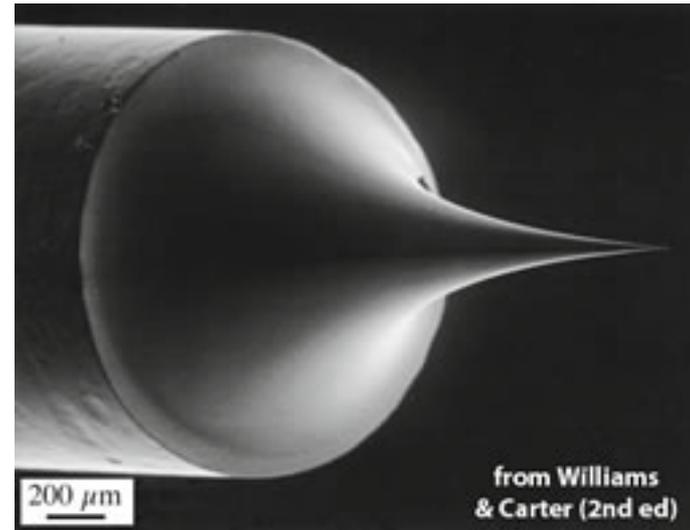
# Sources of electrons

## Schottkey type Field emission gun (FEG) source

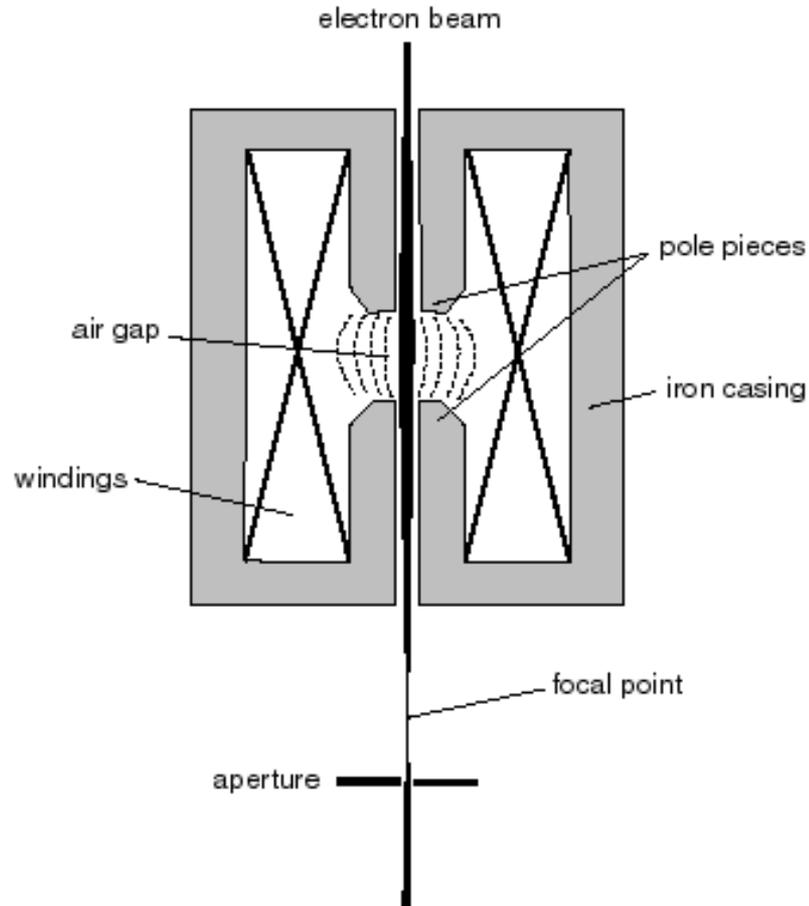
- Single crystal tungsten tip sharpened to 10-25 nm diameter
- Coated with  $ZrO_2$
- Thermally emitted electrons, extracted by strong potential gradient (field emission)
- Accelerated through 100-300 KV
- Extremely bright ( $\sim 500x$  more than tungsten), very coherent
- Energy distribution 1.0 eV
- $> 100$  Kx magnification

## Cold FEG

- No heating required
- Better brightness
- Energy distribution 0.25 eV
- More intense maintenance

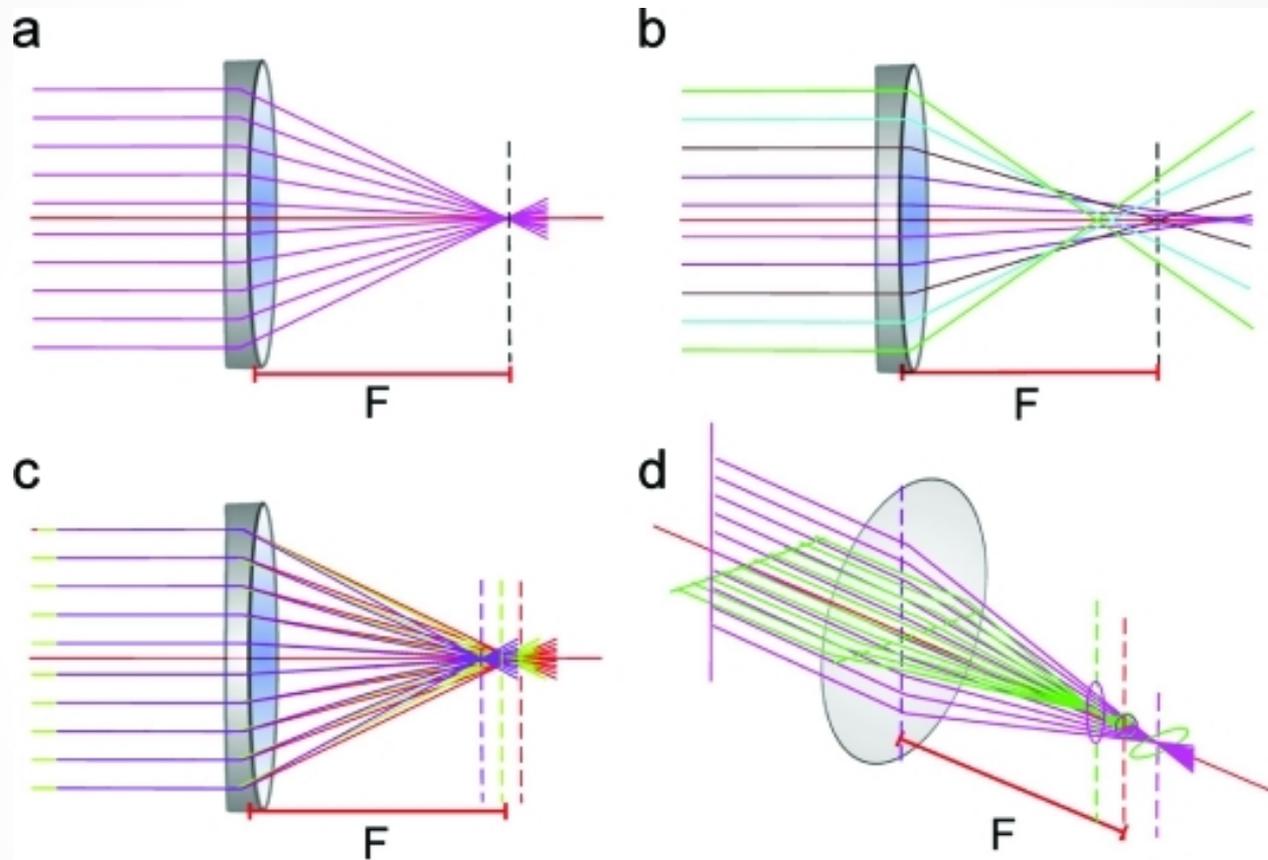


# Lenses in electron microscopy

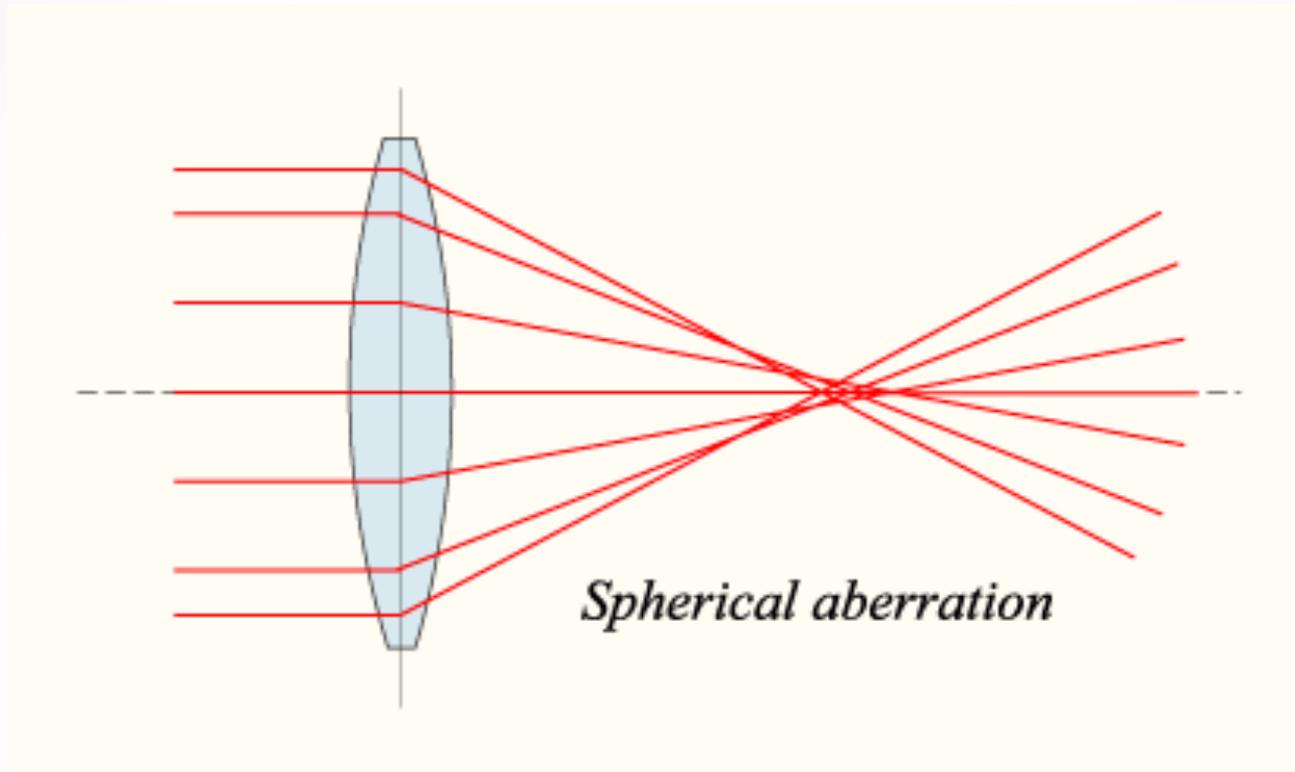


Electromagnetic lenses, varying current in coils alters lens power

# Lens aberrations



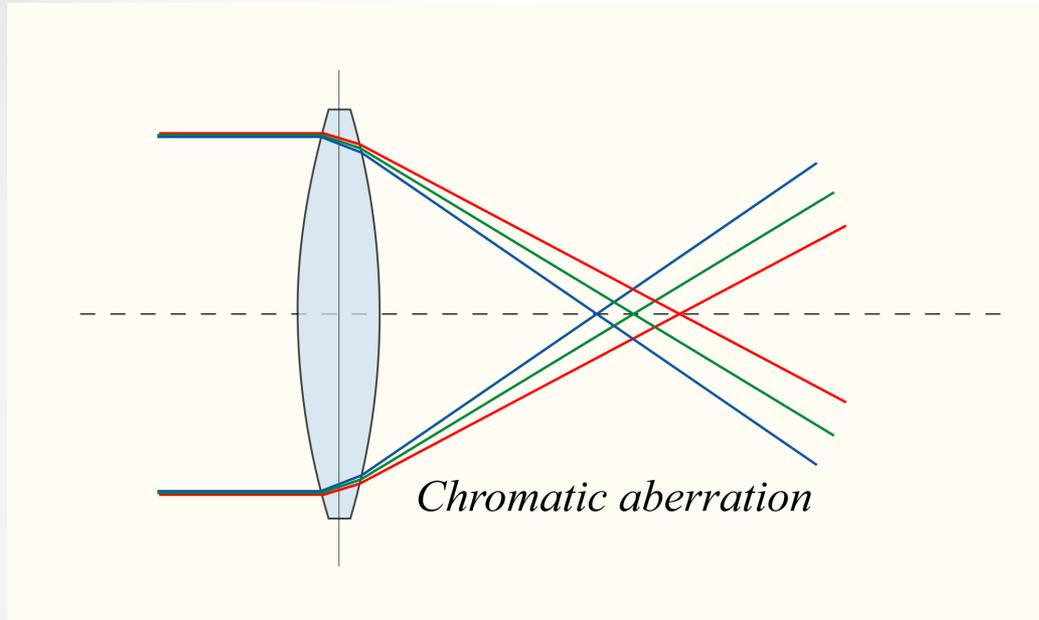
# Lens aberrations



## Spherical aberration

- Diffracted rays with higher angle of incidence converge before the focal point
- correction depends on lens design and manufacture

# Lens aberrations



## Chromatic aberration

- Longer wavelength rays focused more strongly
- Colored halos around images, blurs fine details
- Result of variation in electron energy
- Fixed by stable accelerating voltage

# Lens aberrations



No  
Astigmatism



Regular  
Astigmatism



Irregular  
Astigmatism

## Astigmatism

- Caused by asymmetric magnetic field in lenses
- Point becomes ellipse
- Compensated by stigmator coils

# Interaction of electrons with samples

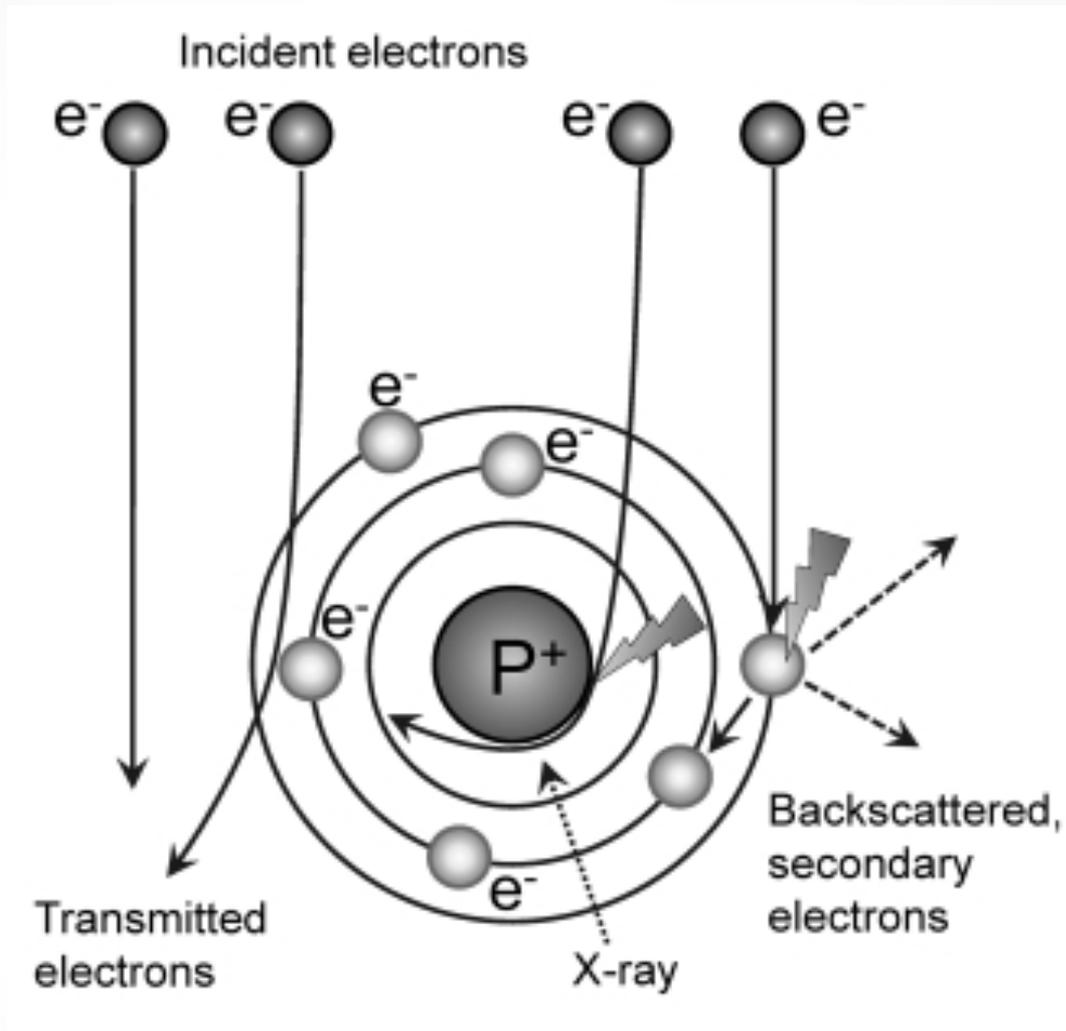
Upon elastic collision of electrons with atom, electrons will be scattered with no change in kinetic energy

- contribute to image formation

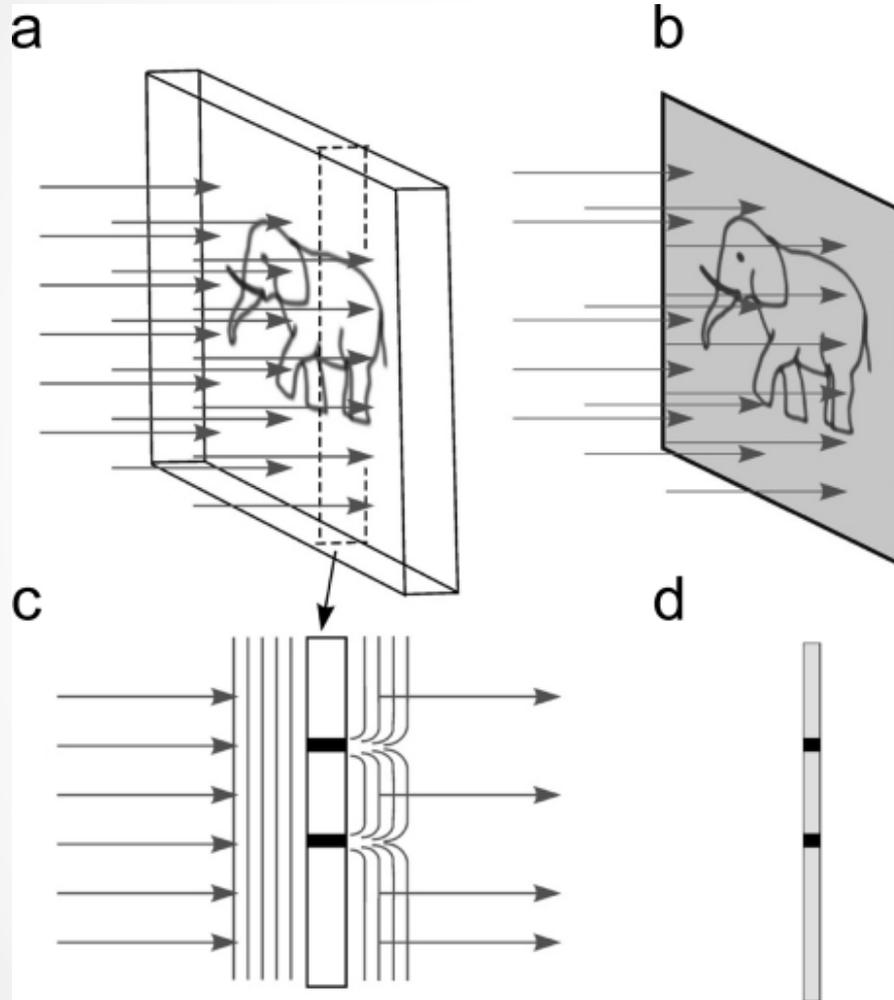
Upon inelastic collision of electrons with atoms, a part of the kinetic energy of electrons is transferred to the atom

- can ionize atoms, generate free radicals, alter chemical bonds, generate X-rays
- contribute to noise

# Interaction of electrons with samples



# Generation of contrast in images

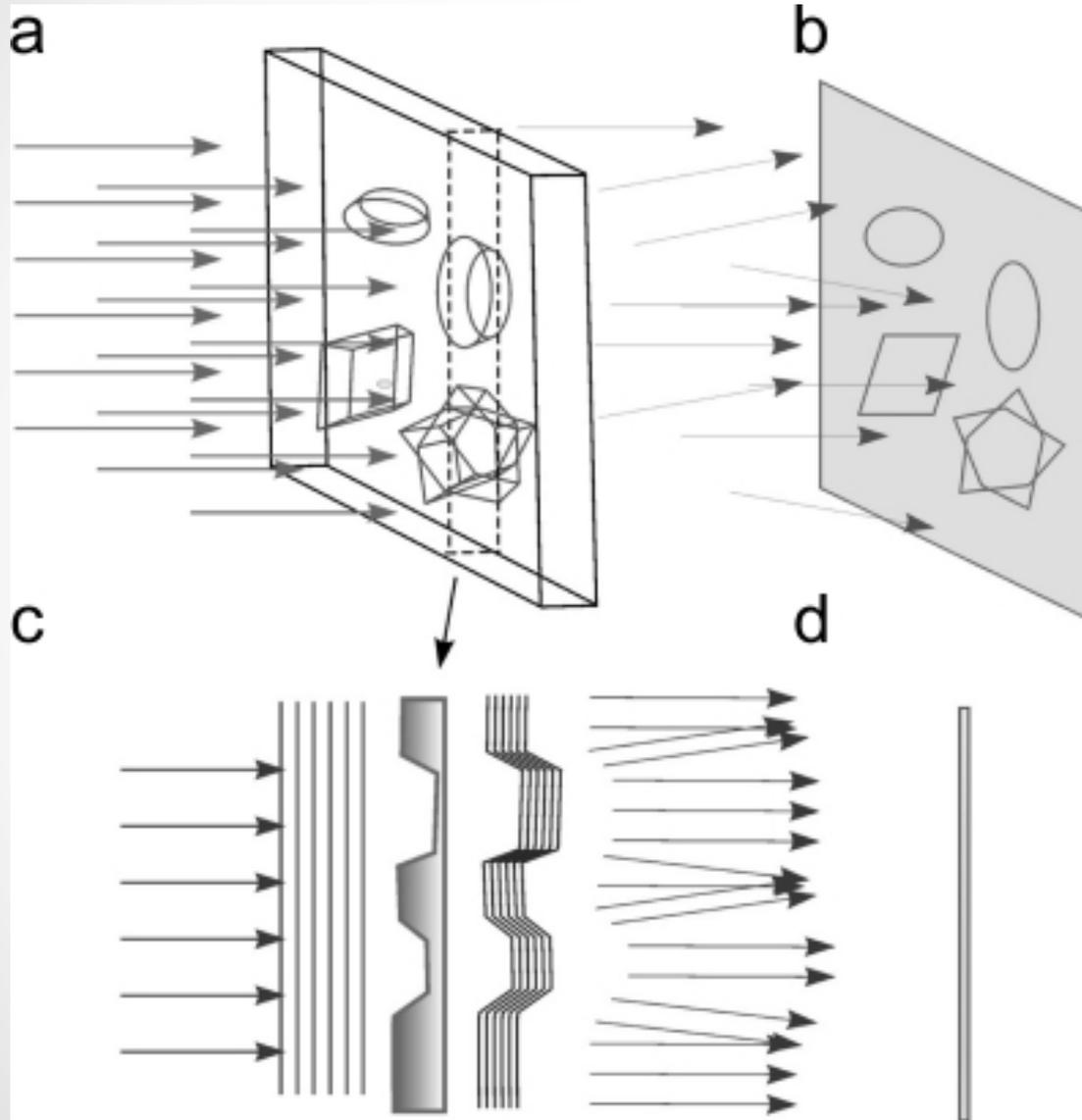


Amplitude contrast:

Part of beam absorbed by the sample  
Produces image contrast

Problem: Biological samples do not  
absorb beam, rather deflect beam  
Intensity difference very small

# Generation of contrast in images



Phase contrast-

Electrons undergo scattering at various angles

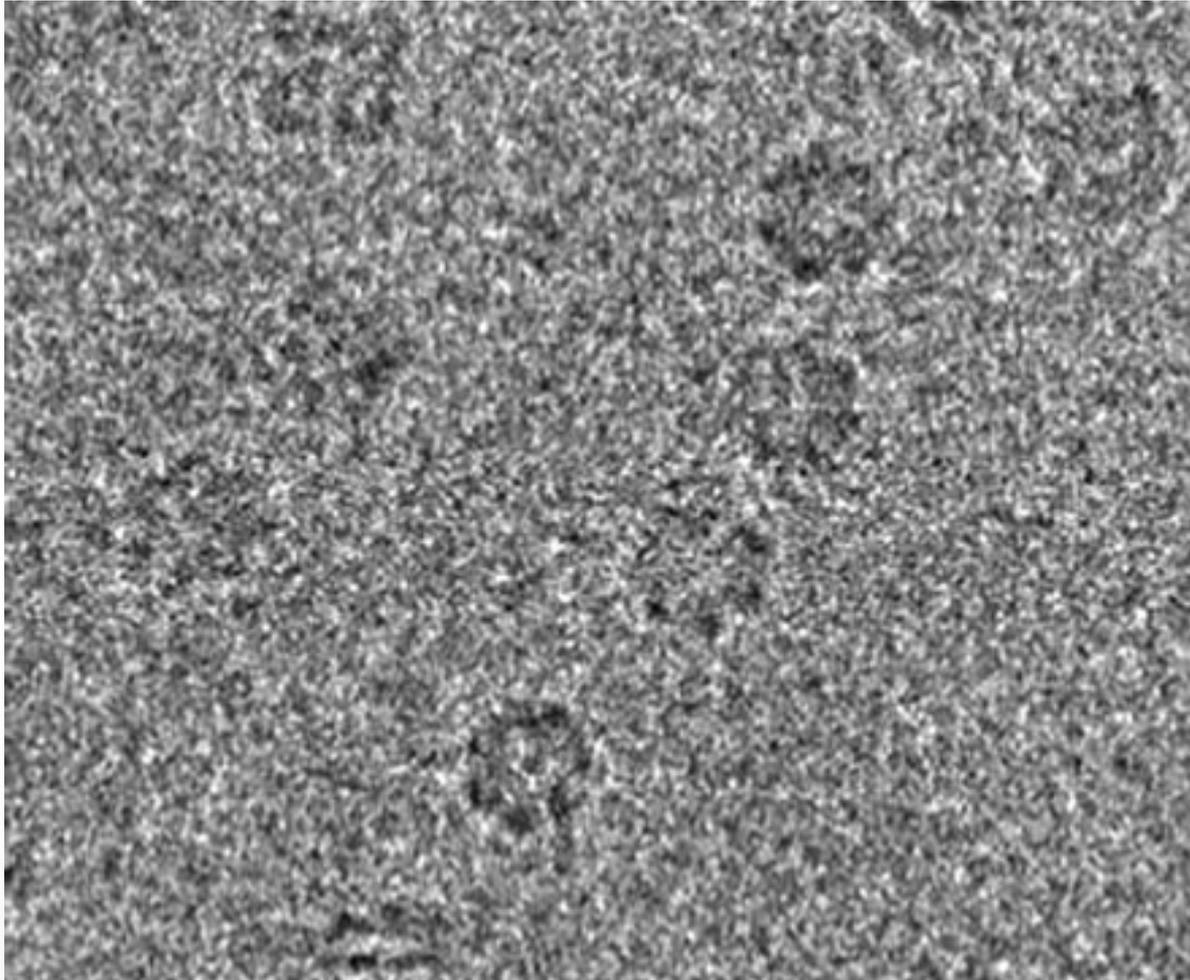
Have different path lengths throughout sample

Emergent beam undergoes constructive or destructive interference with parallel beam

Phase variations may be converted to amplitude variations

## Generation of contrast in images

Biological samples consist of light atoms – C, H, O, N



# Generation of contrast in images

Transparent object varies in refractive index or thickness

Amplitude of emergent wave remains same, phase differs

$$T(x, y) = A_0 \exp [i\phi(x, y)], A_0 = 1$$

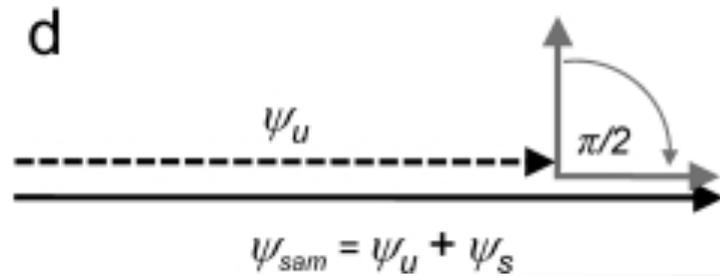
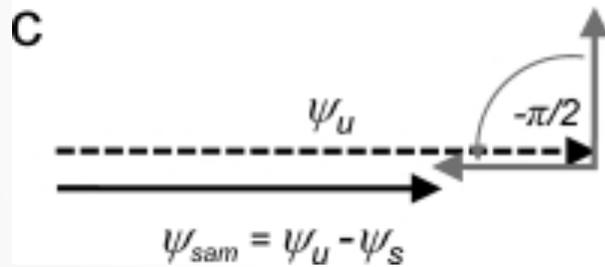
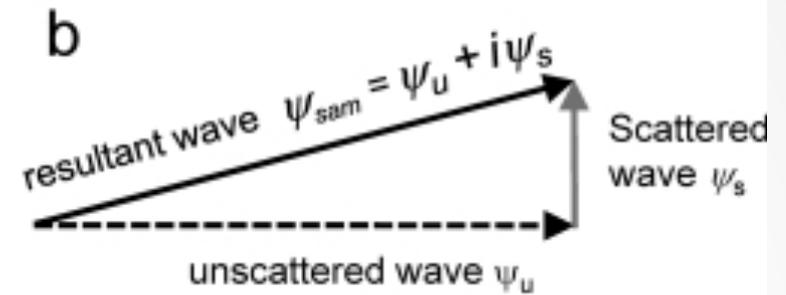
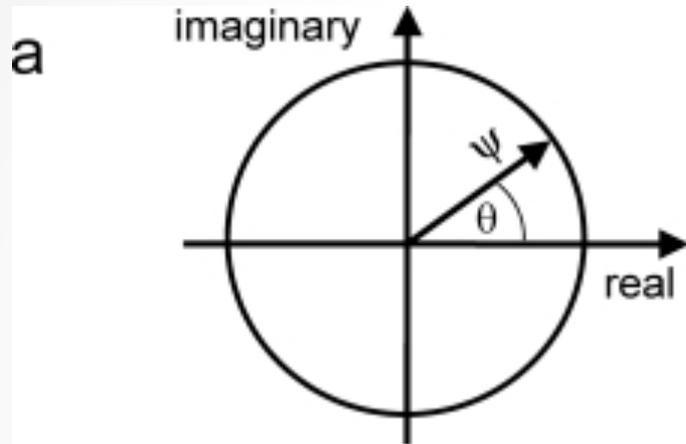
Representation of emergent wave (assuming sample is thin, and phase shift is small):  
 $\exp [i\phi] \approx 1 + i\phi$  (weak phase object)

$$\text{Therefore, } T(x, y) = 1 + i\phi$$

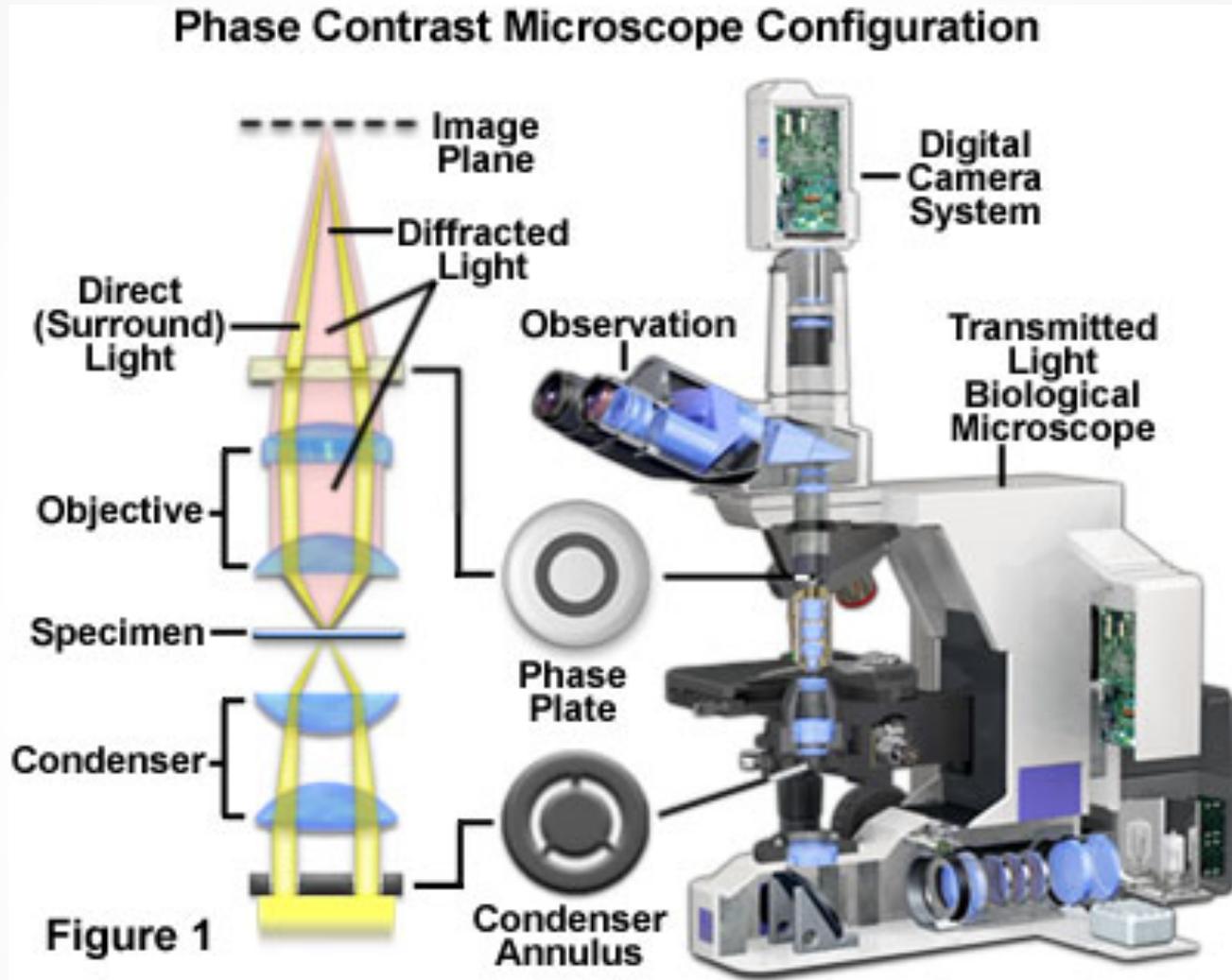
$$\text{Observed intensity: } I^2(x, y) = T^2(x, y) = 1 + i\phi \approx 1$$

$$\text{With additional phase shift of } 90^\circ, \text{ however, } I^2(x, y) = T^2(x, y) = (1 - \phi)^2 \approx 1 - 2\phi$$

# Phase contrast microscopy



# Generation of contrast in images



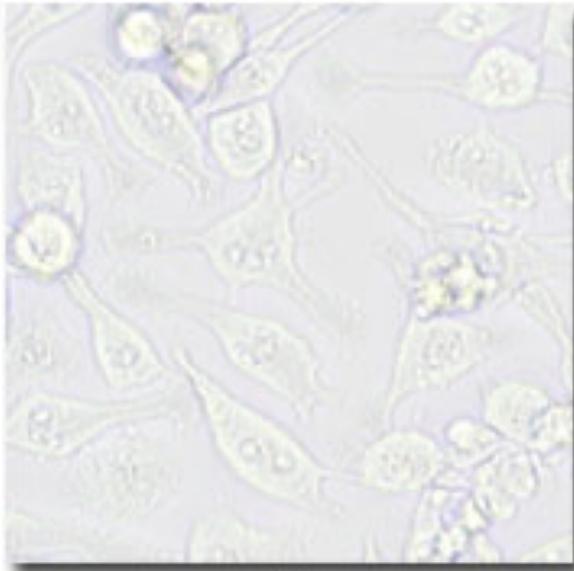
Fritz Zernike (1934) - Phase contrast microscopy

Phase plates introduced in the back focal plane of objective lens

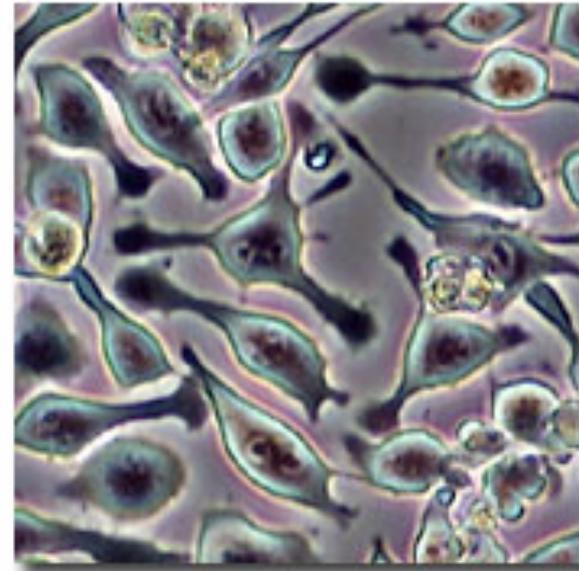
Shifts phase of scattered waves by  $90^\circ$ , amplitude contrast

# Generation of contrast in images

## Living Cells in Brightfield and Phase Contrast



(a)



(b)

Figure 2

Improvement in contrast of biological samples

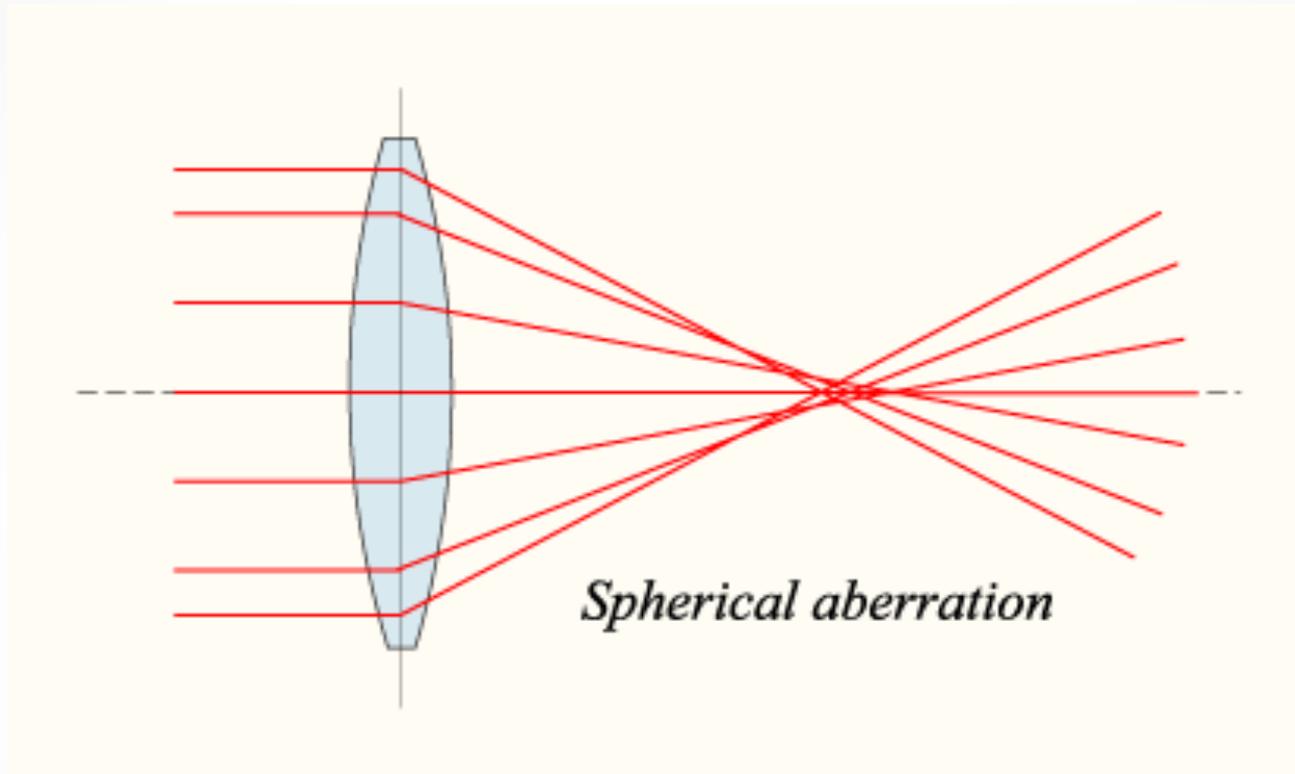
# Generation of contrast in electron microscopy

Combination of various factors generate contrast

- 1) Spherical aberration
- 2) Defocus
- 3) Apertures

Induce phase shift, cut off inelastically scattered electrons

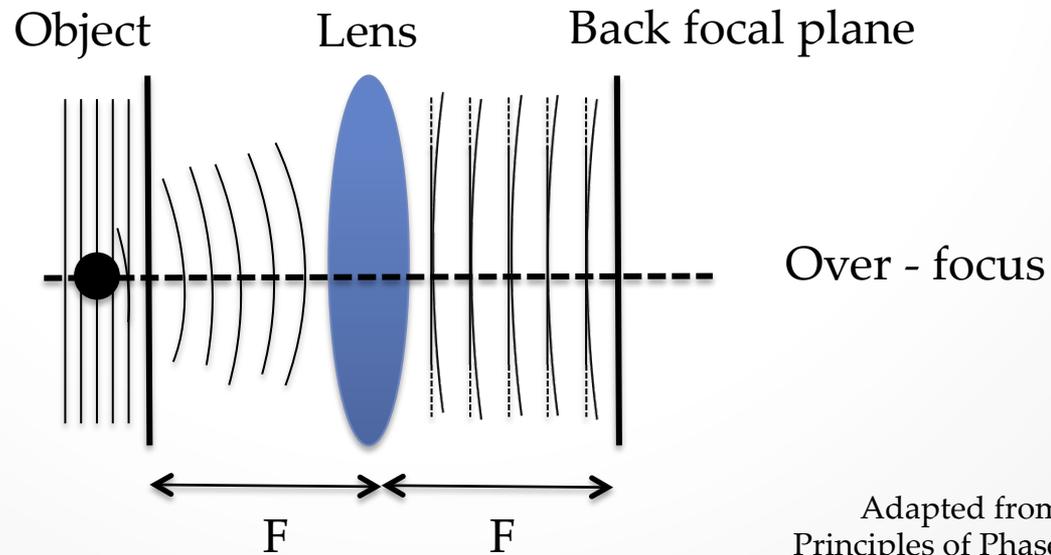
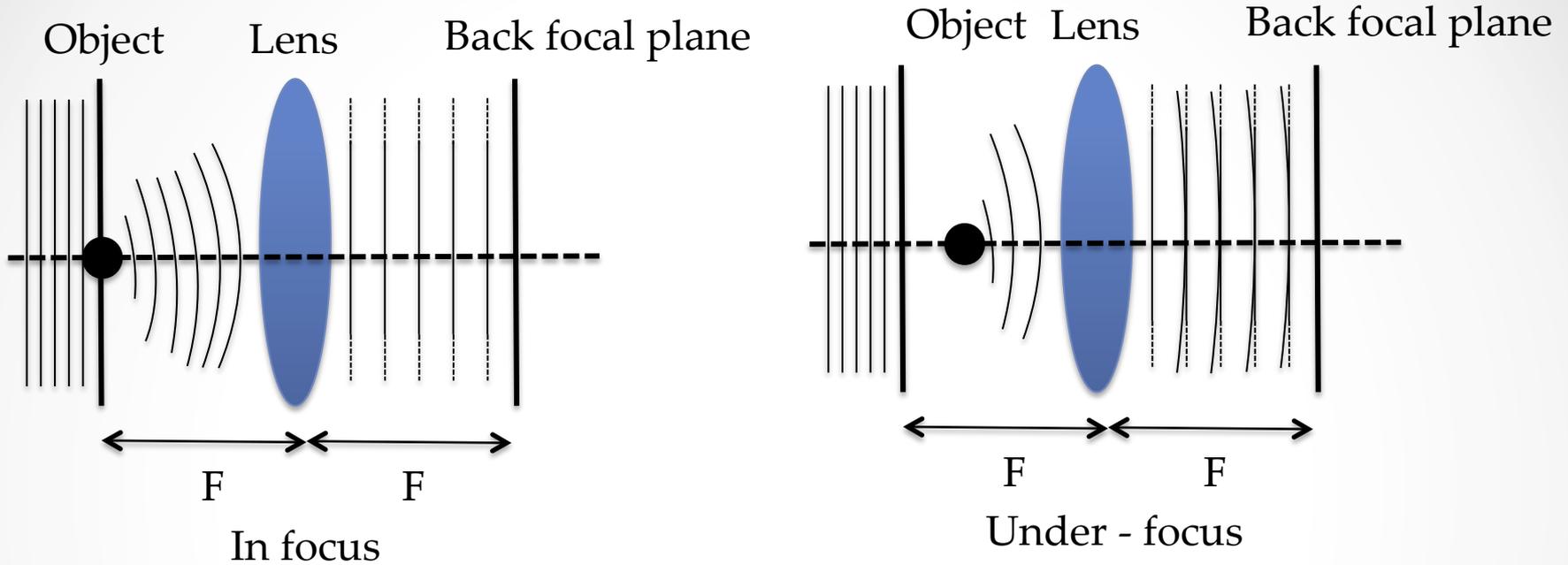
# Generation of contrast in electron microscopy



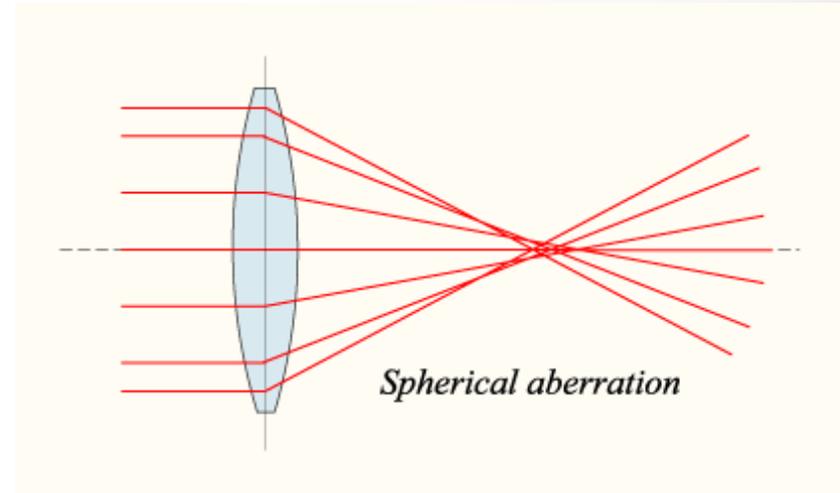
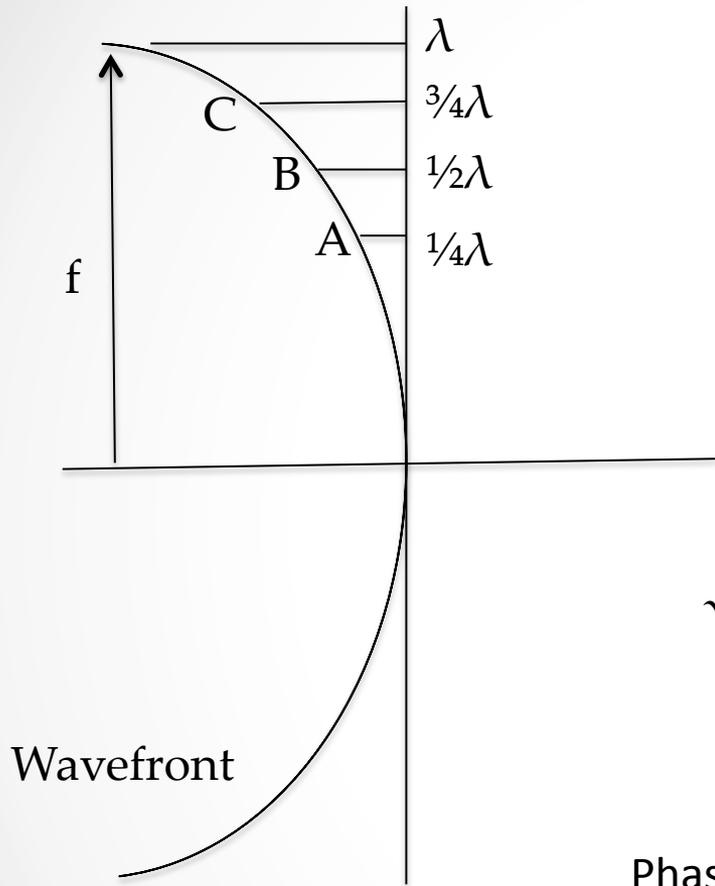
## Spherical aberration

- Diffracted rays with higher angle of incidence converge before the focal point
- correction depends on lens design and manufacture

# Generation of contrast in electron microscopy



# Generation of contrast in electron microscopy



$$\gamma(R) = -2\pi \left( \frac{1}{2} \Delta\lambda R^2 - \frac{1}{4} C_S \lambda^3 R^4 \right)$$

→
→
→

Phase shift      Defocus      Radial frequency      Spherical aberration

Defocus and spherical aberration together cause phase shift at the back focal plane  
 Contrast generation

# Microscope properties affecting image formation

Lens aberrations

Coherence of source

Drift

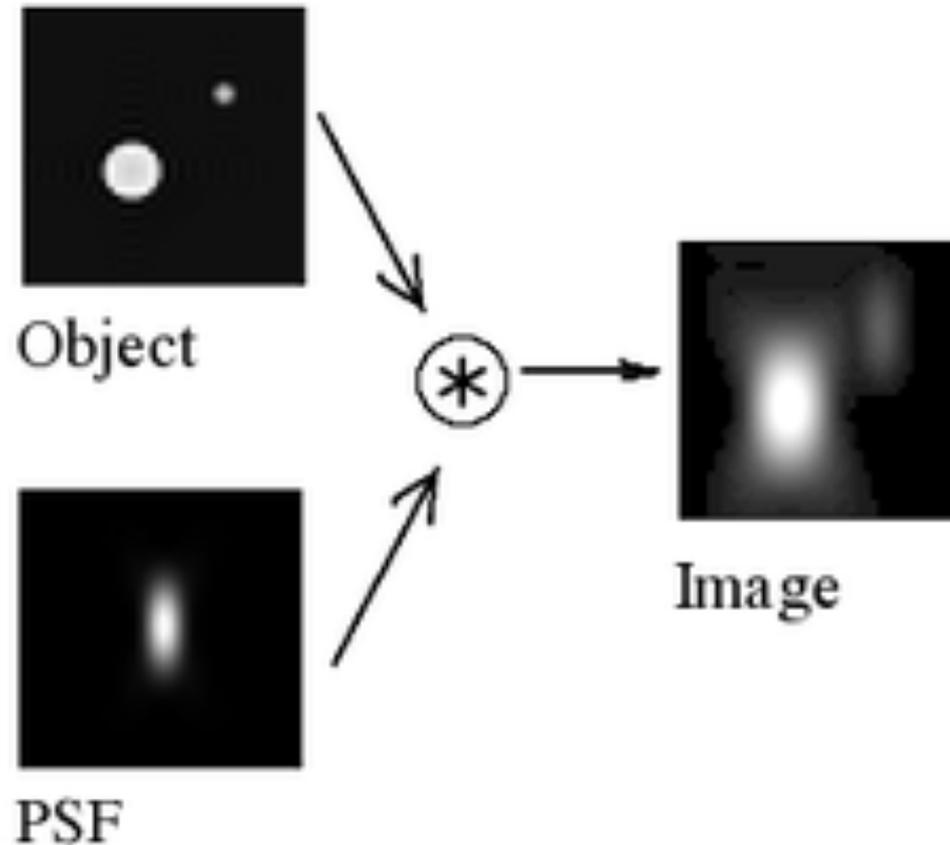
Quality of ice

Alterations in lens current

Quantum noise

Instrumental or environmental instability

# Microscope properties affecting image generation



Point Spread Function (PSF) represents microscope aberrations  
Convolution of Object (FT) with PSF (FT) generates image  
Imperfections transferred to image

# Microscope properties affecting image generation



# Microscope properties affecting image generation

Fourier Transform of PSF = Contrast Transfer Function (CTF)

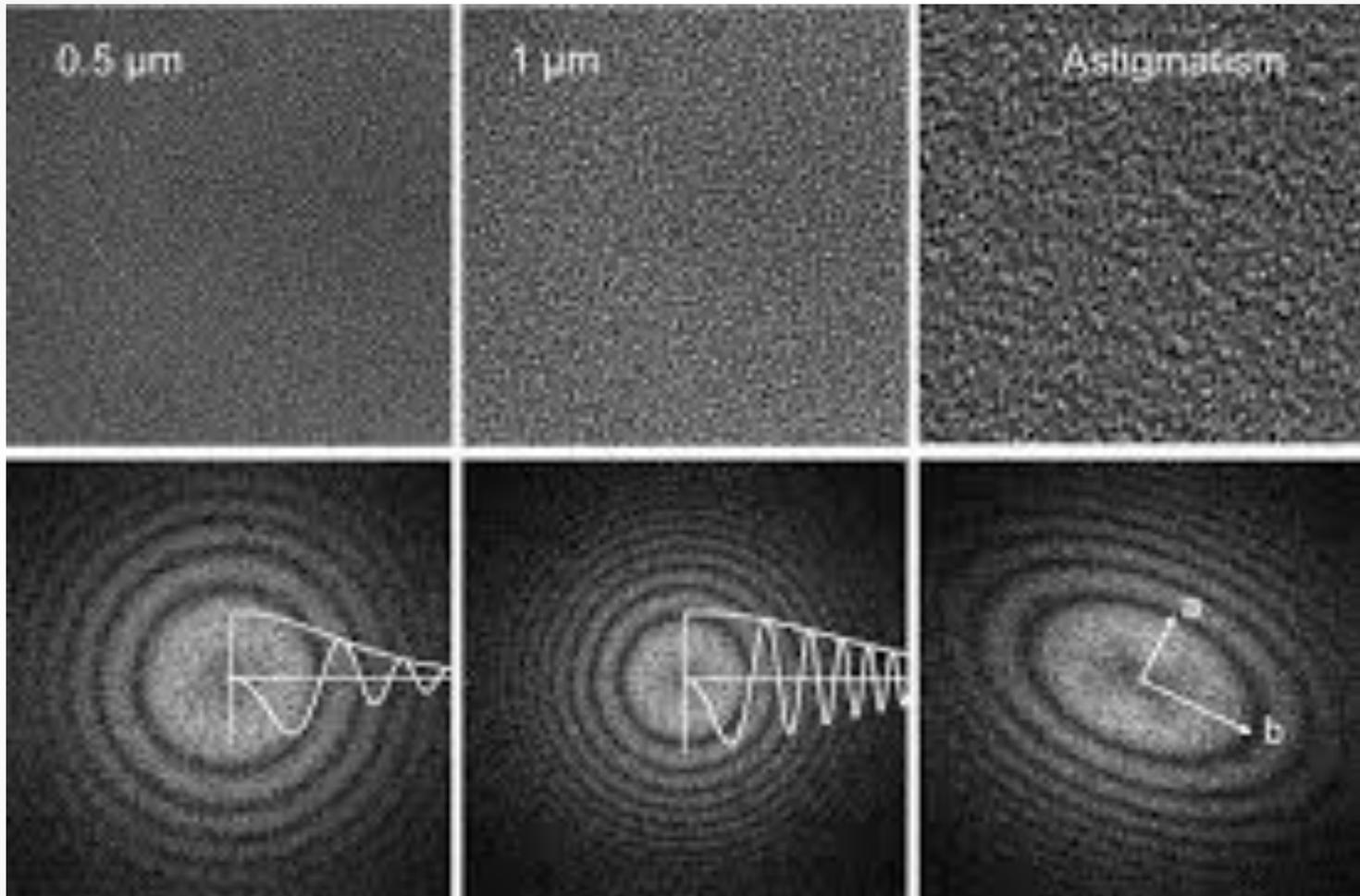
Describes the imaging properties of the objective lens

Can be used to describe the influence of factors on image quality

$$F \{ \overset{\rightarrow}{\Psi}_{\text{obs}}(\mathbf{r}) \} = F \{ \overset{\rightarrow}{\Psi}_{\text{sam}}(\mathbf{r}) \} \cdot \overset{\rightarrow}{\text{CTF}}(\mathbf{R}) \cdot \overset{\rightarrow}{E}(\mathbf{R})$$

$$F \{ \overset{\rightarrow}{\text{PSF}}(\mathbf{r}) \} = \overset{\rightarrow}{\text{CTF}}(\mathbf{R}) \cdot \overset{\rightarrow}{E}(\mathbf{R})$$

# Microscope properties affecting image generation



Effect of defocus and astigmatism on CTF  
Acts as a band pass filter

# Imaging cryo samples – Low dose mode

Incident electrons generate heat

Biological samples degrade

Area to be imaged not exposed until the image is taken

Focusing and alignment done on a different site

Electron dose – 5 -10 electrons/Å<sup>2</sup>

Signal to noise ratio (SNR) very low

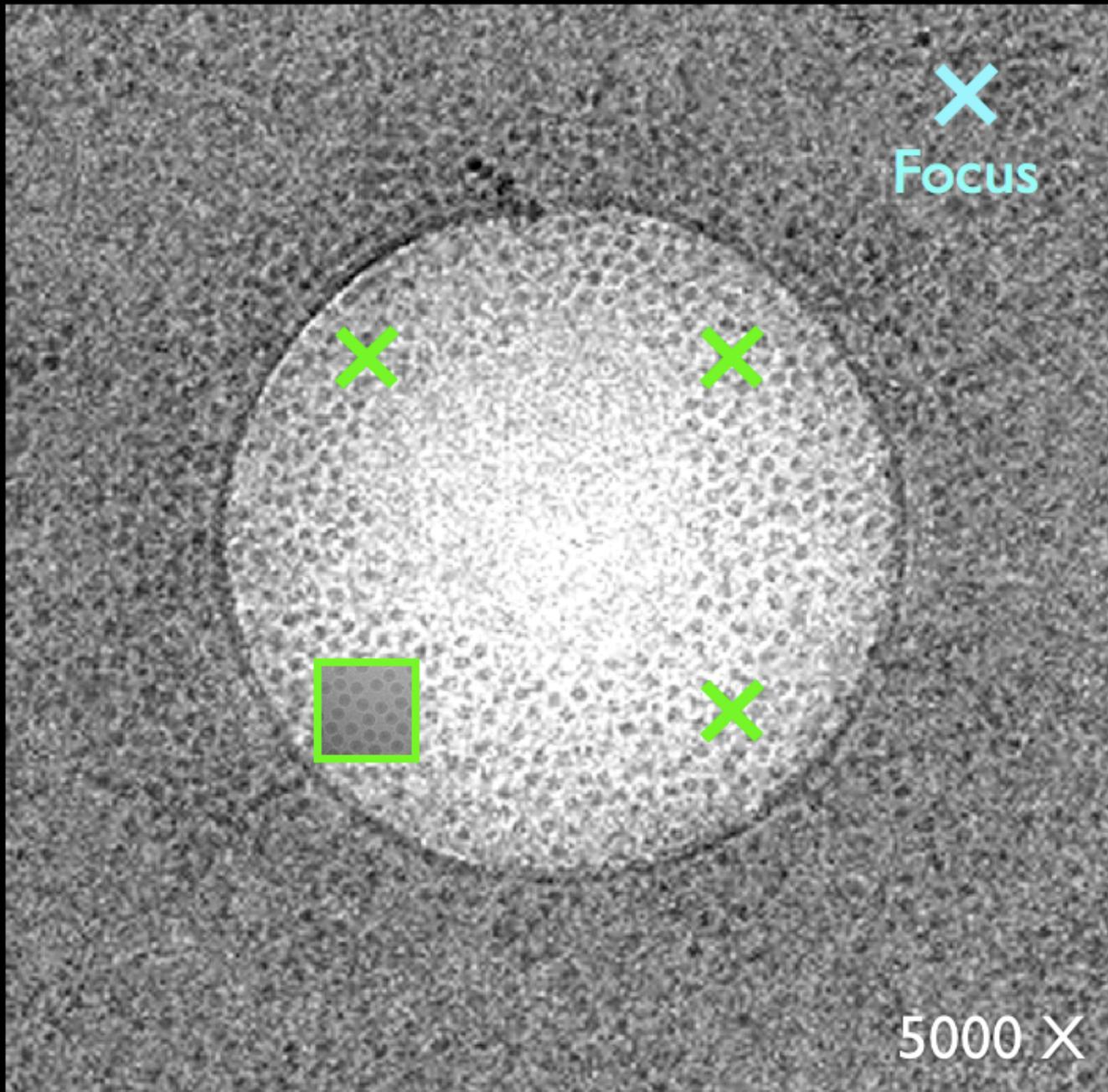


Photo Credit: Gabriel Lander, TSRI

# Methods for contrast and SNR improvement

Sample level: Cryo-negative staining

Microscope level: phase plate, energy filters, aperture size, defocus

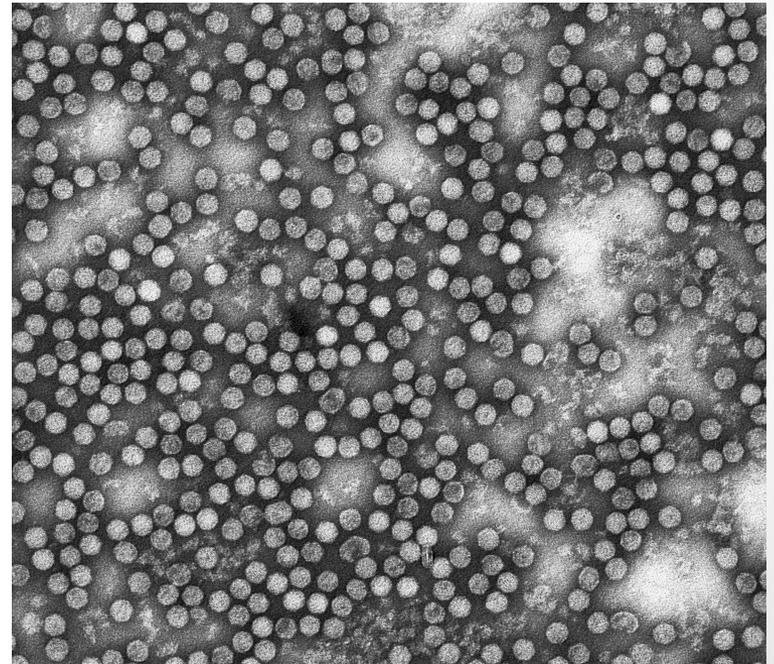
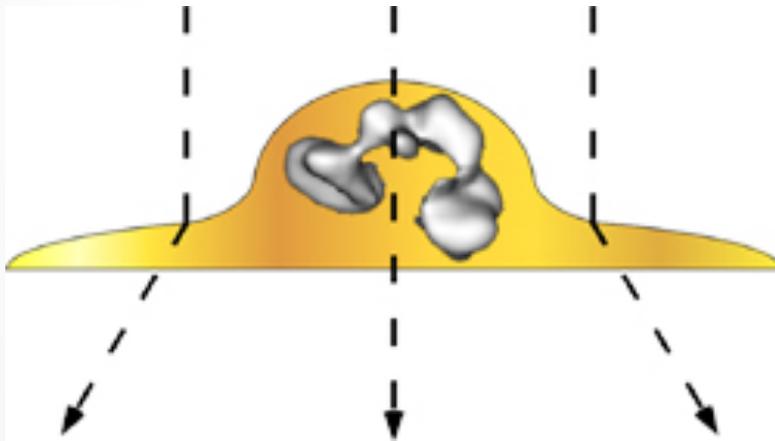
Data collection level: direct detectors, automated collection

## Negative staining with heavy metal

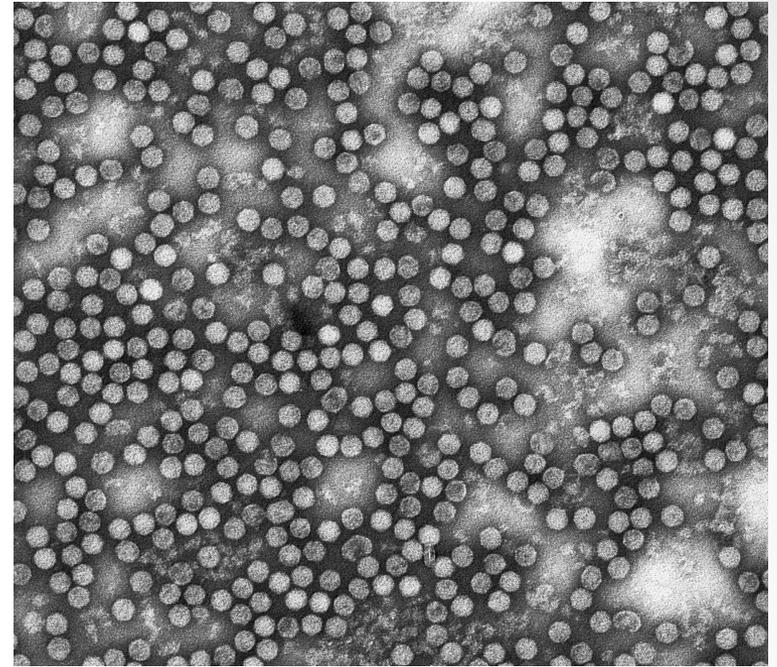
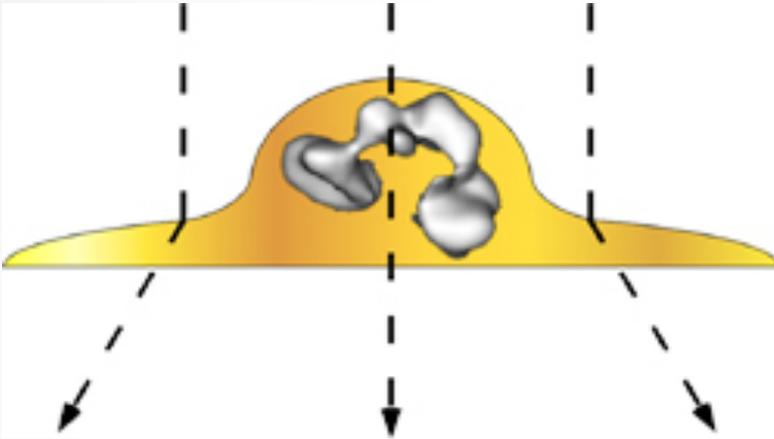
Very small amount of electrons absorbed by biological samples

Heavy metal salt, that absorbs electrons fairly easily, used for negative staining

Uranium, tungsten, molybdenum, vanadium, lead



# Negative staining with heavy metal



Drying step required

May cause dehydration-related damage

Formation of artifacts

Only surface features visible, low resolution

Structural details of external or internal regions not available

Possible to get low resolution reconstructions

Samples may have preferred orientation on continuous carbon film

# Methods for contrast and SNR improvement

Sample level: Cryo-negative staining

Microscope level: phase plate, energy filters, aperture size, defocus

Data collection level: direct detectors, automated collection

# Cryo-negative staining

Prevalent method developed by Marc Adrian in 1998

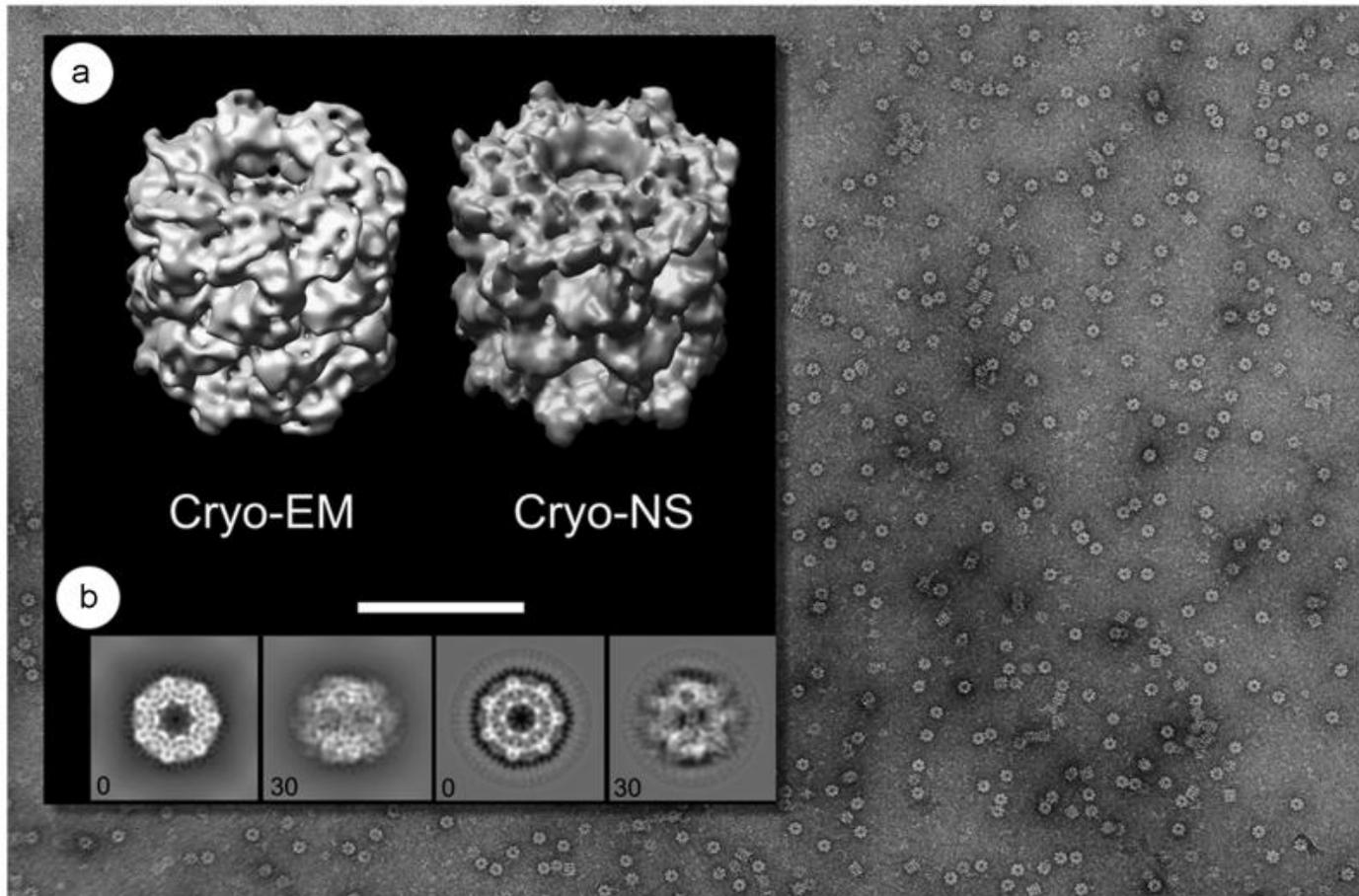
A thin layer of Au/Pd on one side of grid – allows sample spreading

Slurry of ammonium molybdate as staining solution

Quick dip in stain on parafilm, dried for 1-3 s, plunge freezing

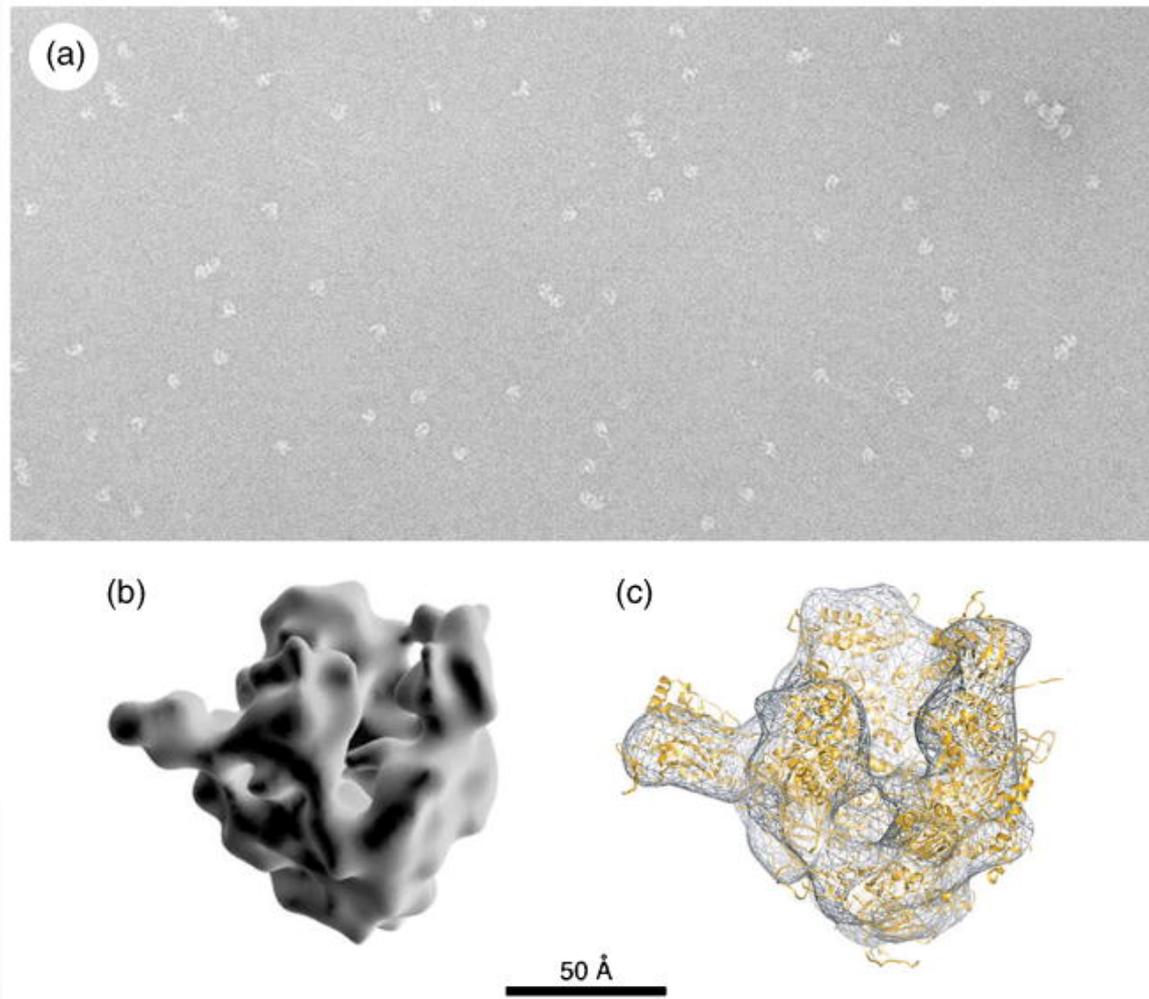
Some dehydration expected

# Cryo-negative staining



Reconstructions of GroEL frozen with and without stain

# Cryo-negative staining



RNA polymerase solved with cryo-negative staining

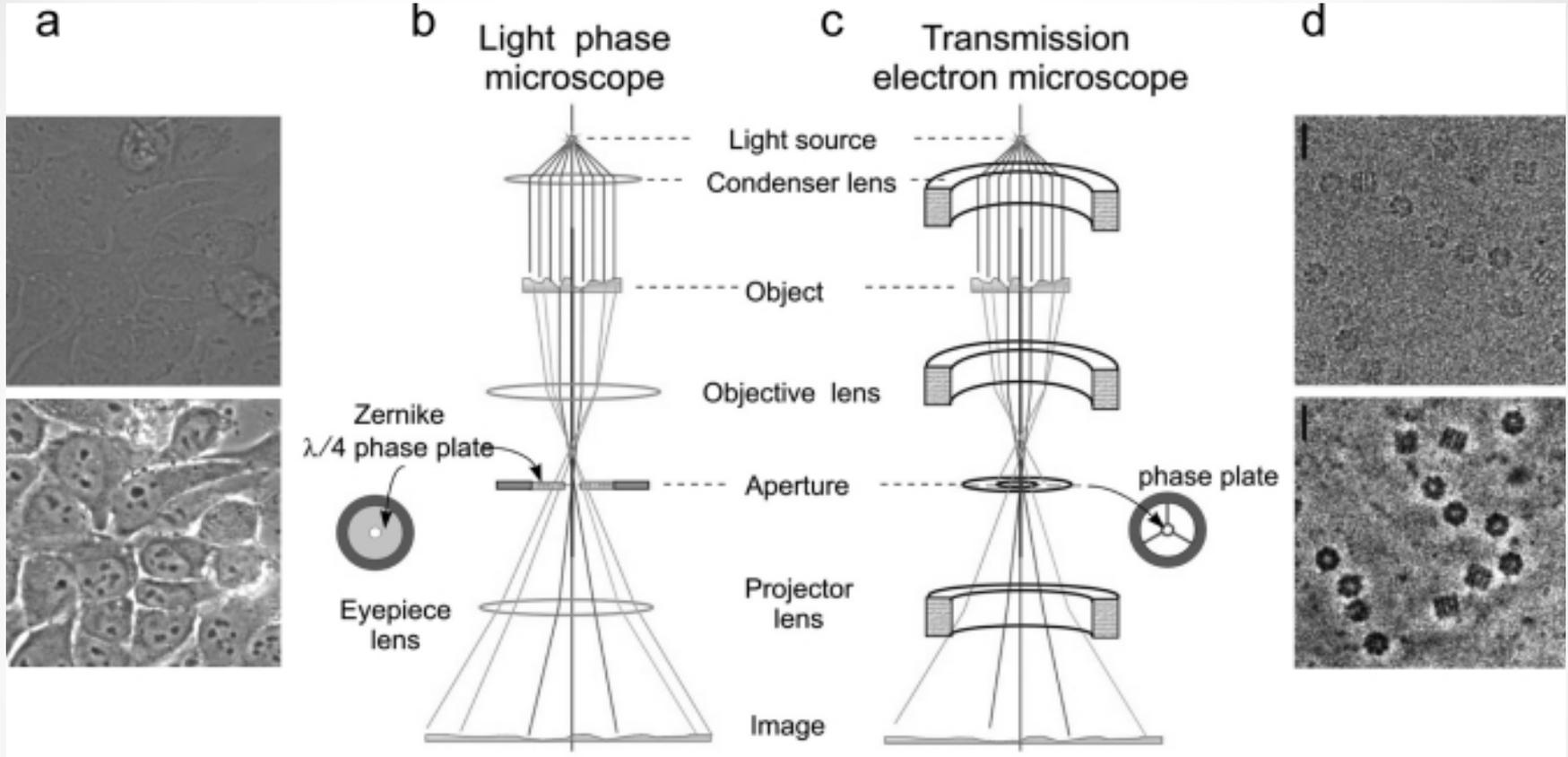
# Methods for contrast and SNR improvement

Sample level: Cryo-negative staining

Microscope level: phase plate, energy filters, aperture size, defocus

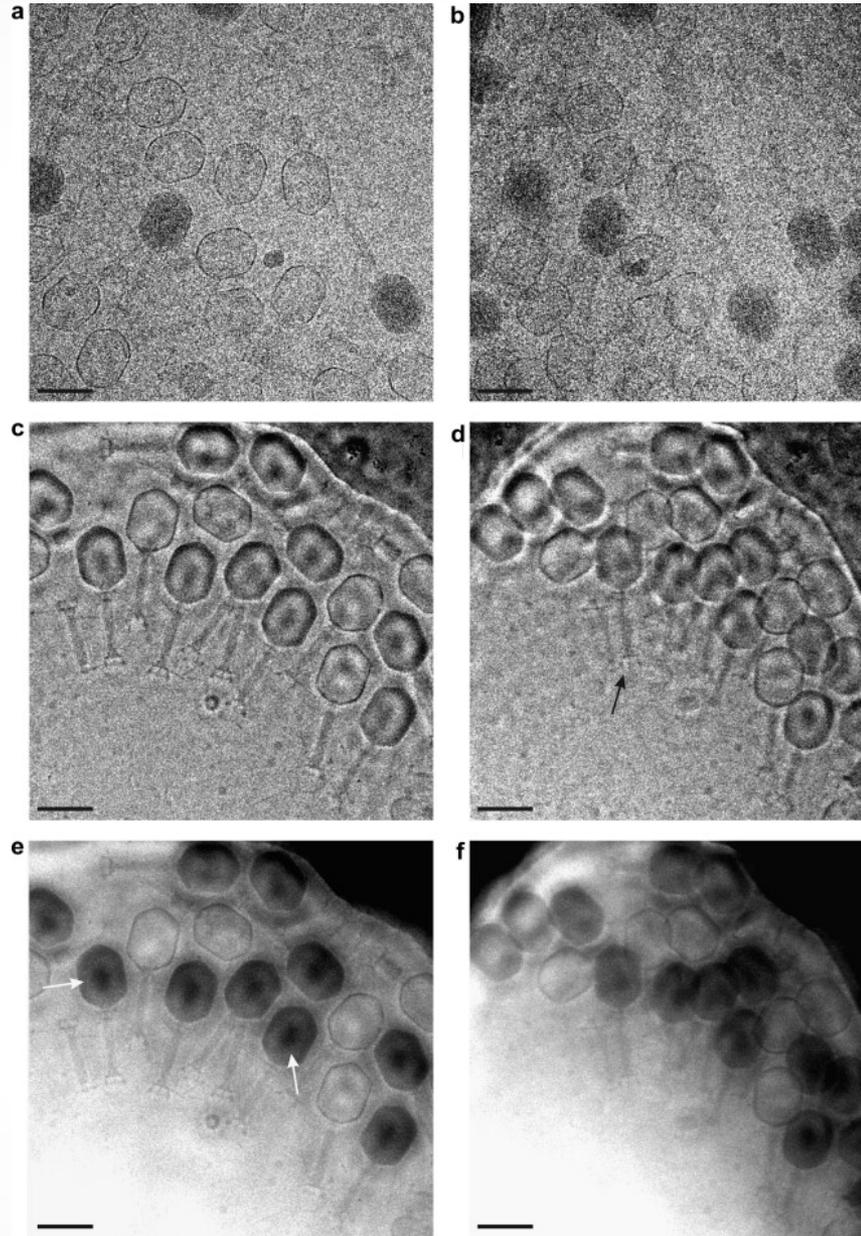
Data collection level: direct detectors, automated collection

# Introduction of phase plate



Phase plates introduced in the back focal plane of objective lens  
Shifts phase of scattered electrons by  $90^\circ$   
Contrast improved upon combination with unscattered electrons  
“Invisible” phase contrast converted into “recordable” amplitude contrast

# CryoET using Zernike phase plate



# Energy filtering

Removal of inelastically scattered electrons

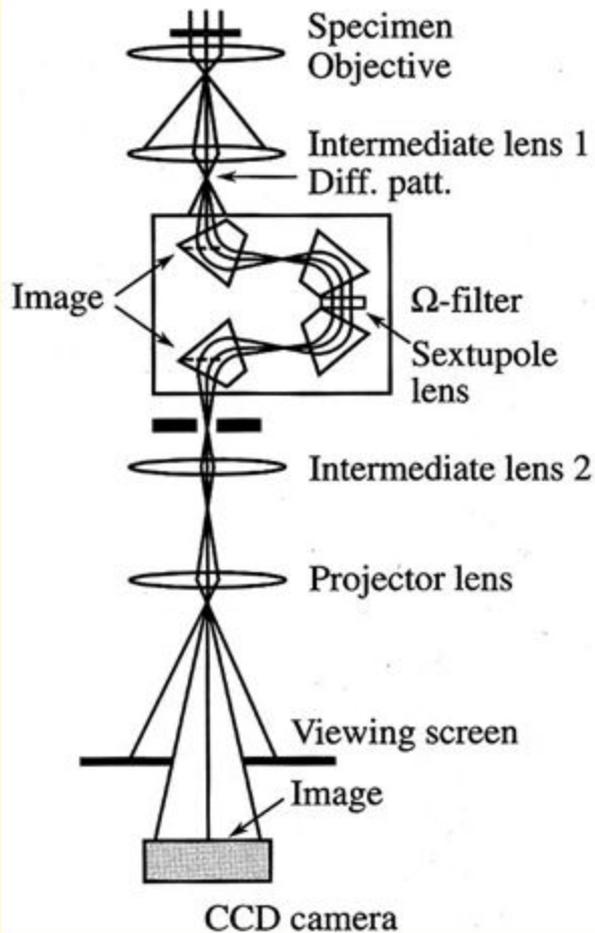
Lower energy, longer wavelength

Chromatic aberration, electrons focused in different planes

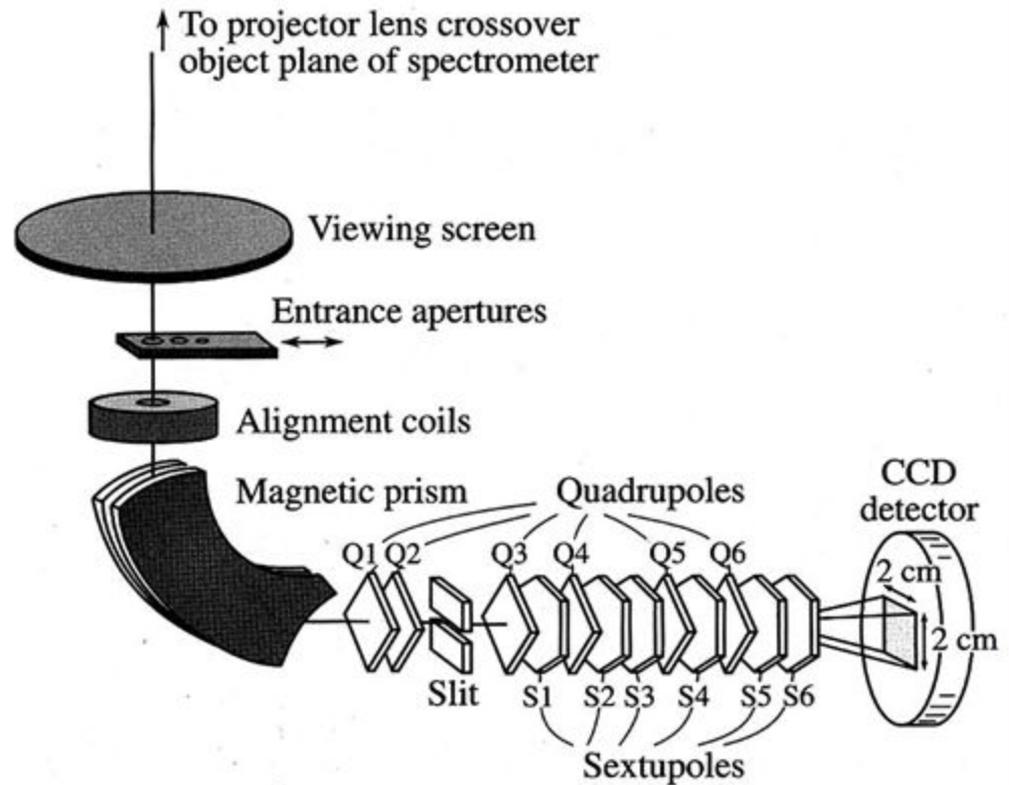
Causes blurriness in image

Removed by in-column or post-column filtration

# EFTEM: In-Column and Post-Column Energy Filters



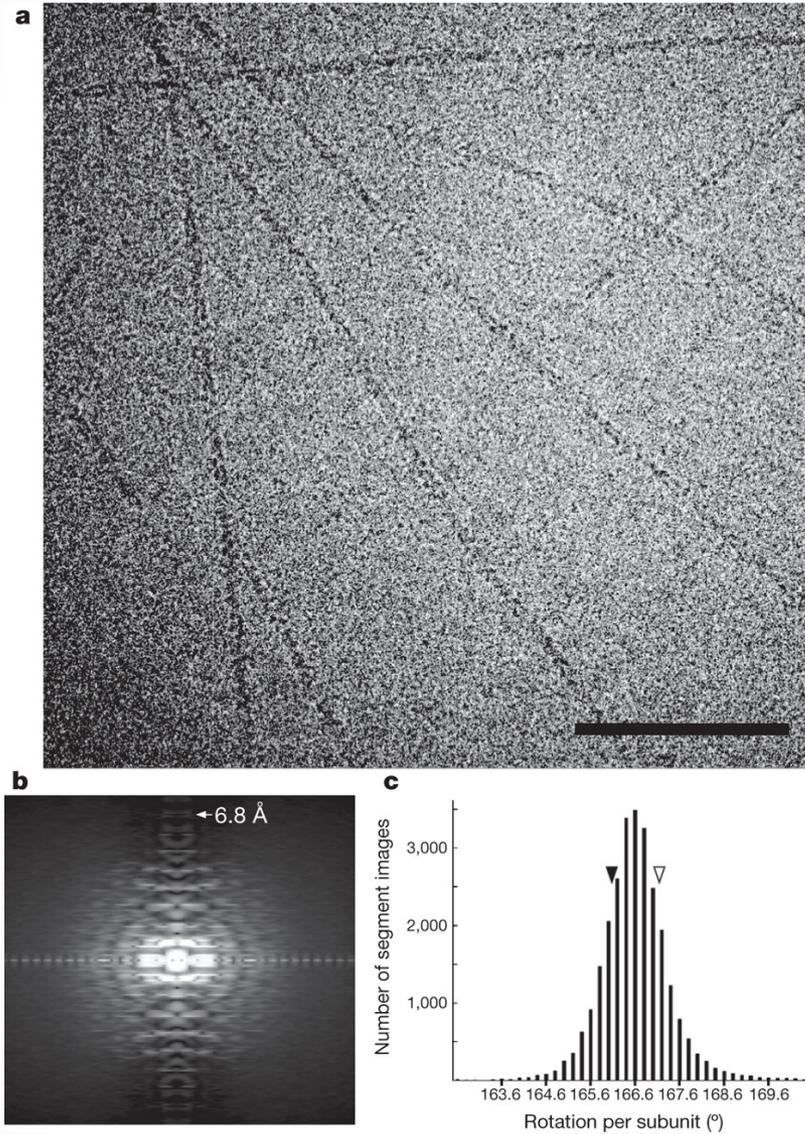
**Omega Filter**



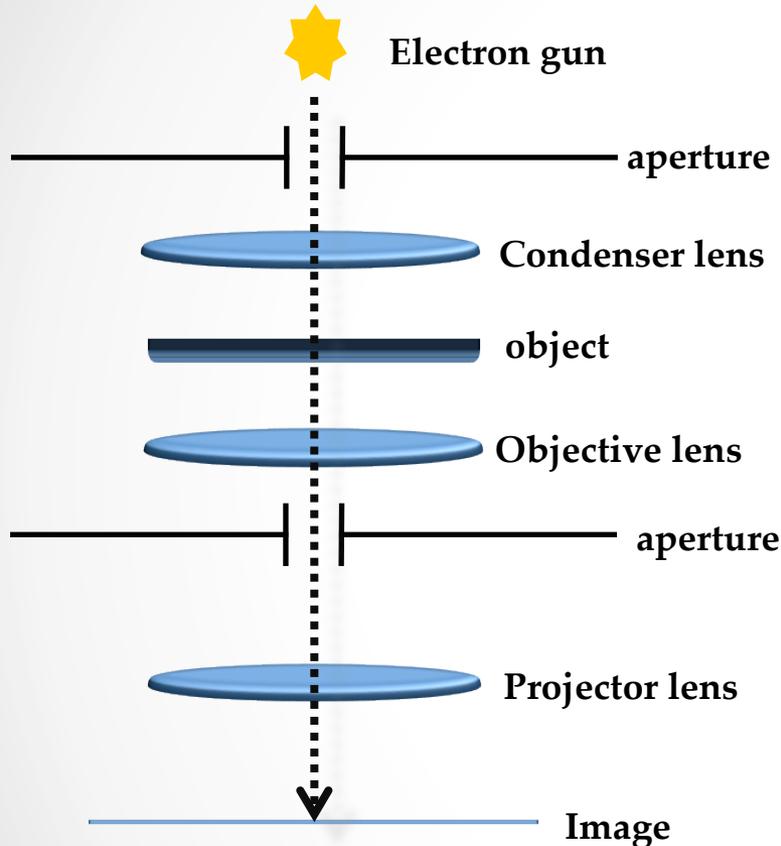
**Gatan Imaging Filter (GIF)**

From Williams and Carter, *Transmission Electron Microscopy*, Springer, 1996

# Imaging of actin filaments using a $\Omega$ type energy filter



# Controlling apertures



Condenser lenses convert diverging electron beam into parallel beam

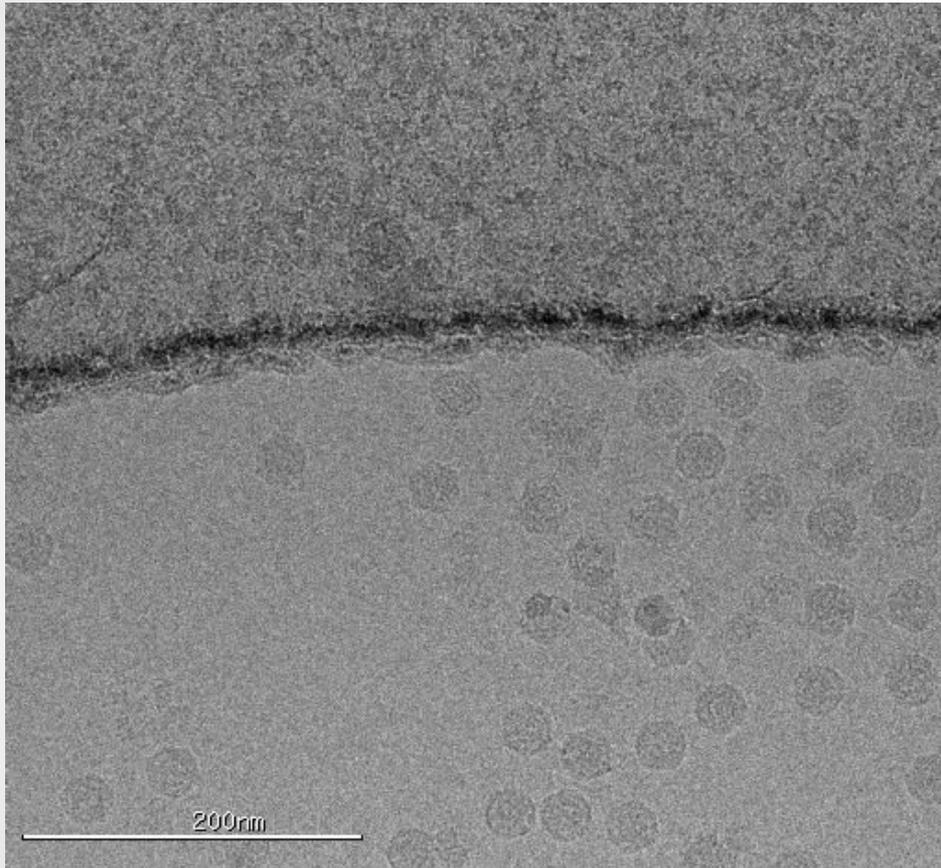
Objective aperture prevents electrons scattered at high angles from reaching image plane, improves contrast

Provides initial magnification (20-50x)

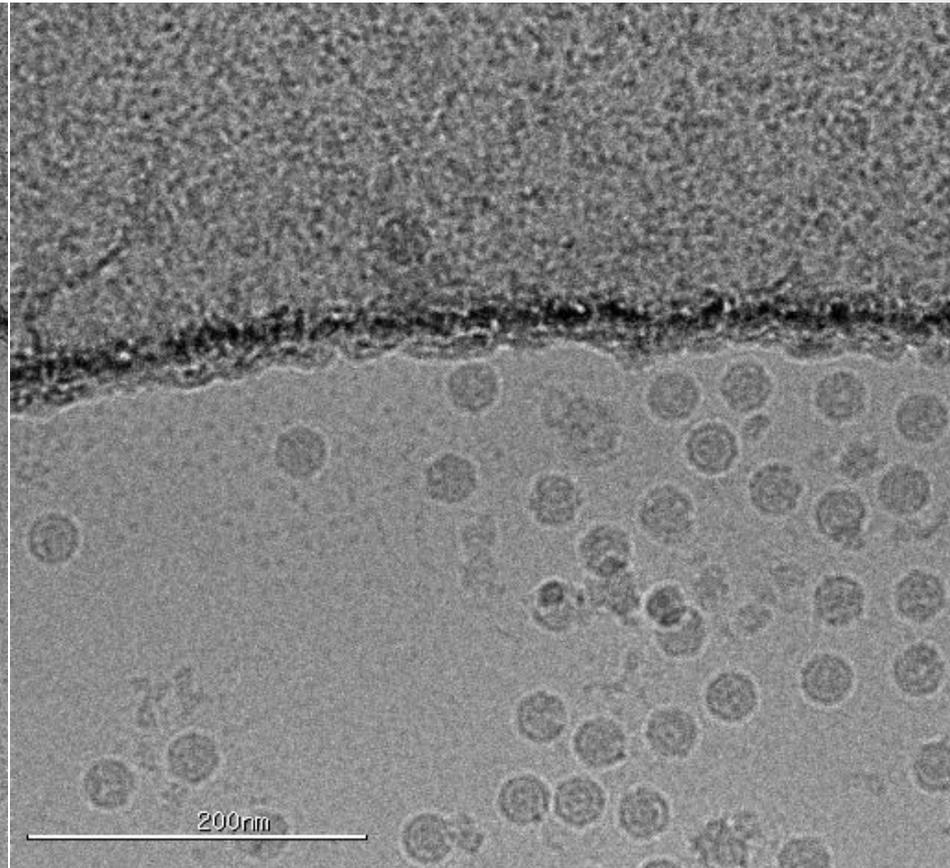
Aberration affects image formation

Intermediate and projector lenses further magnify image

## Images collected at different defocus values



**-1  $\mu\text{m}$  defocus**



**-6  $\mu\text{m}$  defocus**

# Methods for contrast and SNR improvement

Sample level: Cryo-negative staining

Microscope level: phase plate, energy filters, aperture size, defocus

Data collection level: direct detectors, automated collection

# Detection system

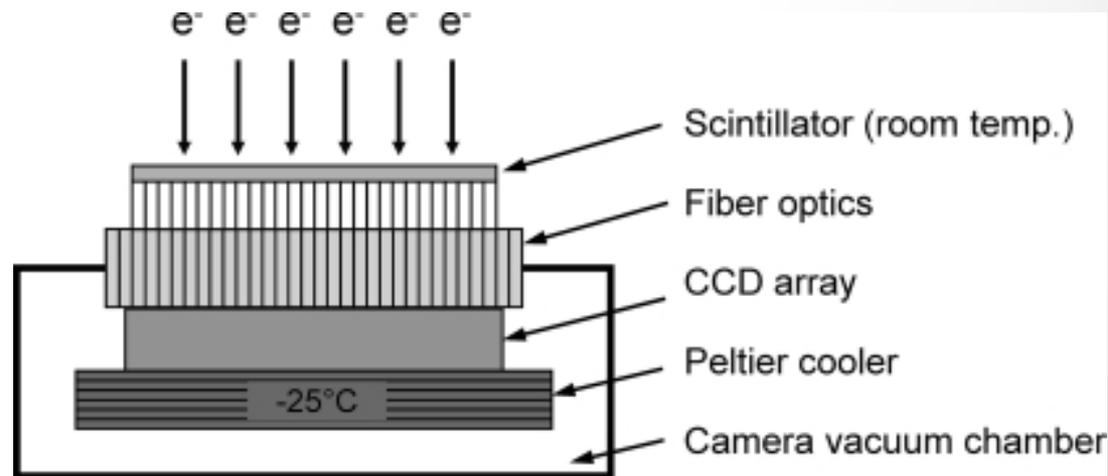
CCD camera

Incident electrons converted to photons

Fiber optics transfer image to charge coupled device sensor

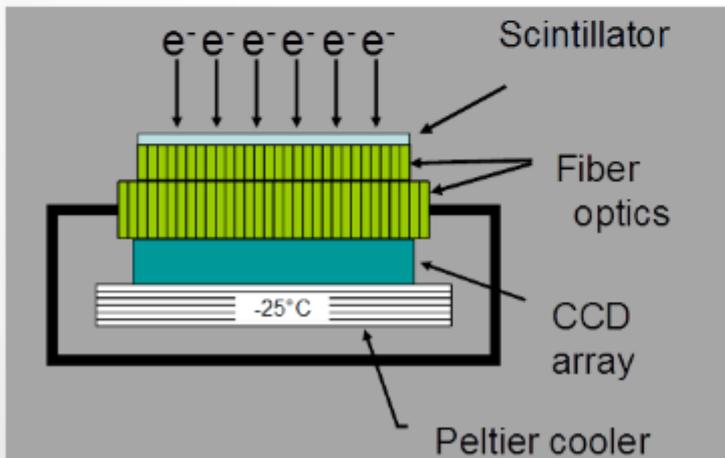
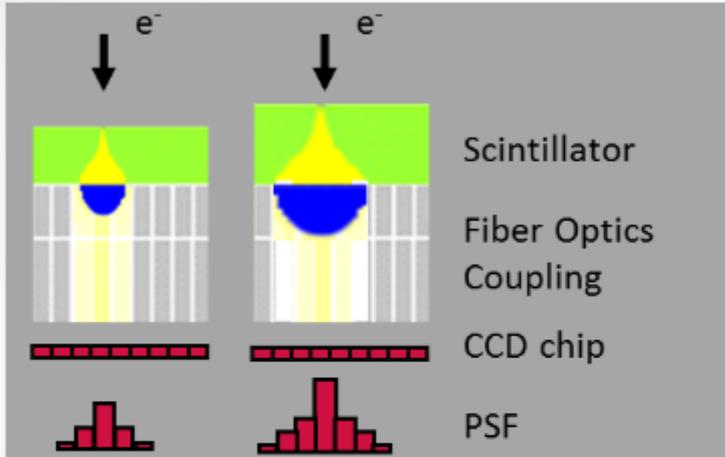
Photons generate electric charge

Charge converted to pixel for readout

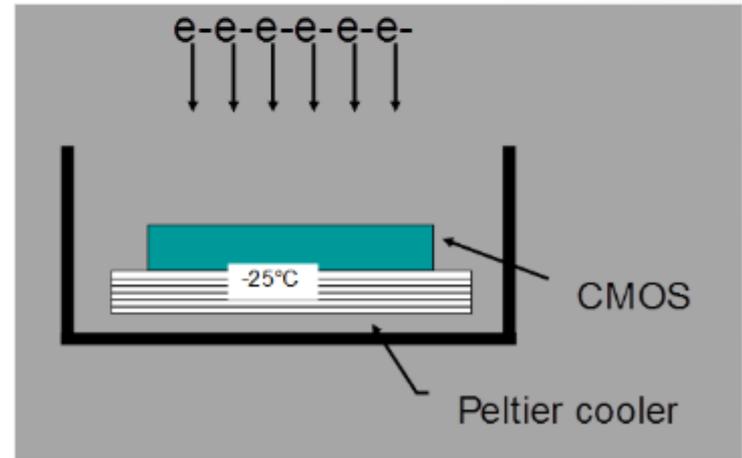
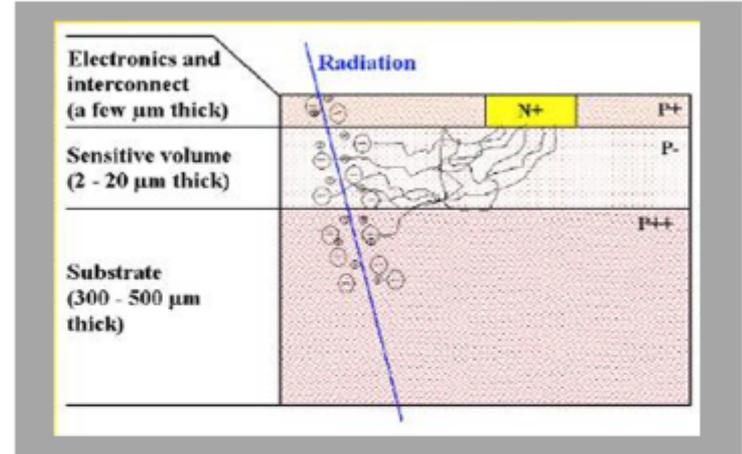


# Direct Electron Detector

**CCD:** multi stage conversion of electron energy via fiber or lens optics



**CMOS:** direct conversion of electron energy without fiber or lens optics



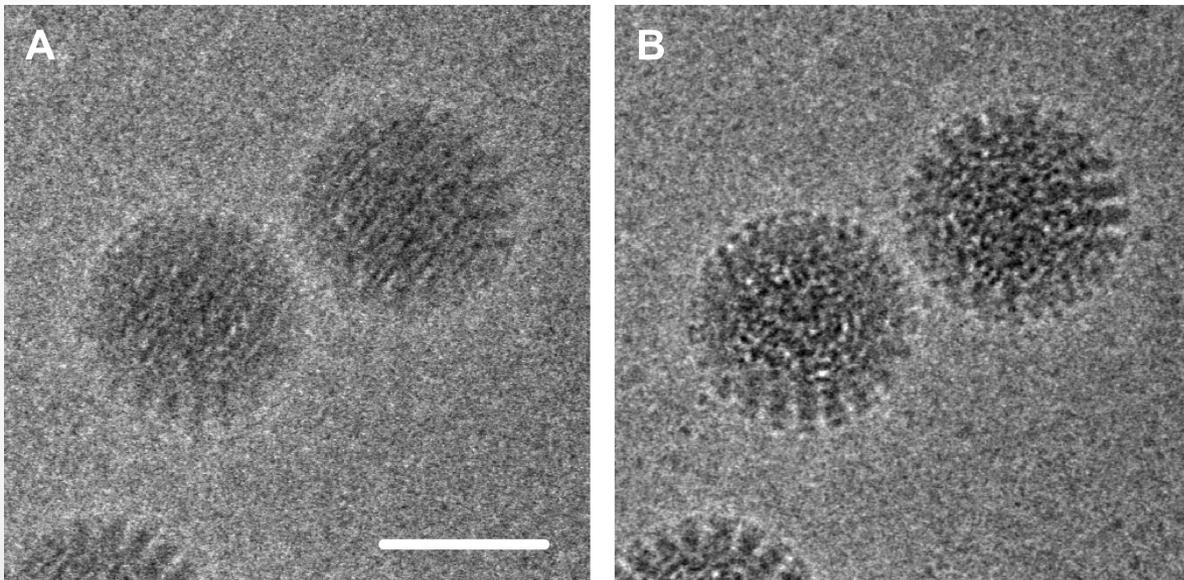
## Advantages of Direct Electron Detector

Direct counting of electrons

Reduced noise from detector

Fast frame rate, correction of beam induced movement possible

Subframe alignment



# Computer controlled data collection

Automation of repetitive operations: Searching for suitable areas for imaging  
Lens setting, stage movement  
Low dose operation  
Large dataset collection  
Basic image FTs

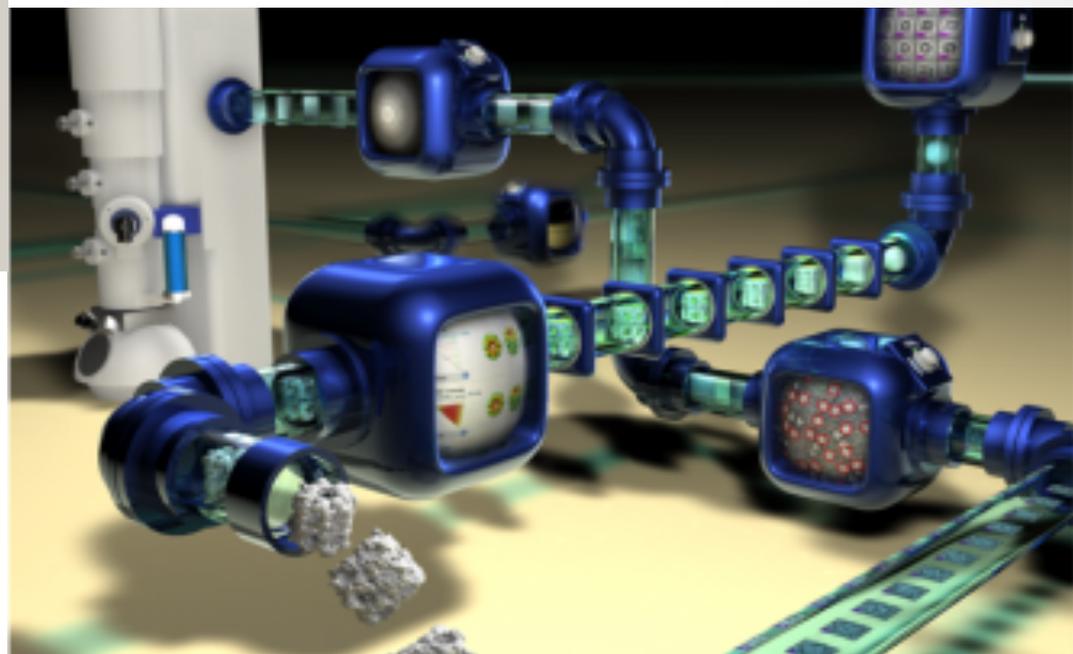
Typically, overview images collected  
cross-correlated with manually collected images  
High-mag recording after low-mag searches

# Computer controlled data collection

**Leginon Import**

Host name *	Database Name *
Leginon DB <input type="text" value="bb02frank15.cpmc.columbia.edu"/>	<input type="text" value="leginondb"/>
Project DB <input type="text" value="bb02frank15.cpmc.columbia.edu"/>	<input type="text" value="projectdb"/>
Leginon Credentials	Database Credentials *
Username <input type="text" value="namho"/>	<input type="text" value="*****"/>
Password <input type="text" value="*****"/>	<input type="text" value="*****"/>

\* Note that the Database Information will be supplied by your System Administrator



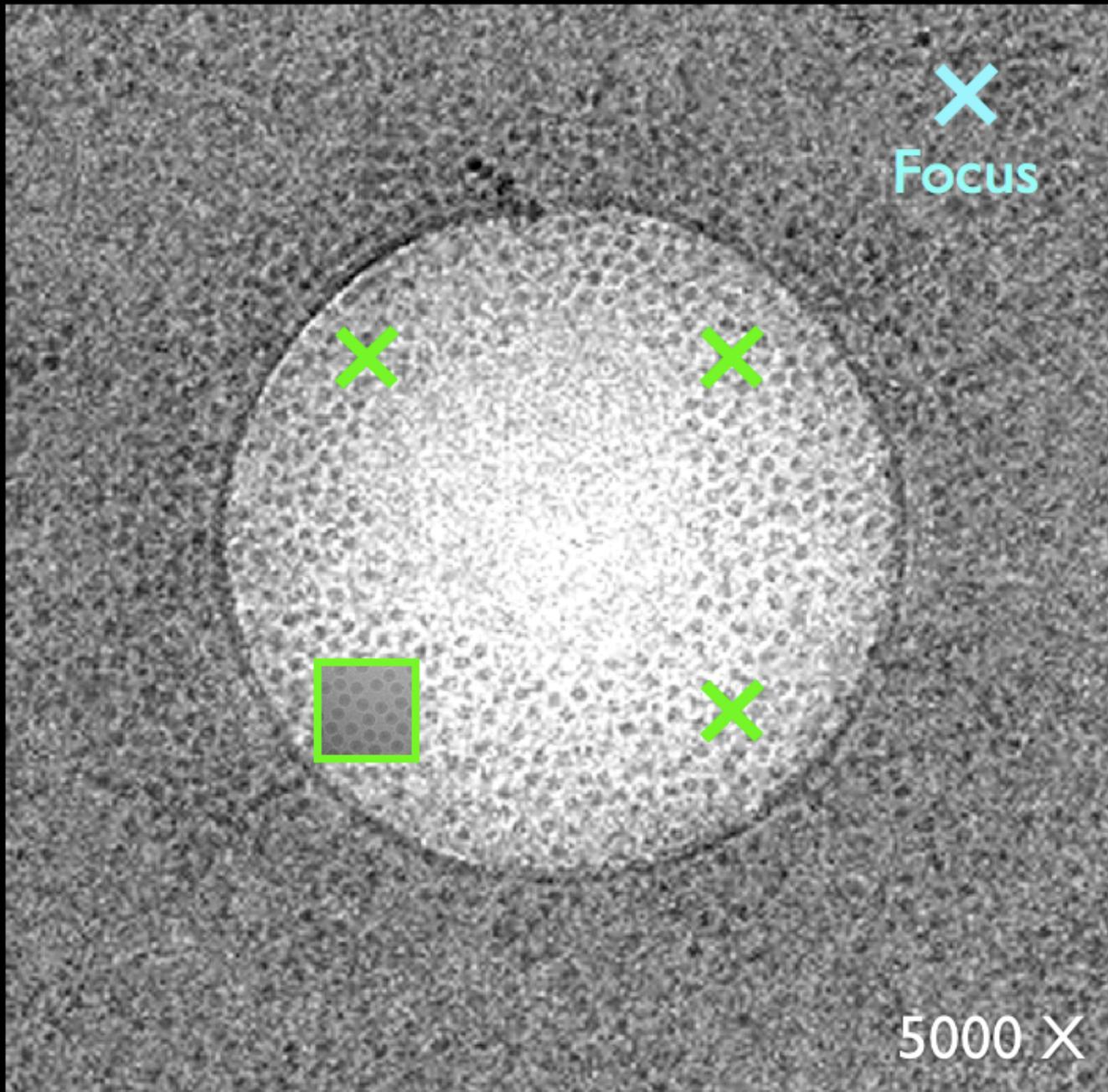
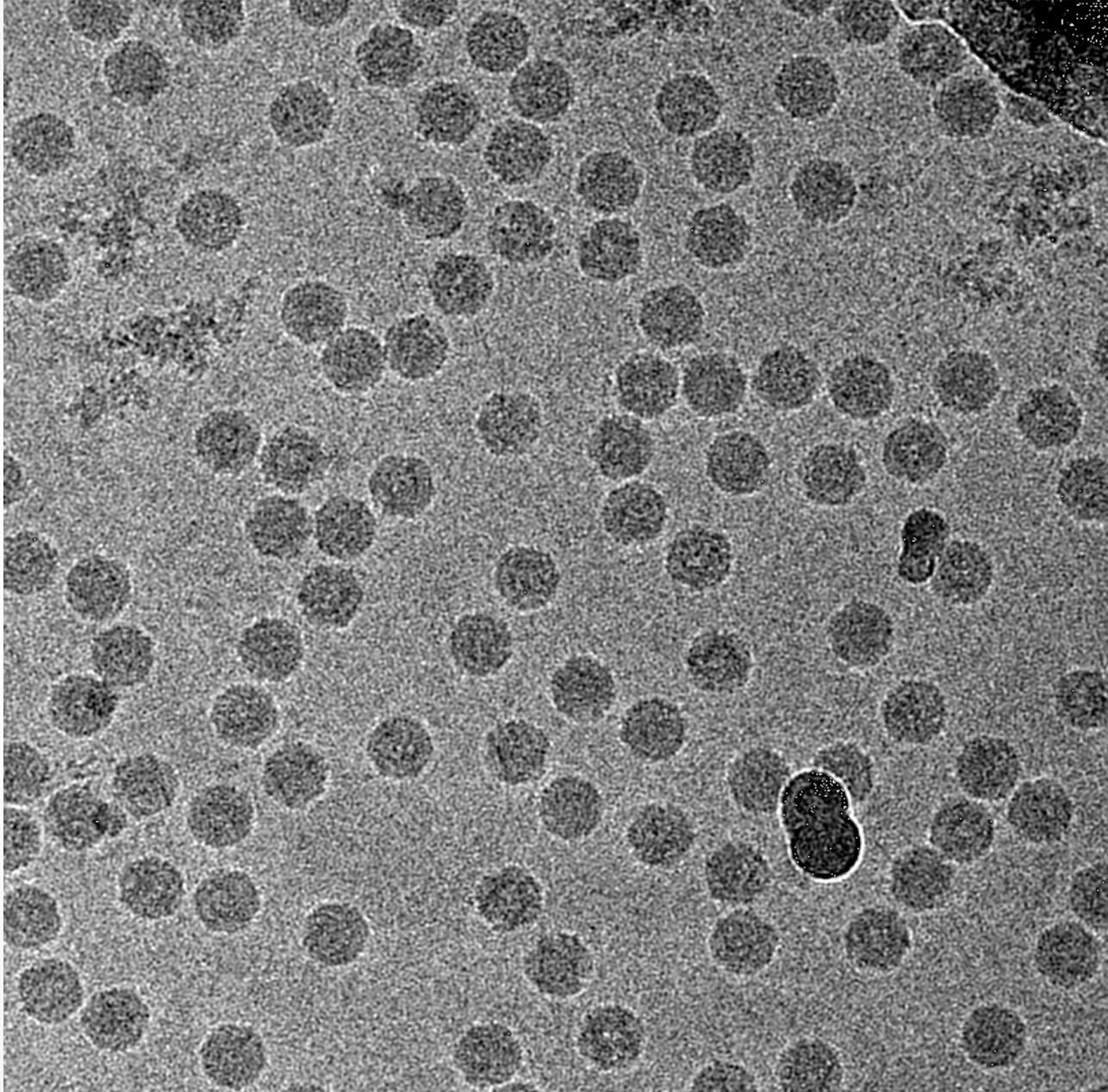
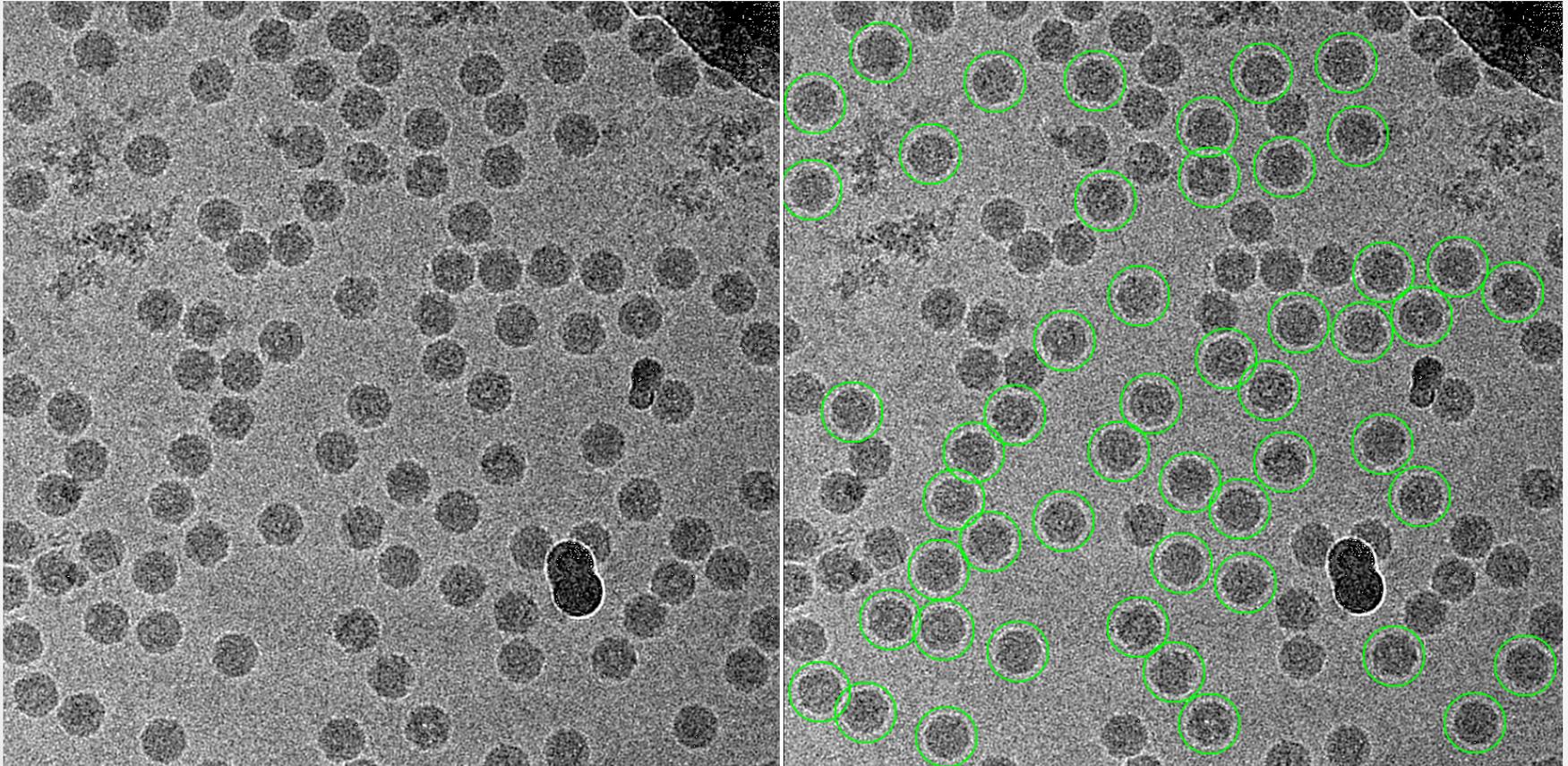


Photo Credit: Gabriel Lander, TSRI

# Particle Picking



# Particle Picking



# Particle Picking

## Template matching:

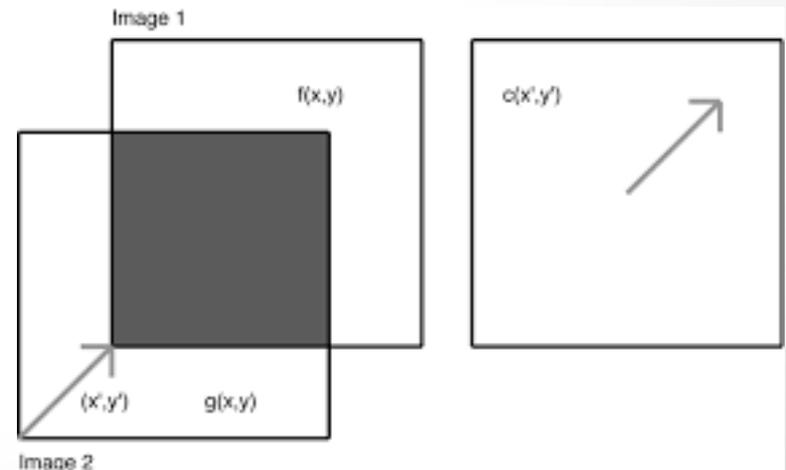
Match between image and reference image scored

Cross-correlation based methods

Sensitive to variations in spatial frequency

Multiple references required to account for different views

Rotationally averaged references/azimuthally averaged particle image



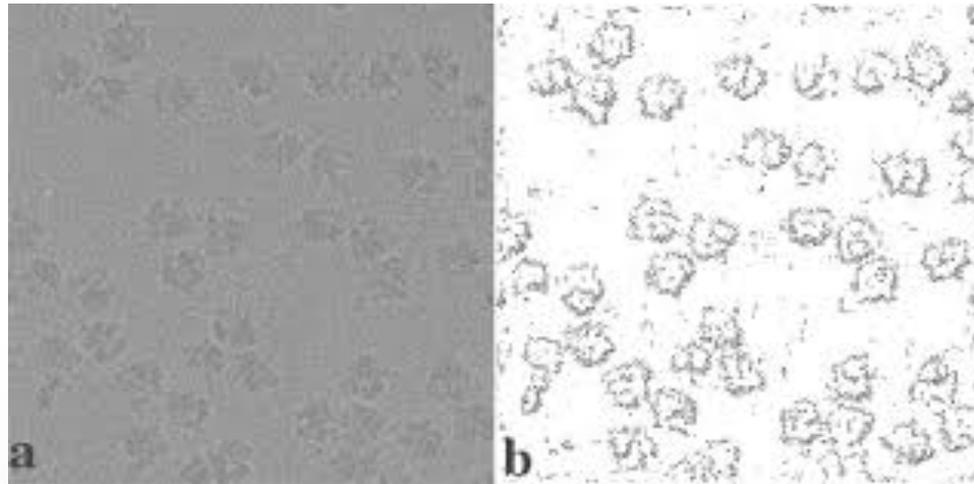
# Particle Picking

## Edge detection:

Identification of blobs in image, assignment of labels to adjacent pixels

Too close or too large “bounding boxes” rejected

Somewhat insensitive to spatial frequency



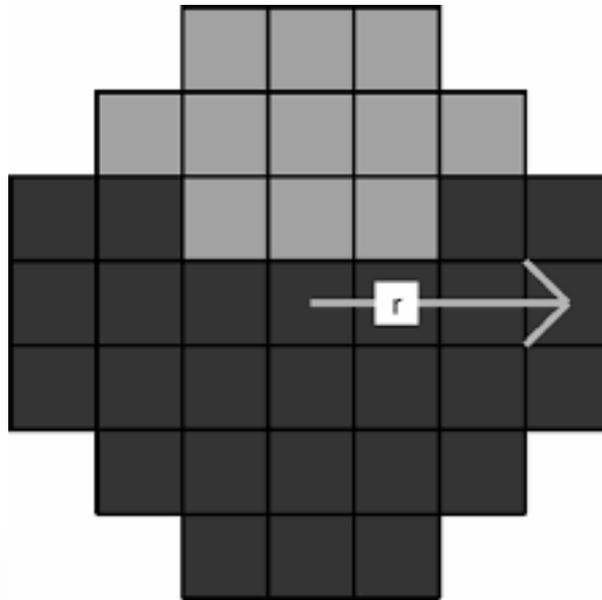
# Particle Picking

## Intensity comparisons:

Objects with uniform internal density selected

Image subjected to horizontal-vertical scan to identify clusters

Post-processing checks



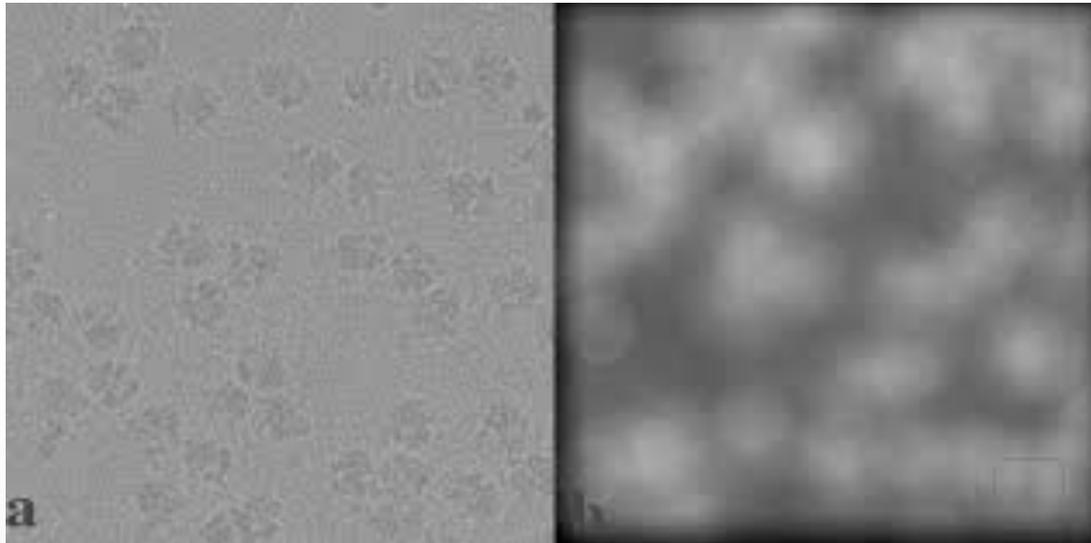
# Particle Picking

## Texture based methods:

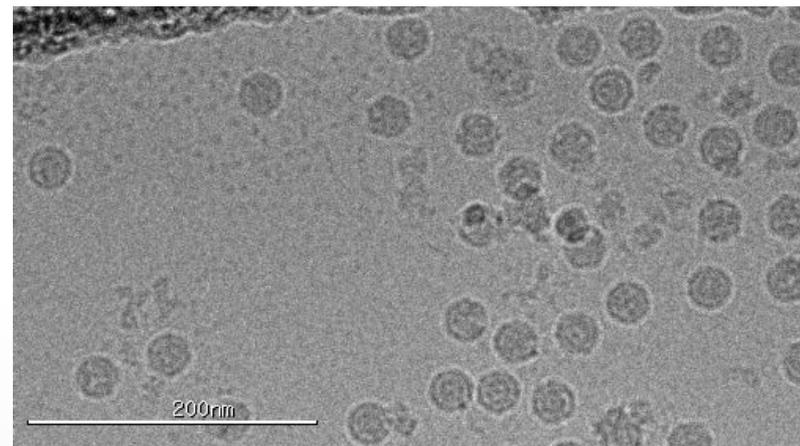
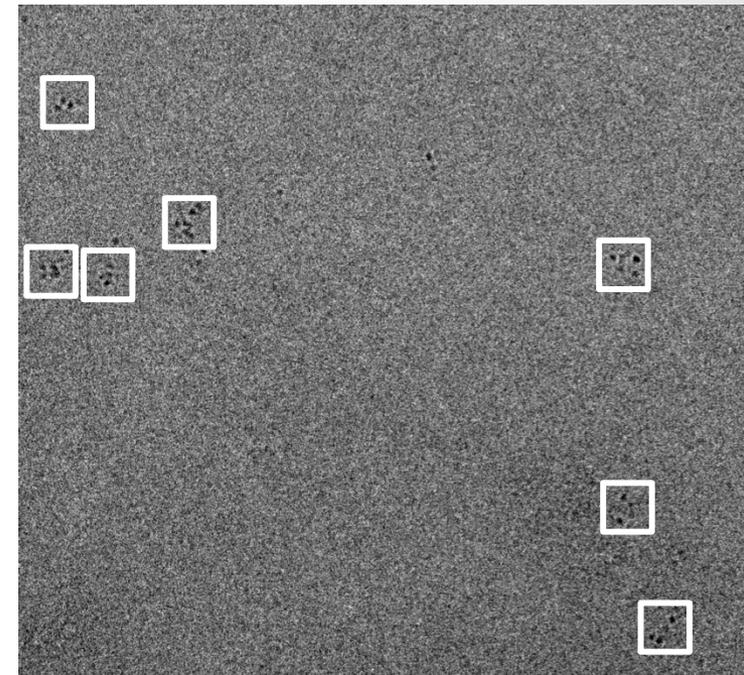
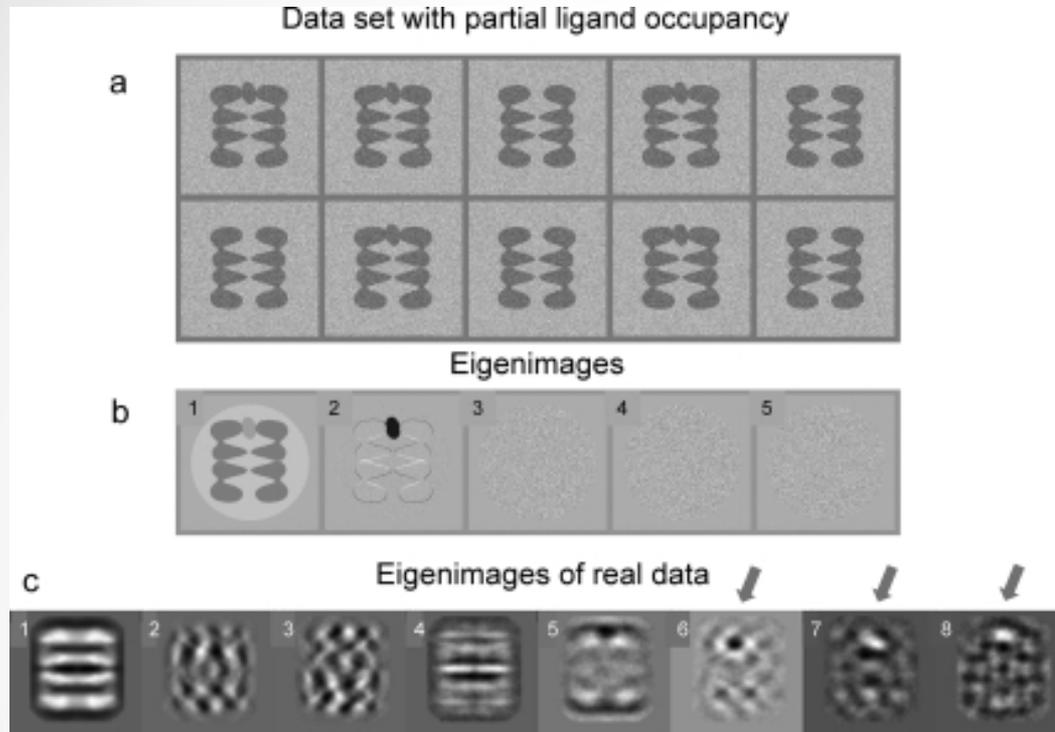
Computes local variance over small area

High values of local variance indicate presence of object

Also detects aggregates/contaminants



# Problems – heterogeneity, contaminants, background noise



# References

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